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PHENOLIC ACIDS: A POLYPHENOL COMPOUND

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INTRODUCTION

Phenolic acids have a spot with an outstandingly improved assembling of phytochemicals, phenolics that are obtain in various food assortments of plant starting in the personage eating schedule. Polyphenols are discretionary metabolites made from a sweet-smelling ring bearing no less than one hydroxyl substituent along with different inverse side get-togethers.

Phenolic compounds most by and large appearing in herbal consolidate phenol carboxylic acid acids, coumarins, flavonoids, tannins, stilbenes, lignin's as well even as lignans Phenolics are joined as initial path of shield substance constituents that opposed to living (defilements, harming) and non-living (supplement insufficiencies or surplus, cold and recognizable light) strain on plants. Similarly, phenolics add to different qualities of food assortments like bitterness, astringency, assortment, flavour, smell and strength against lipid oxidation.

A gathering of plant optional metabolites that dazzled by ton of examination occupied because of their business esteem in material, food, and wellbeing related enterprises are phenolic compounds normally alluded to as "plant phenols"

Phenolic acids have a lot of clinical benefits, for instance, relieving, antibacterial, antiproliferative, antineoplastic and antioxidative activities. Within the living body and pathogenitic assessments recommend that polyphenols strengthen by means of cell support attributes could apply clinical benefits therefore further developing steady diseases related with oxidative mischief.

The clinical benefits of dietary polyphenols are applied by a variety of routine processes, such as metal chelation, chain breaking, equilibrium of enzymatic development, free progressive scrounging, and alteration of signal transduction pathways.

Plants fostered the potential to convey a huge and numerous discretionary metabolites, that are nonessential for run of the mill new development and proliferation, yet, rather accept critical parts in plant watch frameworks, science, and normal change.

The phenolics can be artificially confusing and produced in polymeric construction, such the basic structure of a phenolic chemical is a sweet-smelling ring with at least one hydroxyl social event. PlanPhenolics from plants can be crucial to the extent that they are created, such as hydroxybenzoic acids (C6-C1) and benzoquinones (C6), either it can be intentionally confusing and occurred in the form of polymeric construction, namely (C6-C3) lignin as well as (C6-C3-C6) united tannins.

CHEMISTRY:

Two classes of phenolic acids in view of C1eC6 and C3eC6 spines which are hydroxybenzoic as well as hydroxycinnamic acids, that is universally tracked down regarding herbal substance near different range [1].

Gallic, p-hydroxybenzoic vanillic, acid syringic, and the protocatechuic acids are examples of hydroxybenzoic acids.

Hydroxycinnamic acids generally get within food varieties as well as refreshments are sinapic cinnamic acids, caffeic, p-coumaric, ferulic.

Through the activity of shikimate, pentose phosphate, phenolic molecules phenylpropanoid digestive biological pathways, are obtained through a short supply of biosynthetic precursors, like pyruvate, acetic acid derivation, Acetyl CoA, malonyl CoA, and some other amino acids [2] [3]. Phenylalanine and, to a less Tyrosine as well as its extent are twoof the main protein (amino acids) present within combination of phenolics in plants [4] [5].

Generally speaking, solvent phenolics are phenolic compounds that can be extracted into fluid or watery-natural dissolvable mixtures. These involve phenolic compounds that are present in unbound or non-formed forms, further also phenolic compounds that are formed by dissolving the carbohydrates as ester as well as ether (etherified) securities [1] [6]. Furthermore, not soluble bound phenol carboxylic molecules, mainly hydroxycinnamic acids (esterified) and converted into glucose(sugar) deposits of the polysaccharides, giving connection among the polymers of cell wall, that can be obtained from the leftover buildup following the extraction of the dissolvable phenolics [7]. They also form CeC connections and ether bonds with lignins [8]. According to certain studies, formed and insoluble bound phenolics may be administered under colonic maturation and in a variety of basic and acidic gastro-digestive conditions. They may also provide therapeutic benefits even at nearby locations, such as the gastrointestinal epithelium, and after retention [9] [11].

Phenolic acids, that contains the derivatives of hydroxybenzoic and hydroxycinnamic acids, which are essentially free and produced in grains [12].

PHENOLIC ACIDS PRESENT IN PLANTS:

Phenolic acids provide a vast number of functions in plants, which includes being the main components of the cell wall, enhancing the absorption of supplements, promoting seed germination in the face of abiotic stress, acting as indicator particles in systemic as well as local plantsdefence reactions as well as influencing nomination regarding microorganisms that benefit plant in the root zone [14] [15].

In plants, the shikimate pathway primarily uses two sweet-smelling amino acids (L-phenylalanine as well as L-tyrosine) to biosynthesize polyphenol acids (14).

Generally, three steps are given for the production regarding phenol carboxylic acid:

- Deamination
- Oxidation
- Methylation

Temporarily, the amino acids phenylalanine and tyrosine deamine produce p-coumaric and cinnamic corrosives, respectively, which are not phenolic corrosives.

Benzoic corrosive (C6-C1) is occurred when side chain of ethyl is removed by the cinnamic corrosive (C6-C3). All the phenol carboxylic acids are essentially derivatives of benzoic (hydroxylated) and also of cinnamic corrosives.

Phenolic acids compounds are present plant parts and can exist as simple glycosides, depsides, or insoluble forms as underlying cell wall components linked with xylenes, gelatin, and lignin [16].

Rosmarinic corrosive, which is an ester of two caffeic corrosive atoms, is a model of a depside, which are unusually regular particles that are ester-connected and composed of at least two units of the same or distinct phenolic corrosive.

ISOLATION AND CHARACTERIZATION OF PHENOLIC ACIDS:

The ability of phenolic acids to separate from plant tissues is what makes them unique. Generally speaking, the major phenolic intensifier due to sheer variety complicates the investigation. A number of phenolic compounds are highly susceptible to chemical changes like hydrolysis, oxidation and isomerization [17], therefore preliminary procedure will be conducted while the test handling to ensure the measurand reliability [18] [19].

Depending on the food material's nature and molecule size, the synthetic nature of phenolic compounds, the conditions and capacity time, and the presence of substances that slow down the detachment contact, test readiness can range from a simple process to one that is highly complex. [20] [21]. After

being ground to a powder with a specific molecular size, grains, vegetables, oilseeds, nuts, and dried leafy foods are defatted using a natural dissolvable such as hexane. [22, 23], [24], [25], [26], and [29]. The solvent used to extract phenolic compounds from plant yields a complex mixture of diverse dissolved compounds [21].

The location of phenolic compounds and whether they occur as free, produced, or bound compounds influence different extraction techniques. Water and open fluid natural solvents that are used to complete the strong fluid removal of free and dissolvable esters of phenolic acids. Ethyl acetic acid derivation, high temperature water, methanol, cHa)2CO, and methanol-smelling salts are among the solvents used. Soxhlet extraction [30], vortexing followed by centrifugation [18] [19], ultrasound-assisted extraction [22], mechanical blending by homogenization [31] [32], continuous turning extraction, compressed fluid extraction [33], and microwave-assisted extraction [34] [35] are some of the strategies that alter the announced extraction times.

Ultrasound extraction method is considered as an alternative for the traditional methods when separating the phenolic compounds because it provides high efficiency while needing less energy and water [36]. Ultrasound waves increase the intensity of the extraction process by degrading the cell wall and reducing molecular size, they also help in the transfer of cellular components into the solvent by dissolving the air pockets formed during the cavitation process [38].

Depending on the extraction techniques used, different phenolic chemicals may be supplied [23]. Filtration or centrifugation are used to separate the remaining insoluble accumulation. The percentage of dissolvable to-test affects the extraction of phenolic chemicals [21]. Enzymatic, corrosive, or soluble base treatments are used to deliver bound phenolic compounds [4]. Numerous experiments have demonstrated the use of basic hydrolysis, that can be either lengthy hydrolysis when processing time exceeds 16 hours or fast hydrolysis (from 1 to 4-6 hours) [30] [33]. Basic hydrolysis reactions have been allowed to occur at ambient temperature by a variety of agents.

In order to limit the isomerization of combinations, including phenolic acids, certain tests were carried out in secrecy and with dormant air, such as argon or nitrogen [40]. Not many investigations

have involved corrosive hydrolysis for the recuperation of bound phenolics [33]. Enzymatic responses are further utilized to deliver unfree phenolic acids. Proteins namely cellulases, amylases, and pectinases are used to break down starch bonds [42] [53] [54]. The hydroxyl bunches on the fragrant rings and an acetal or hemiacetal bond across the starch components are broken during enzymatic hydrolysis, releasing the phenolic acids. Following the extraction of phenolic substances, spectrophotometric analysis (SA), GC (gas chromatography) or HPLC may be used to confirm their content. Phenolic substances are often separated and evaluated using GC and HPLC techniques [21].

Phenolic compound analysis is a common application of HPLC. Table 4 provides models for the different locators and systems used to examine food ingredients. To identify the primary characteristics of phenolic compounds, HPLC is used in conjunction with mass spectrometry, 1 H and 13C atomic attractive reverberation spectrometry, or infrared spectroscopy.

RECOVERY OF PHENOLIC ACIDS FROM BOTANICAL SOURCES:

It is possible to extract phenolic acids from lively, frozen, or stored plant material. Although, before phenolic corrosive extraction, herbal biological mass is usually dried using air or freeze then further ground in a uniform particle (powder). Since proteins in newly formed cells may potentially contaminate phenolic acids, it is anticipated that the extraction from dry biomass will also be extracted from new biomass. The phenolic content in plant biomass has been observed to be preserved in higher amounts when compared to air-drying, freeze-drying [56].

The ideal degree of concentrate immaculateness, the synthetic complication of plant biological mass as well as substance qualities of the perfect phenolic corrosive (such as extremity) are some of the factors that are specifically tailored to the extraction cycle [56] [57].

Generally speaking, the cycle begins with a crude phenolic extraction by solution of 20%-half ethyl alcohol, methyl alcohol or the mixture of solvents (water). To release non-soluble phenolic acids to their dissolvable free structures, the unrefined concentrate is subsequently subjected to base hydrolysis on a regular basis using 1-4M sodium hydroxide. Although enzymatic hydrolysis by enzymes regarding a substitute to hydrolysis(base), typically most costly as well as not even drilled. Nonpolar solvents such as hexane can be used in fluid fractionation to remove lipids from unrefined concentrations.

If phenol carboxylic acids are used in their purest form, they are refined in the rough concentrates using further fluid or strong stage extraction (SPE) procedures. SPE is typically used because it is quick, sensitive, and an economical technique that uses sections with different types of sorbents. In order to obtain and separate phenolic acids depending upon their pKa and science, C18 sections are often commonly used for partition of phenol carboxylic acids. Common work technique consists of altering pH of unrefined concentrate or pH and extremity of carrier.

The following sections will demonstrate both emerging solutions that utilizes microwave or compact form of solution as removal specialists, as well as conventional approaches like SE and UAE. In emerging extraction strategies like SFE control of trial state been utilized for quickly isolate phenol carboxylic acids depending upon physical and chemical uses from various constituents in the plant biological mass, whereas majorly regarding techniques will need extra advancements as SPE to decontaminate phenol carboxylic acids by rough concentrates.

ULTRASOUND EXTRACTION:

The kilohertz range of acoustic waves that form through the dissolvable creating cavitation bubbles is included in the UAE [58] [59]. A shockwave-induced malfeasance to lay out the cell wall works on the mass trade of phenolic heightens across cell layers into plan just as vapor-filled cavities implode at exterior layer of plant test structure [59].

In any case, ultrasonic waves have been given the attention they deserve in order to corrupt a few phenolic acids and create very reactive hydroxyl progressives within the gas bubbles ([59]). After extraction, the concentrate is separated from the plant growth using filtration. For the plant test under investigation, the UAE extraction show is frequently enhanced in terms of temperature, dissolvable, and dissolvable to biomass extent.

Supercritical fluid extraction:

It is a most clear process that uses compressed liquids as solvents. The formation of a fluid (supercritical) starts with a solvent that is subjected for strain and temperature which is more than its critical point, making liquid as well as gas phases identical state one another. The physicochemical characteristics of both fluid as well as gas form are displayed by supercritical fluids [60] [61] [62]. They have a thickness of 0.3 to 0.8 g/cm3, a consistency of 104 to 103 g/scm, as well as scattering quantity which is most similar by the fluid as well as gas (103 to 104 cm2/s). Because of these characteristics (poor consistency, somewhat high diffusivity), the ideal dissolvability can be affected by altering the strain and temperature, which in turn alters the thickness of the supercritical fluid.

Supercritical fluids have higher diffusivity rates and less consistency than regular solvents, yet they still retain solvating qualities. Overall, supercritical fluids can quickly diffuse through materials and have preferred transport qualities over typical regular solvents, which further enhances the capacity to extract the best compounds and produce the highest yield. Supercritical (CO-2) are more often utilized supercritical liquid by its mild fundamental temperature-31.3°C as well as pressure-72.9atm. Since CO-2 which is gas at room temperature, it is definitely possible to separate or destroy it in order to get a free concentrate that is soluble.

In any event, utilizing supercritical CO 2 has the disadvantage of having a low limit and occasionally being less effective. (60) (62).

SOXHLET EXTRACTION:

The most popular technique for getting rid of phenolics and other plant auxiliary metabolites is SE. In this phenol is removed by herbal biological mass through dissolving in the dissolvable component. A tiny thimble and a Soxhlet measuring unit (often formed of cellulosic compound) to hold model, as well as a percolator a flowing water shower linked to the Soxhlet assembly as well as a warming sign are necessary for the straightforward SE methodology.

Using SE has the advantage of avoiding the requirement for a filtration push to recover the dissolvable with removed phenolics because plant development is contained in the thimble rather than being directly mixed in with the dissolvable. Over the course of two to twelve hours, the SE connection is routinely extended, typically using a 40% to 60% ethanol or methanol in water plan that dissolves at reflux.

However, other solvents as CH3)2CO, acetonitrile, or pure water have also been used as soluble. The most typical answer is ethyl or methyl alcohol in water. Although it is clear that the SE show must be adjusted to take into consideration the plant biomass used, a good starting point may be to perform SE using a half-ethanol in water game plan and an extraction period longer than four hours at temperatures lower than 100°C.

impact of phenolic acids on human health:

Phenol carboxylic acids are substances that affect natural structures. They prevent illnesses and are linked with biological actions that can improve human health, they stand out in the pharmacological and restorative exploration domains. Oxidative pressure in cells causes aging and a variety of Chronic disorder like type 2 diabetes, heart disorders, as well as disease. Oxidative pressure is brought on by excessive amounts of responsive oxygen species (ROS and RON), such as hydrogen peroxide and peroxynitrite and free radicals in cell., which in turn causes DNA, lipids, molecules, and proteins to oxidize

The term "cell reinforcements" refers to particles that have the ability to both reduce and eliminate the material reactivity of ROS, RON, and free revolutionaries. Normal cell reinforcements, phenolic acids have redox characteristics that enable them to function as singlet oxygen quenchers, hydrogen benefactors, and decreasing specialists [63]. Phenolic acids' synthetic design is responsible for their cell reinforcement movement: (1) OH groups which can give an electron or a [H} molecule to the radicals; (2) a fragrant framework can cause an unpaired electron to become decentralized [64]. Protocatechuic corrosive, p-coumaric, ferulic acids, caffeic, gallic, cinnamic, and p-hydroxybenzoic as well as methylated, sulfated, and concentrated forms of the parent phenol carboxylic acids, were evaluated for their ability to reinforce cells [65].

When compared to the parental phenolic acids, altered phenolic corrosive metabolites often exhibited less cell reinforcement activity. Additionally, these evaluations illustrated the significance of location as well as quantity of OH bunches in original structure [63]. The general conclusions drawn from these analyses are that 1) monophenols are weaker extreme foragers than polyhydroxyphenols, and the cell reinforcement action increases as the number of hydroxyl bunches in the parental phenolic corrosive design increases. For example, p-coumaric corrosive, a monophenol, has a lower cell reinforcement action than protocatechuic and caffeic corrosives. Similarly, gallic corrosive, a triphenol, has a stronger effect on cell reinforcement than caffeic and protocatechuic corrosives.

The ability of the phenolic corrosive to prevent cancer is dependent on the location of the OH group with respect to the carboxyl group; OH, in the meta position are more effective than those in the ortho and para locations. Functional groups in both ortho and para positions are impacted by the carboxylic bunch on the benzene ring's ability to take up electrons. Presence by a few methoxy substitutions at locations ortho to the hydroxyl bunch increases the cell reinforcement movement of monophenols. For example, ferulic corrosive is far more effective than p-coumaric corrosive, yet synaptic corrosive is more persuasive. Thus, syringic acid is found to be more effective than p-hydroxybenzoic acid and vanillic acid. 4) Compared to hydroxybenzoic corrosive subordinates, hydroxycinnamic corrosive subsidiaries frequently exhibit higher cancer prevention agent exercises

Hydroxycinnamic corrosive subsidiaries have a more grounded H-giving capacity and revolutionary adjustment due to the presence of the carboxylate bunch in the middle between the phenyl ring and the ring. Despite their ability to prevent cancer, phenolic acids can also regulate cell flagging cycles

during irritation [23] and have been shown to inhibit the overproduction of fiery arbiters, which can cause a variety of cell damage. According to one review, proinflammatory cytokines can be reduced by chlorogenic corrosive [65].

Another evaluation found that when given at 10 mg/kg body weight, syringic corrosive might reduce serum cytokine levels and smother hepatic aggravation in mice [54]. Because rosmarinic corrosive inhibits lipoxygenase and cyclooxygenases, it reduces the fiery reaction in chronic illnesses [64] and has a protective effect against organ damage [66]. Additional applications for health include cancer hiding and the therapy of malignant development. In the case of mammary organ, liver cells, such as ellagic, protocatechuic, caffeic, and gallic corrosives; cervix adenocarcinomas; lymphoblastic leukemia; and cervical carcinomahave demonstrated cellular toxicity as well as antineoplastic actions [43] [62].

The antidiabetic characteristics of phenolic acids have also been explained; for instance, gallic and protocatechuic acids shown a fixation subordinate hindrance of important catalysts linked to type 2 diabetes [59]. Another analysis revealed that in diabetic animals, syringic corrosive increased insulin levels and the translocation of glucose metabolic proteins while decreasing plasma glucose levels [45]. By preventing bacteria nucleic corrosive amalgamation, enzymatic movement, cytoplasmic layer capacity, and energy digestion, phenolic acids also function as antimicrobial specialists [65]. It has been determined that p-coumaric, protocatechuic, and caffeic acids show strong antibacterial action against bacteria that are Gram-positive and Gramnegative. [65] and limit the growth of Staphylococcus aureus EP167 and pathogenic Escherichia coli O157:H7 [34].

While coumaric(P) corrosive has shown to pair the genomic of bacterial DNA, impairing cell's purpose as well as ultimately causing bactericidal action [17], caffeic corrosive's antibacterial effect has been linked to its capability which pauses or stops the bacterial RNA polymerase catalyst [19]. Phenol carboxylic acids are also used in preventing aging, shielding the skin from ultraviolet (UV) light, ripeness therapy, and hepatotoxicity prevention. Ferulic acid, for example, shows a strong UV absorption capacity, which makes it a good skin defence specialist [65], approval as a flavour enhancers in some countries to stop the lipid peroxidation [11], longer half-life in the blood compared to other cell reinforcements, such as L-ascorbic acid [18], protection against coronary heart disease, lowering cholesterol, and increasing sperm rationality [12].

IN VITRO AND IN VIVO METHODS OF ACTION:

Three basic systems—hydrogen iota move (Cap), electro move proton move (ETPT), and successive proton misfortune electron move (SPLET)—are the reason behind defensive exercises involving phenolic chemicals [22] [63] [55]. Phenolic acids generate phenolic revolutionaries during the free extremist search period, which could be counterbalanced by creating the intra-subatomic hydrogen bonds, expanding delocalization, and forming electrons upgraded through reverberation adjustment [58].

When in their free form, phenolic acids are maintained, and when bonded, they can be transported throughout the body. In its free form, ferulic acid can be taken from the abdomen of a mouse and is further converted into ferulic acid in the liver [43]. Furthermore, it has been observed that human subjects release Ferulic acid in its glucuronide and sulfate forms, which suggests that the body retains and digests it [18]. An ester links phenolic acids that are bonded together, particularly hydroxycinnamates like p-coumaric and ferulic acids in cereal grains., to the Plant cells contain arabinoxylans that are aromatic walls

Ferulic and di-ferulic acids from grain wheat can be delivered by GIT esterase enzyme from the digestive mucosa and microbiota.

CONCLUSION:

Bioactive substances called phenolic acids affect by plant's process, environment, as well as growth. The majority of phenol carboxylic acids are found by bonded structures as framed esters, insoluble hidden fragments of cell wall by gelatin, xylenes, as well as lignin, or like direct glycosides. Generally speaking, the idea is to identify the phenolic acids that are relevant regarding a particular use as well as identification.

These extraction methods, expertise level, and personal preference will all affect the guarantee of a phenolic destructive extraction system. Perhaps the best option for the model is to begin with standard procedures if the assessment is exploratory and time is not a constraint. In any case, the use of new techniques, such as SFE and Pro, is a popular choice if the extraction's goal is to regain a large amount of phenolic acids in the small amount of time and low cost.

Phenolic acids are widely examined for their use in prosperity because of their anticancer, soothing, cell-supporting qualities, among other things. The significance of this topic is implied by the enormous number of publications that have been published on phenolic destructive evaluation and their extraction from plants throughout the past ten years.

REFERENCES:

- 1. Shahidi, F., & Yeo, J. (2016). Insoluble-bound phenolics in food. Molecules, 21(9), 1216.
- 2. Robards, K., Prenzler, P. D., Tucker, G., Swatsitang, P., & Glover, W. (1999). Phenolic compounds and their role in oxidative processes in fruits. Food chemistry, 66(4), 401-436.
- 3. Randhir, R., Lin, Y. T., Shetty, K., & Lin, Y. T. (2004). Phenolics, their antioxidant and antimicrobial activity in dark germinated fenugreek sprouts in response to peptide and phytochemical elicitors. Asia Pacific journal of clinical nutrition, 13(3).
- 4. Shahidi, F. (2002). Phytochemicals in oilseeds. In Phytochemicals in nutrition and health (pp. 155-172). CRC Press.
- 5. Herrmann, K., & Nagel, C. W. (1989). Occurrence and content of hydroxycinnamic and hydroxybenzoic acid compounds in foods. Critical reviews in food science & nutrition, 28(4), 315-347.
- 6. Shahidi, F., & Chandrasekara, A. (2015). The use of antioxidants in the preservation of cereals and low-moisture foods. In Handbook of antioxidants for food preservation (pp. 413-432). Woodhead Publishing.
- 7. Ishii, T., & Hiroi, T. (1990). Linkage of phenolic acids to cell-wall polysaccharides of bamboo shoot. Carbohydrate research, 206(2), 297-310.

- 8. Grabber, J. H., Ralph, J., & Hatfield, R. D. (2000). Cross-linking of maize walls by ferulate dimerization and incorporation into lignin. Journal of agricultural and food chemistry, 48(12), 6106-6113.
- 9. Andreasen, M. F., Landbo, A. K., Christensen, L. P., Hansen, Å., & Meyer, A. S. (2001). Antioxidant effects of phenolic rye (Secale cereale L.) extracts, monomeric hydroxycinnamates, and ferulic acid dehydrodimers on human low-density lipoproteins. Journal of Agricultural and Food Chemistry, 49(8), 4090-4096.
- 10. Andreasen, M. F., Kroon, P. A., Williamson, G., & Garcia-Conesa, M. T. (2001). Intestinal release and uptake of phenolic antioxidant diferulic acids. Free radical biology and medicine, 31(3), 304-314.
- 11. Chandrasekara, A., & Shahidi, F. (2012). Bioaccessibility and antioxidant potential of millet grain phenolics as affected by simulated in vitro digestion and microbial fermentation. Journal of functional foods, 4(1), 226-237.
- 12. Chandrasekara, A., & Shahidi, F. (2011). Determination of antioxidant activity in free and hydrolyzed fractions of millet grains and characterization of their phenolic profiles by HPLC-DAD-ESI-MSn. Journal of Functional Foods, 3(3), 144-158.
- 13. Chandrasekara, A., & Shahidi, F. (2011). Antiproliferative potential and DNA scission inhibitory activity of phenolics from whole millet grains. Journal of Functional Foods, 3(3), 159-170.
- 14. Cheynier, V., Comte, G., Davies, K. M., Lattanzio, V., & Martens, S. (2013). Plant phenolics: recent advances on their biosynthesis, genetics, and ecophysiology. Plant physiology and biochemistry, 72, 1-20.
- 15. Rivas-San Vicente, M., & Plasencia, J. (2011). Salicylic acid beyond defence: its role in plant growth and development. Journal of experimental botany, 62(10), 3321-3338.
- 16. Gorshkova, T. A., Salnikov, V. V., Pogodina, N. M., Chemikosova, S. B., Yablokova, E. V., Ulanov, A. V., ... & Lozovaya, V. V. (2000). Composition and distribution of cell wall phenolic compounds in flax (Linum usitatissimum L.) stem tissues. Annals of botany, 85(4), 477-486.
- 17. Jeandet, P., Breuil, A. C., Adrian, M., Weston, L. A., Debord, S., Meunier, P., ... & Bessis, R. (1997). HPLC analysis of grapevine phytoalexins coupling photodiode array detection and fluorometry. Analytical Chemistry, 69(24), 5172-5177.
- 18. Montedoro, G., Servili, M., Baldioli, M., & Miniati, E. (1992). Simple and hydrolyzable phenolic compounds in virgin olive oil. 1. Their extraction, separation, and quantitative and semiquantitative evaluation by HPLC. Journal of agricultural and food chemistry, 40(9), 1571-1576.
- 19. Montedoro, G., Servili, M., Baldioli, M., & Miniati, E. (1992). Simple and hydrolyzable phenolic compounds in virgin olive oil. 2. Initial characterization of the hydrolyzable fraction. Journal of Agricultural and Food Chemistry, 40(9), 1577-1580.
- 20. Naczk, M., & Shahidi, F. (1989). The effect of methanol-ammonia-water treatment on the content of phenolic acids of canola. Food Chemistry, 31(2), 159-164.
- 21. Naczk, M., & Shahidi, F. (2006). Phenolics in cereals, fruits and vegetables: Occurrence, extraction and analysis. Journal of pharmaceutical and biomedical analysis, 41(5), 1523-1542.
- 22. Chandrasekara, N., & Shahidi, F. (2011). Effect of roasting on phenolic content and antioxidant activities of whole cashew nuts, kernels, and testa. Journal of Agricultural and Food Chemistry, 59(9), 5006-5014.
- 23. Chandrasekara, A., Rasek, O. A., John, J. A., Chandrasekara, N., & Shahidi, F. (2016). Solvent and extraction conditions control the assayable phenolic content and antioxidant activities of seeds of black beans, canola and millet. Journal of the American Oil Chemists' Society, 93, 275-283.
- 24. Madhujith, T., & Shahidi, F. (2006). Optimization of the extraction of antioxidative constituents of six barley cultivars and their antioxidant properties. Journal of Agricultural and Food Chemistry, 54(21), 8048-8057.
- 25. Madhujith, T., Izydorczyk, M., & Shahidi, F. (2006). Antioxidant properties of pearled barley fractions. Journal of agricultural and food chemistry, 54(9), 3283-3289.
- 26. Madhujith, T., & Shahidi, F. (2007). Antioxidative and antiproliferative properties of selected barley (Hordeum vulgarae L.) cultivars and their potential for inhibition of low-density lipoprotein (LDL) cholesteroloxidation. Journal of Agricultural and Food Chemistry, 55(13), 5018-5024.
- 27. Liyana-Pathirana, C. M., & Shahidi, F. (2005). Antioxidant activity of commercial soft and hard wheat (Triticum aestivum L.) as affected by gastric pH conditions. Journal of agricultural and food chemistry, 53(7), 2433-2440.
- 28. Liyana-Pathirana, C., Dexter, J., & Shahidi, F. (2006). Antioxidant properties of wheat as affected by pearling. Journal of agricultural and food chemistry, 54(17), 6177-6184.
- 29. Liyana-Pathirana, C. M., & Shahidi, F. (2006). Importance of insoluble-bound phenolics to antioxidant properties of wheat. Journal of agricultural and food chemistry, 54(4), 1256-1264.
- 30. Sun, R. C., Sun, X. F., & Zhang, S. H. (2001). Quantitative determination of hydroxycinnamic acids in wheat, rice, rye, and barley straws, maize stems, oil palm frond fiber, and fast-growing poplar wood. Journal of Agricultural and Food Chemistry, 49(11), 5122-5129.
- 31. Krygier, K., Sosulski, F., & Hogge, L. (1982). Free, esterified, and insoluble-bound phenolic acids. 1. Extraction and purification procedure. Journal of Agricultural and Food Chemistry, 30(2), 330-334.
- 32. Krygier, K., Sosulski, F., & Hogge, L. (1982). Free, esterified, and insoluble-bound phenolic acids. 2. Composition of phenolic acids in rapeseed flour and hulls. Journal of agricultural and food chemistry, 30(2), 334-336.
- 33. Bonoli, M., Marconi, E., & Caboni, M. F. (2004). Free and bound phenolic compounds in barley (Hordeum vulgare L.) flours: Evaluation of the extraction capability of different solvent mixtures and pressurized liquid methods by micellar electrokinetic chromatography and spectrophotometry. Journal of Chromatography A, 1057(1-2), 1-12.
- 34. Beejmohun, V., Fliniaux, O., Grand, É., Lamblin, F., Bensaddek, L., Christen, P., ... & Mesnard, F. (2007). Microwave-assisted extraction of the main phenolic compounds in flaxseed. Phytochemical Analysis: An International Journal of Plant Chemical and Biochemical Techniques, 18(4), 275-282.
- 35. Pomponio, R., Gotti, R., Hudaib, M., & Cavrini, V. (2002). Analysis of phenolic acids by micellar electrokinetic chromatography: application to Echinacea purpurea plant extracts. Journal of Chromatography A, 945(1-2), 239-247.
- 36. Wang, J., Sun, B., Cao, Y., Tian, Y., & Li, X. (2008). Optimisation of ultrasound-assisted extraction of phenolic compounds from wheat bran. Food chemistry, 106(2), 804-810.

- 37. Wang Jing, W. J., Sun BaoGuo, S. B., Cao YanPing, C. Y., Tian Yuan, T. Y., & Li XueHong, L. X. (2008). Optimisation of ultrasound-assisted extraction of phenolic compounds from wheat bran.
- 38. Paniwnyk, L., Beaufoy, E., Lorimer, J. P., & Mason, T. J. (2001). The extraction of rutin from flower buds of Sophora japonica. Ultrasonics sonochemistry, 8(3), 299-301.
- 39. Paniwnyk, L., Beaufoy, E., Lorimer, J. P., & Mason, T. J. (2001). The extraction of rutin from flower buds of Sophora japonica. Ultrasonics sonochemistry, 8(3), 299-301.
- 40. Dabrowski, K. J., & Sosulski, F. W. (1984). Composition of free and hydrolyzable phenolic acids in defatted flours of ten oilseeds. Journal of agricultural and food chemistry, 32(1), 128-130.
- 41. Yan, J. J., Cho, J. Y., Kim, H. S., Kim, K. L., Jung, J. S., Huh, S. O., ... & Song, D. K. (2001). Protection against β- amyloid peptide toxicity in vivo with long-term administration of ferulic acid. British journal of pharmacology, 133(1), 89-96.
- 42. Yan JiJing, Y. J., Cho JaeYoung, C. J., Kim HeeSung, K. H., Kim KyoungLi, K. K., Jung JunSub, J. J., Huh SungOh, H. S., ... & Song DongKeun, S. D. (2001). Protection against β-amyloid peptide toxicity in vivo with long-term administration of ferulic acid.
- 43. Yang, TK Basu, B. Ooraikul, F. (2001). Studies on germination conditions and antioxidant contents of wheat grain. International Journal of Food Sciences and Nutrition, 52(4), 319-330.
- 44. Yu, H., Hong, J., & Wu, D. (1999). Effect of sodium ferulate on proliferation of rabbit aortic smooth muscle cells induced by oxidized LDL. Zhongguo Zhong yao za zhi= Zhongguo Zhongyao Zazhi= China Journal of Chinese Materia Medica, 24(6), 365-6.
- 45. Yu, J., Vasanthan, T., & Temelli, F. (2001). Analysis of phenolic acids in barley by high-performance liquid chromatography. Journal of agricultural and food chemistry, 49(9), 4352-4358.
- 46. Yu Jing, Y. J., Thava Vasanthan, T. V., & Temelli, F. (2001). Analysis of phenolic acids in barley by high- performance liquid chromatography.
- 47. Yu, L., Haley, S., Perret, J., Harris, M., Wilson, J., & Qian, M. (2002). Free radical scavenging properties of wheat extracts. Journal of agricultural and food chemistry, 50(6), 1619-1624.
- 48. Yun, K. J., Koh, D. J., Kim, S. H., Park, S. J., Ryu, J. H., Kim, D. G., ... & Lee, K. T. (2008). Anti-inflammatory effects of sinapic acid through the suppression of inducible nitric oxide synthase, cyclooxygase-2, and proinflammatory cytokines expressions via nuclear factor-κB inactivation. Journal of agricultural and food chemistry, 56(21), 10265-10272.
- 49. Yun KyungJin, Y. K., Koh DuckJae, K. D., Kim ShiHye, K. S., Park SeungJae, P. S., Ryu JongHoon, R. J., Kim DeogGon, K. D., ... & Lee KyungTae, L. K. (2008). Anti-inflammatory effects of sinapic acid through the suppression of inducible nitric oxide synthase, cyclooxygase-2, and proinflammatory cytokines expressions via nuclear factor-κB inactivation.
- 50. Zang, L. Y., Cosma, G., Gardner, H., Shi, X., Castranova, V., & Vallyathan, V. A. L. (2000). Effect of antioxidant protection by p-coumaric acid on low-density lipoprotein cholesterol oxidation. American Journal of Physiology. Cell Physiology, 279(4), C954-C960.
- 51. Zhao, Z., Egashira, Y., & Sanada, H. (2004). Ferulic acid is quickly absorbed from rat stomach as the free form and then conjugated mainly in liver. The Journal of nutrition, 134(11), 3083-3088.
- 52. Zhao ZhaoHui, Z. Z., Egashira, Y., & Sanada, H. (2004). Ferulic acid is quickly absorbed from rat stomach as the free form and then conjugated mainly in liver.
- 53. Zhou, Z., Robards, K., Helliwell, S., & Blanchard, C. (2004). The distribution of phenolic acids in rice. Food Chemistry, 87(3), 401-406.
- 54. Zhou ZhongKai, Z. Z., Robards, K., Helliwell, S., & Blanchard, C. (2004). The distribution of phenolic acids in rice.
- 55. Zielinski, H., Kozlowska, H., & Lewczuk, B. (2001). Bioactive compounds in the cereal grains before and after hydrothermal processing. Innovative Food Science & Emerging Technologies, 2(3), 159-169.
- 56. Stalikas, C. D. (2007). Extraction, separation, and detection methods for phenolic acids and flavonoids. Journal of separation science, 30(18), 3268-3295.
- 57. Maldonado, A. F. S., Mudge, E., Gänzle, M. G., & Schieber, A. (2014). Extraction and fractionation of phenolic acids and glycoalkaloids from potato peels using acidified water/ethanol-based solvents. Food Research International, 65, 27-34.
- 58. Bubalo, M. C., Ćurko, N., Tomašević, M., Ganić, K. K., & Redovniković, I. R. (2016). Green extraction of grape skin phenolics by using deep eutectic solvents. Food chemistry, 200, 159-166.
- 59. He, B., Zhang, L. L., Yue, X. Y., Liang, J., Jiang, J., Gao, X. L., & Yue, P. X. (2016). Optimization of ultrasound- assisted extraction of phenolic compounds and anthocyanins from blueberry (Vaccinium ashei) wine pomace. Food chemistry, 204, 70-76.
- 60. Lang, Q., & Wai, C. M. (2001). Supercritical fluid extraction in herbal and natural product studies—a practical review. Talanta, 53(4), 771-782.
- 61. Lang QingYong, L. Q., & Wai, C. M. (2001). Supercritical fluid extraction in herbal and natural product studies-a practical review.
- 62. Reverchon, E., & De Marco, I. (2006). Supercritical fluid extraction and fractionation of natural matter. The Journal of Supercritical Fluids, 38(2), 146-166
- 63. Sytar, O. (2015). Phenolic acids in the inflorescences of different varieties of buckwheat and their antioxidant activity. Journal of King Saud University-Science, 27(2), 136-142.
- 64. Cuvelier, M. E., Richard, H., & Berset, C. (1992). Comparison of the antioxidative activity of some acid-phenols: structure-activity relationship. Bioscience, biotechnology, and biochemistry, 56(2), 324-325.
- 65. Zhao, Z., & Moghadasian, M. H. (2008). Chemistry, natural sources, dietary intake and pharmacokinetic properties of ferulic acid: A review. Food Chemistry, 109(4), 691-702.