



A Literature Review of Antioxidant Activity of P. Betle Leaf Extract.

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ABSTRACT:

Piper betle leaf has been considered a natural antioxidant. The P. betle extract shows high antioxidant activity, mostly due to presence of phenolic and flavonoid compounds in the P. betle. Eugenol was discovered to possess antioxidant properties and its percentage ie. (4.86%) was calculated from betel leaf using the Ultrasound extraction process and acidified water used as solvent. The P. betle leaf phenolic content were found to protect photosensitization- mediated lipid peroxidation in rat liver or by free radical scavenging activity and thus enhance antioxidant activity. In a few studies it also been reported that the P. betle with high antioxidant activities suppress the proliferation of cancer cells. In previous study, the study evaluated the anti-oxidative activity of the extract of P. betle leaves (PBLs) on the basis of Cu(2+)- mediated oxidation and it thus has ability to prevent foam cell formation in humans for oxidised low density lipoprotein (ox LDL)-induced lipid accumulation in macrophages. P. betle leaf have numerous properties like antiseptic, anti-oxidant, anti-cancer, anti-inflammatory. P. betle leaf contains many Phytochemicals such as alkaloids, flavonoids, tannis, essential oil, steroids, saponins and suger too.

Keywords: P.betle leaf; Anti-oxidant ; Eugenol ; Ultrasound extraction; Free radical.

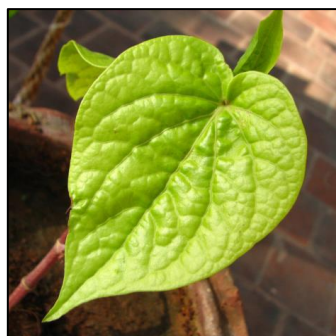
Introduction:

In human, the oxidation process during normal metabolism produces unstable molecule called free radicals. A molecule with one or more than one unpaired electron in its outer shell is known as free radicals. Free radicals causes damage to cell membrane and structure. An anti-oxidant is any substance that can delay or prevent the oxidation of a substrate and protect cells from the damage.^[1] Anti-oxidant are the potential solution to the disease caused by free radical such as neurological disease, cancer, pulmonary disease and condiovascular diseases.^[2] These free radicals can involve in the aging process, thus betel leaf extract can be used as anti -aging agent. Anti-oxidant is used in inhibition of reactive oxygen species (ROS) for treating diseases.^[3]

Mechanism of action:

The anti-oxidant substance works by scavenging activity on free radicals ie. Reactive oxygen species (ROS). From previous studies, it was reported that betel leaf extract exhibited the free radical scavenging activity.^[4,5] In previous study, the study evaluated the anti-oxidate activity of the extract of Piper betle leaves (PBLs) on the basis of Cu(2+)-mediated oxidation and it thus has ability to prevent foam cell formation in human for oxidised low density lipoprotein (oxLDL)-induced lipid accumulation in macrophages. PBLs up regulated the protein levels of the class A and Class B scavenger receptors, its upstream regulator liver X receptor (LXR) and the membrane lipid transporter ABCA1 in macrophages that are exposed to oxLDL. This result minimise the damage of vessels caused by the oxLDL and to prevent foam cell formation and lipid accumulation.^[6]

Information about Piper betle leaf :



[34]

Piper betle L ; synonym: Piper betel Blanco or betel vine, belong to the family *Piperaceae*.^[7] The betel leaf grows on a vine that makes roots. In India, betel leaves are heart-shaped and dark green. They are also referred to as Paan.^[8] Leaf extract and Purified bioactive compounds of P. betle have many beneficial properties like antiseptic, anti-oxidant, anti-cancer, anti-inflammatory. P. betle contains many phytochemicals such as alkaloids, flavonoids, tannins, essential oil, steroids, saponins and sugar too.^[9] India developed a broad variety of P. betle leaves that are traditionally and currently used as a mouth fresher after every meal in India. P. betle is also found and been used by various other countries like Malaysia, Thailand and Srilanka.^[10] P. betle has a qualities including a potent pungent and aromatic flavor, so the Asian people utilize it as a masticator.^[11]

Chemical constituents:

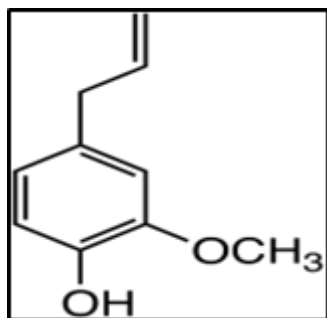
The main phyto-constituent of the betel leaf are mainly phenols & terpenoids. The preliminary studies of aqueous & methanol extract of betel leaves shows the presence of alkaloids, flavonoids, tannins, sterols, phenols, glycosides, saponins & terpenoids as chemical constituent.^[7]

Table no.1^[8]

Sr.no	Chemical compounds	Qt %
1	Eugenol	48.41%.
2	Acetyl Eugenol	14.05%.
3	Alliylpyrocatechol diacetate	0.71%
4	Allylpyrocatechol monoacetate	0.23%
5	Camphene	0.48%
6	Cyrophyllene	1.3%
7	Chavibetol	53%
8	Chavibetol acetate	15.5%
9	Chavicol	11.08%
10	Eucalyptol	1.58%
11	Eugenol acetate	28%
12	Methyl acetate	1.45%
13	α -Copaene	6.43%
14	α -Pinene	0.23%
15	β -Cubebene	13.06%
16	1,8 cineol	0.04%
17	3, methoxy cinnamonaldehyde	1.38%

Eugenol is a phenolic compound from the class of phenylpropanoid. The chemical composition of Eugenol is (4-allyl 2-methoxyphenol). Eugenol is a natural antioxidant.^[35]

Eugenol



Eugenol is pale yellow liquid and has an oily consistency. It has a spicy aroma. Eugenol is sparingly soluble in water and well soluble in organic solvent. Eugenol is a monoamine oxidase (MAO) inhibitor & it also exhibit micro-protective properties. It inhibits the generation of reactive oxygen species and prevents the formation of reactive forms of nitrogen & it also increases the cyto-antioxidant potential. The antioxidative property of Eugenol is due to its structure, it allows it to fix phenoxy radicals by receiving donated Hydrogen atoms.^[36]

Species:

There are over 100 different types of species of betel leaf are discovered. Over 40 are found in India Only. From them, Bagerhati, Ghanagete & Kauri are considered to be rich in antioxidant.^[14] There are mainly 5 cultivator of betel vine These care Desawari, Bangla, Kapoorin, Meetha & Sanchi. Kaptori & Sanchi are common in peninsular India. Bangla & Desivan are common in North India. Meetha is grown in West Bengali Only. The season for optimum cultivation are in between November to February.^[13]

Methods for extraction of Eugenol :**1.Steam distillation method :-**

-It is the most popular Method for isolating eugenol. In this procedure, essential oil is first collected from the plant.

-Eugenol is then extracted by mixing the essential oil with 3% solution of sodium potassium hydroxide. The product of this reaction is phenolic alkali salt.

- Next, the extracts insoluble component is using steam distillation or solvent extraction.^[37]

2.Solvent extraction method :-

-Another Method for extracting ethanol is by employing different solvents, such as methanol, ethanol, and petroleum ether. Using an appropriate organic solvent in a Soxhlet system, extraction is performed.^[15]

- The extracted materials are concentrated at 50°C to complete the process.

3.Ultrasound extraction method :-

-Ultrasonic baths and probes, which use piezoelectric transducers as their source of ultrasonic power, are typically used for UAE procedures. The solid matrix is distributed in the solvent in a stainless steel tank that is coupled to a transducer in an ultrasonic bath. Because of the probe's higher ultrasonic intensity (tip), probe-based systems are frequently chosen over bath systems and are effective tools for extracting bioactive chemicals due to their non-uniform.^[16]Through the probe, the ultrasonic energy is emitted into the sample.^[17]

-The diameter of the laboratory scale probe tip that is commercially available ranges from 2mm, which can handle samples up to 5mL, to 25mm, which can handle samples up to 1litre. Pectin was extracted from pomegranate^[18] and jackfruit peels,^[19]and polysaccharide from rambutan peels^[20] using a 20mm flat tip probe. Xu et al^[21], extracted pectin from grapefruit peel using 13mm and 25mm probes.

Evaluation Parameter for Antioxidant property:

For the study of the antioxidant effects in betel leaf extract, the inhibition of lipid peroxidation, superoxide generation and the activities of superoxide or hydroxyl radical scavenging are usually evaluated.^[22]

Table no.2 ^[22]

Sr. No.	Parameter	Observation
1	FeCl ₂ - Ascorbic Acid stimulated Lipid Peroxidation in Rat Liver Homogenate	Significant increase in GMDA (Malandialdehyde) in test compared to normal control.
2	Inhibition of Xanthine Oxidase	The activity of xanthine oxidase was inhibited.
3	Free Radical Scavenger Activity	Antioxidant enzyme such as SOD, P-ase, GSH, C-ase, phenolic,etc., lower the steady state concentration of ROS in cells & various tissue and there by prevent oxidative substance.
4	Enzyme assay from Herbal Extract by following method:	SOD activity is determined.

5	(a) Assay for DPPH free radical scavenging activity	IC50 value denote the concentration of sample which is required to scavenge 50% DPPH free radicals.
6	Phospholipid Peroxidation	Inhibits peroxidation membrane lipid.

Free Radical Scavenger Activity:

1. Samples are dissolved in 0.1m PBS to each concentration. Then 50ul of sample solution and 400ul working solution (xanthine/cytochrome c) are added to 530ul H₂O, mixed and then 20ul of 1 units/ml of xanthine oxidase/ PBS are added, mixed vigorously and screened for 2 minutes at 550 nm.
2. Previous studies evaluated that the ESR technique is used to confirm the free radical scavenger activity.
3. Antioxidant enzymes which include super-oxide-dismutase (SOD), peroxidase (P-ase) and catalase (Case) enzyme, natural antioxidant include reduced glutathione (GSH), phenolics (tocopherols, flavonoids, phenolic acids, etc) contain activity of lowering the steady-state concentration of ROS in cells and many tissue and thus prevent oxidative substance.^[23]

Assay for DPPH free radical scavenging activity:

1. DPPH (1,1-diphenyl-2-picryl-hydrazyl) is considered as a simple, rapid, stable, inexpensive and can be used widely, controlled method.^[24]
2. DPPH can make stable free radicals in aqueous or ethanol solution.^[23] Then, the free radical will undergo further reaction and thus create a stable product, while DPPH will accept an hydrogen or electron radical to become stable molecule.^[25,26]
3. The antioxidant potency was previously evaluated through free radical scavenging by test samples, the change of optical density of DPPH radicals is monitored. [Lee et al, 1998]
4. Reaction mixtures containing test samples (5ul of methanol: water 7:3 plant extract, dissolved in dimethyl sulphoxide (DMSO)) and 316umol/l DPPH ethanol solution (95ul, final DPPH concentration 300umol/l) in 96- well microtitre plates are incubated at 37°C or 30min and absorbance was measured at 515nm. The % of inhibition by sample treatment is determined by comparison with a DMSO-treated control group. IC50 values denote the concentration of sample which is required to scavenge 50% DPPH free radicals.^[23]

Result & Discussion:

The data gathered shows that eugenol in betel leaf shows higher anti-oxidant activity by the mechanism of scavenging activity. From previous study, the result indicate that the presence of phenolic compound contributes to the antioxidant activity of the leaves and thus act as hydrogen donor, singlet oxygen quencher, heavy metal chelator, reducing agent.^[27] The studies reported that increased production of free radicals in body escorts disturbance in antioxidant status, leading to cellular oxidative damage.^[28] Eugenol acts as bioactive phytochemical against SOD (Superoxide dismutase) and MDA (Malondialdehyde) profiles in hypercholesterolemic rats.^[29] The previous study reported that phenolic compound and flavonoids compound of betel leaf have both chemotherapeutic and chemo-preventive effects.^[30,31,32] The study also result into showing anti-cancer activity of the P. betle extract due to high anti-oxidant activity.^[33]

Conclusion:

From the data gathered it was concluded that by Ultrasound Extraction method more eugenol extract is obtained. There are various methods for extraction but by ultrasound extraction the yield of eugenol is more. As study indicates that P. betle leaf extract has phenolic and flavonoid compound result into more anti-oxidant activity. Various formulation can also be prepared from betel leaf like cream, transdermal patches, suspension and many more. It was also seen that betel leaf can be used for ulcer treatment.

Reference:

1. <https://www.betterhealth.vic.gov.au/health/healthyliving/antioxidants>
2. RINTU D, SHINJINI M, KAUSTAB M, PRAMATHADHIP P, UMESH PS AND BANERJEE ER. 2015. Anti-oxidant and anti-inflammatory activities of different varieties of Piper leaf extracts (Piper betle L.). Nutr Food Sci 5(5): 1-15.
3. HOQUE MM, RATTILA S. SHISHIR MA, BARI ML, INATSU Y AND KAWAMOTO S. 2012. Antibacterial activity of ethanol extract of betel leaf (Piper betle L.) against some food borne pathogens. Bangladesh J Microbiol 28(2): 58-63,
4. LEE OH, LEE BY, LEE J, LEE HB, SON JY, PARK CS, SHETTY K AND KIM YC. 2009. Assessment of phenolics-enriched extract and fractions of olive leaves and their antioxidant activities. Bioresour Technol 100(23): 6107-6113.

5. SHAH SK, GARG G, JHADE D AND PATEL N. 2016. Piper betle: phytochemical, pharmacological and nutritional value in health management. *Int J Pharm Sei Rev Res* 38: 181-189.
6. Gwo-Chin Ma I, Pei-Fang Wu, Hsien-Chun Tseng, Charng-Cherng Chyau, Hsiu-Chin Lu, Fen-Pi Chou Affiliation IInstitute of Biochemistry and Biotechnology, College of Medicine, Chung Shan Medical University, Taichung, Taiwan. 128729@cch.org.tw.
7. <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC9170825/>
8. https://wjpr.s3.ap-south-1.amazonaws.com/article_issue/eac4f875dfde7735afbb5aa5af087216.pdf
9. Khan, Z., Bashir, O., Hussain, J.I., Kumar, S., & Ahmad, R. (2012). Effects of ionic surfactants on the morphology of silver nanoparticles using Paan (Piper betel) leaf petiole extract. *Colloids and Surfaces B, Biointerfaces*, 98, 85–90. 10.1016/j.colsurfb.2012.04.033.
10. Ekambaram P, Balan C. Efficacy of salivary and diastase extracts of Piper betle in modulating the cellular stress in placental trophoblast during preeclampsia. *Pharmacognosy Res* 2019;11:25-30.
11. Tholkappian, C. (2014). Organic and conventional betel leaf cultivation: Cost–benefit analysis. *International Journal of Research*, 1(4), 929–938.
12. Heliawati, L., Lestari, S., Hasanah, U., Ajiati, D., & Kurnia, D. (2022). Phytochemical profile of antibacterial agents from red betel leaf (piper scrotum Ruiz and Pav) against bacteria in dental caries. *Molecules*, 27(9), 2861. <https://doi.org/10.3390/molecules27092861> (Basel, Switzerland).
13. <https://actascientific.com/ASNH/pdf/ASNH-07-1170.pdf>
14. <https://www.sciencedirect.com/science/article/abs/pii/S0308814604001050>
15. G. Wenqiang, L. Shufen, Y. Ruixiang, T. Shaokun and Q. Can, Comparison of essential oils of clove budsextracted with supercritical carbon dioxide and otherthree traditional extraction methods, *Food Chem.*, 2007, *101*, 1558-1564.
16. Chemat F., Rombaut N., Meullemiestre A., Turk M., Perino S., Fabiano-Tixier A.S., Abert-Vian M. Review of green food processing techniques. Preservation, transformation, and extraction. *Innov. Food Sci. Emerg. Technol.* 2017;41:357–377.
17. Adetunji L.R., Adekunle A., Orsat V., Raghavan V. Advances in the pectin production process using novel extraction techniques: a review. *Food Hydrocolloids*. 2017;62:239–250.
18. Moorthy I.G., Maran J.P., Muneeswari S., Naganyashree S., Shivamathi C.S. Response surface optimization of ultrasound assisted extraction of pectin from pomegranate peel. *Int. J. Biol. Macromol.* 2015;72:1323–1328.
19. Moorthy I.G., Maran J.P., Ilakya S., Anitha S.L., Sabarima S.P., Priya B. Ultrasound assisted extraction of pectin from waste Artocarpus heterophyllus fruit peel. *Ultrason. Sonochem.* 2017;34:525–530.
20. Maran J.P., Priya B. Ultrasound-assisted extraction of polysaccharide from Nephelium lappaceum L. fruit peel. *Int. J. Biol. Macromol.* 2014;70:530–536.
21. Xu Y., Zhang L., Bailina Y., Ge Z., Ding T., Ye X., Liu D. Effects of ultrasound and/or heating on the extraction of pectin from grapefruit peel. *J. Food Eng.* 2014;126:72–81.
22. Mukherjee P. 2002, Quality control of herbal drugs, first edition, Pharmaceutical publication by BUSINESS HORIZONS, INDIA:562-565.
23. Mukherjee P. 2002, Quality control of herbal drugs, first edition, Pharmaceutical publication by BUSINESS HORIZONS, INDIA:563-564.
24. Sirivibulkovit, K.; Nouanthavong, S.; Sameenoi, Y. Paper-based DPPH Assay for Antioxidant Activity Analysis. *Anal. Sci.* 2018, 34, 795–800. [CrossRef]
25. Blois, M.S. Antioxidant Determinations by the Use of a Stable Free Radical. *Nature* 1958, 181, 1199–1200. [CrossRef]
26. Ionita, P. The Chemistry of DPPH-Free Radical and Congeners. *Int. J. Mol. Sci.* 2021, 22, 1545. [CrossRef] [PubMed]
27. JHA A, UPADHYAY A, RASANE P and SINGH HB. 2011. Quantitative studies of phytochemicals of selected green leafy vegetables and their antioxidant potential. *Med Plant-Int J Phytomed Rel Ind* 3(2): 113-117.
28. E. Hopps, D. Noto, G. Caimi and M. R. Averna, A novel component of the metabolic syndrome: the oxidative stress, *Nutr., Metab. Cardiovasc. Dis.*, 2010, 20, 72–77.
29. A. Munisa, W. Manalu, T. Wresdiyati and N. Kusumorini, The Effect of Clove Leaf Methanol Extract on the Proles of Superoxide Dismutase and Malondialdehyde in the Liver of Rabbits under Hypercholesterolemia Condition, *Transl. Biomed.*, 2015, 6, 1–5, DOI: 10.21767/2172-0479.100012.

30. Seeram NP, Zhang Y, Nair MG. Inhibition of proliferation of human cancer cells and cyclooxygenase enzymes by anthocyanidins and catechins. *Nutr Cancer* 2003; 46: 101–106.
31. Thangapazham RL, Singh AK, Sharma A, Warren J, Gaddipati JP, Maheshwari RK. Green tea polyphenols and its constituent epigallocatechin gallate inhibits proliferation of human breast cancer cells in vitro and in vivo. *Cancer Lett* 2007; 245: 232–241.
32. Li WY, Chan SW, Guo DJ, Yu PHF. Correlation between antioxidative power and anticancer activity in herbs from traditional Chinese medicine formulae with anticancer therapeutic effect. *Pharm Biol* 2007; 45: 541–546.
33. Nakagawa Y, Suzuki T, Nakajima K, Ishii H, Ogata A. Biotransformation and cytotoxic effects of hydroxychavicol, an intermediate of safrole metabolism, in isolated rat hepatocytes. *Chem Biol Interact* 2009; 180: 89-97.
34. <https://images.app.goo.gl/t2hvFSMBihd5ZMdp9>
35. <https://www.sciencedirect.com/topics/agricultural-and-biological-sciences/eugenol>
36. <https://www.mdpi.com/1422-0067/22/7/3671>
37. S. K. Mahapatra, S. P. Chakraborty, S. Majumdar, B. G. Bag And S. Roy, Eugenol protects nicotine-induced superoxide Mediated oxidative damage in murine peritoneal Macrophages in vitro, *Eur. J. Pharmacol.*, 2009, 623, 132–140