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## **Innovative Scaffolds: Enhancing the Delivery of Drugs and Biologics**

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### **ABSTRACT:**

Biodegradable materials called scaffolds are used to introduce medications, genes, and cells into the body. They replicate biological mechanisms that stimulate cell adhesion, extension, proliferation, and differentiation and are administered as implants or injections to aid in tissue regeneration. Scaffold matrices' biocompatibility, biodegradability, interface adherence, drug distribution, porosity, binding affinity, stability, and loading capacity all affect how well they carry drugs and cells. Applications for scaffolds include medication delivery, tissue engineering, antibiotic delivery, wound healing, bone repair, and diabetes treatment. Various processes, such as solvent diffusion, pyrogen leaching, emulsion and phase separation, electrospinning, and stereolithography, are employed in their fabrication.

**Keywords:** diabetes, scaffolds, biologics, embryonic stem cells.

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### **Introduction:**

Biodegradable scaffolds, which are infused with cells, proteins, growth hormones, and genes, aid in the generation of tissues. The concept of tissue reconstruction was first proposed by Robert Langer and colleagues in the early 1990s, using artificial polymer matrices to receive cell transplants. A number of factors have limited the adoption of the novel scaffolding technology from the research labs in the clinic, including a lack of understanding regarding the necessary materials and design elements, the need for a thorough scale-up procedure for biomaterials with intricate three-dimensional structures; the degradation of scaffold material in clinical applications and their regulatory requirements are unclear. The necessity to use computer-aided design methods for making scaffolds. The necessity of improving material comprehension.

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### **Applications of scaffolds:**

Tissue engineering requires a scaffold to act as a source for cell adhesion, extension, proliferation, and differentiation. The target tissue will grow when the scaffold resembles the biological requirements of the regeneration tissue. For tissue engineering, the ideal material should generally have a high capacity for absorbing liquids, sufficient gas permeability, biocompatibility, and antibacterial properties to protect the skin from further infections, dehydration, and resulting tissue damage. monitoring cell density and location in addition to sufficient levels of fibrinogen, thrombin, and aprotinin. determined the ideal cell culture parameters for the proliferation and differentiation of embryonic stem (ES) cells in fibrin scaffolds (protease inhibitors). Determining the proper dosages of thrombin and fibrinogen for the neural lineage progenitor cells generated from embryonic stem cells. It is necessary to add aprotinin, a plasmin inhibitor, to the media in order to prevent scaffold destruction by ES cells. The best cell preparation technique for good ES cell proliferation and differentiation in 3D cultures was found to be using a single intact EB enclosed within a fibrin scaffold [1]. After incubation for 14 days within fibrin scaffolds, these cells differentiated into neurons and astrocytes. Employing biopolymers with proven biocompatibility and bioresorbable have shown that an injectable, in situ-forming, non-toxic, biodegradable polymer scaffold may be created without the need for additional crosslinking agents. Rapid crosslinking and gelation with gelatine in the presence of borax is feasible to produce a truly injectable system, as demonstrated by the use of oxidized alginate with the appropriate molecular weight and degree of oxidation.

### **Drug delivery:**

Chondrocytes were given using elastic hydrogel scaffolds, which are constructed of hydrophobic poly(-caprolactone) and hydrophilic poly (ethylene glycol) (PEG) for the growth of neocartilage rabbits. These scaffolds are made by and are biodegradable. Chondrocytes were employed to study the in vitro cell interactions of salt leaching, an often-used technique. While the hydrogel scaffold with a comparatively low PEG concentration showed poorer chondrogenic development, the scaffold with a high PEG content indicated better chondrocyte cell proliferation. Drug delivery using dextran hydrogels has been investigated in the past, but there was a catch: tissue engineering had already made use of dextran's non-adhesive properties. It was possible to develop microporous, cell-adhesive, and cell-penetrable qualities [2-5]. Due to specific interactions between the substance and the cell, covalent Neurite outgrowth and cell adhesion were enhanced by ECM-derived peptide therapy. used a twofold emulsion/solvent extraction method to encapsulate doxorubicin in poly (D, L-lactide-co-glycoside) (PLGA).

**Wound healing:**

In order to improve stability and solubility, integrated curcumin into chitosan nanoparticles (CSNPs). The resulting Curcumin-CSNPs were then implanted into collagen scaffolds (a unique nanohybrid scaffold) for improved tissue regeneration applications. The results of this study indicate that a synergistic combination of collagen, an established wound healer acting as a scaffold, chitosan, an anti-inflammatory and antioxidant drug carrier, and curcumin is a valuable tactic for addressing various pathological signs of diabetic wounds and enhancing wound healing capability. A collagen/chitosan (COL-CS) composite scaffold loaded with L-glutamic acid (LGA) was developed by Sanpaolo to aid in the healing of diabetic wounds. The characterization results of the composite scaffold showed that a crosslinked scaffold exhibits excellent porosity, low matrix degradation, and sustained drug release when compared to a non-crosslinked scaffold. Growth factors like Basic Fibroblast Growth Factor (BFGF) and vascular endothelial growth factor (VEGF) are widely known to speed up wound healing and promote cell proliferation. Losi and colleagues synthesized a scaffold consisting of poly(ether)urethane, polydimethylsiloxane, and fibrin using PLGA nanoparticles [6].

**Treatment for diabetes:**

The ability of human embryonic stem cells to differentiate into insulin-producing cells was demonstrated, yet only 1-3 percent of the cells in differentiated embryoid bodies (EBs) were insulin-positive. A five-step procedure can be utilized to create islet-like cells from human embryonic stem cells (ES), according to research. Exendin-4 was utilized in place of nicotinamide. This work suggests that transforming human embryonic stem cells (ES cells) into islet-like cells could be achieved with a five-step technique that includes exendin-4. Additionally, it raises the possibility of using scaffolds to move islet-like cells to desired locations. In order to improve the biological and pharmacokinetic properties of xanthine derivatives used in the treatment of diabetes, synthesized novel chitosan formulations. To evaluate the effectiveness of the optimized formulations, measurements of particle size, shape, degree of swelling, Fourier transform infrared analysis, and X-ray diffraction were made.

**Healing of the bones:**

An infection from an open fracture can delay bone healing, result in limb loss, or possibly result in death. As of right now, treating infected open fractures with antibiotic-loaded non-biodegradable poly (methyl methacrylate) (PMMA) beads is the clinical standard of care. The infection is subsequently controlled, and bone grafting is carried out. Tissue regeneration includes the formation of new blood vessels, which can take up to six weeks in segmental defects [7-10]. To prevent infection of the implant and allow for the unhindered growth of new bone and vascularization, a biodegradable bone graft containing a slow-releasing antibiotic is required.

**Administration of antibiotics:**

Antibiotics must be administered in a targeted and timely manner in order to treat dental, periodontal, and bone infections with high local bioactivity and minimal systemic adverse effects. In order to prevent bacteria from growing for an extended period of time, a 3-D porous tissue engineering scaffold was created in this study that could deliver antibiotics in a regulated manner. loaded the water-soluble antibiotic medication doxycycline (DOXY) into polylactic acid nanospheres using a modified water-in-oil (w/o/o) emulsion technique. Next, the prefabricated nanofibrous PLLA scaffolds with a well-connected, microporous structure were filled with PLGA nanospheres (NS).

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**Methods of preparation:**

In order to create an efficient scaffold with the best mechanical and mass-transport capabilities, the first stage in the manufacturing process is to create a base material and turn it into a 3D structure. In the same way that elastomeric polymers will provide appropriate functionality for soft tissue, titanium, for instance, will not be compliant when it comes to soft tissue repair. Additionally, there are middle-of-the-road polymers that are functionally acceptable for both soft and hard tissues, such as bone. Conventional techniques, including solvent diffusion, emulsion and pyrogen leaching, and electrospinning, are utilized to create scaffolds with excellent structural qualities and high porosity [11].

**3.1 Diffusion of solvents:**

chloroform (25% w/v) was used to cast PLA into ceramic Molds, and the solvent was allowed to evaporate at room pressure or in a vacuum at 1500 Hg. Until there was no more solubilized PLA in the mold, the process was repeated.

**1.2 Seepage of Pathogens:**

Pyrogens with varying forms are utilized to obtain the appropriate pore shape. Zhang and associates developed a combination of compression moulding and pyrogen leaching methods for polylactic acid (PLA)/poly-glycoside (PGA) scaffolds with a ratio of 85:15 of spherical and cubic-shaped pyrogens, respectively, by using the vibrating-particle approach, a type of pyrogen leaching. There was a 78%–97% porosity observed.

**1.3 Phase division and emulsion:**

Thermally induced phase separation (TIPS) was used by Nam and Park to build scaffolds of poly-L-Lactides (PLLA) and PLGA with a connected pore structure and fine structural features [12-15]. In order to extract the solvent, the polymer is dissolved in a solvent, rapidly frozen in liquid nitrogen, and then freeze-dried for three days.

#### **1.4 Spine Electrodes:**

Materials of sizes varying from nanometres to millimetres are utilized, such as collagen, chitosan, PCL, PLGA, and others. A powerful electric field is applied to the nozzle to melt the polymer droplet in solution, creating the fibres. Extruded fibres are amorphous unless they spin along a revolving drum to arrange themselves. Qualities of anisotropic effective scaffold are generated by amorphous fibres, whereas aligned fibres create anisotropic effective scaffold characteristics.

#### **1.5 The technique of stereolithography**

requires the application of a material that can be photopolymerized. Cooke and colleagues were the first to show how to fabricate scaffolds from polypropylene fumarate (PPF) by utilizing a biodegradable resin mixture consisting of diethyl fumarate (DEF), PPF, and bicycle phosphine oxide (BAPO) photoinitiated. With the aid of stereolithography, Lee et al. produced PPF scaffolds with a variety of 3D porous topologies. The porosity varied from 30 to 63%, the compressive moduli from 15 to 40 MPa, and the pore diameters from 500 to 900  $\mu$ m. These substances may be used to replace trabecular bone [16].

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### **4. Assessment:**

#### **4.1 Tests for Cytotoxicity:**

The term "cytotoxicity" describes a set of molecular processes that disrupt the synthesis of macromolecules, leading to various types of structural, functional, and cellular damage. In these investigations, substances extracted from scaffolds that have been examined are exposed to different cell culture lines. When potentially hazardous substances are present, cell cultures show rapid toxicity and are especially sensitive to minute concentrations of leachable chemicals.

#### **4.2 Assay for indirect contact:**

These experiments determine how the medicines leak from the scaffolds. The extraction test assesses the morphological alterations, growth inhibition, and metabolic activity of the cell. Scaffolds are incubated for 12 to 72 hours at a specific temperature of 37 °C. These are viability assays to evaluate the extract's impact on cell cytotoxicity. In order to determine viability, the most fundamental method is trypan blue exclusion. Under a light microscope, live cells seem transparent because they may release trypan blue from their cytoplasm, while nonviable cells appear blue.

#### **4.3 Assays using direct contact:**

In order to investigate the cytocompatibility of decellularized scaffolds, most researchers choose to employ the direct contact experiment since it enables them to analyse the attachment, motility, and dispersion of reseeded cells within the scaffolds. A sample comes into direct contact with cells during surface culturing. Cells are examined morphologically and by viability tests to check for signs of toxicity at different times.

#### **4.4 Risk of pathogenicity:**

Pigs are commonly employed as xenograft donors despite the fact that they differ from humans immunologically. Because they are simpler to breed, pigs are utilized. Typically, these animals are home to viruses like the porcine endogenous retrovirus (PERV), which, it was discovered, was integrated into the pig genome even in animals that were designated as pathogen-free (SPF). Polymerase chain reaction (PCR) is a specialized assay for identifying proviral PERV in decellularized tissues.

#### **4.5 The ability to produce immunity:**

To prevent rejection and vascular thrombosis rejection and stimulate immunity, it is crucial to make sure that there are no cellular components, antigens, or nucleic materials present.

#### **4.6 Genetics:**

DNA remnants must be removed from the ECM because they have the potential to cause unfavourable host responses in vivo and increase the risk of exoantigen transmission. Using tools like spectroscopy, slot blot, and Pico Green makes quantifying DNA easy. The length of any remaining DNA fragments could be ascertained by gel electrophoresis.

#### 4.7 Galactose Assessing:

The recipient's reaction to decellularized tissues post-implantation is crucial, especially in cases where xenogeneic decellularized tissues are being used. This includes monitoring the recipient's immune system. Remaining on the cell surface, the alpha-Gal (Gal1-3Gal1-4GlcNAc-R) epitope plays a crucial role in the rejection of xenogeneic tissue. The gal epitope is expressed in most mammalian organs, with the exception of humans and higher apes. Immunohistochemistry and PCR can be used to quantify the amount of Gal present in decellularized tissues.

#### 4.8 In vivo assessment:

Biological testing of scaffolds derived from people, pigs, and cattle has been done through animal testing prior to clinical application in a variety of animal species, including swine, mice, and rats. A biomaterial that has been implanted in the body may cause a host reaction. The degree of this response and in vivo performance against scaffolds can be classified into an acute and chronic inflammatory response followed by a long-lasting granulation tissue phase, depending on the properties of the implant, such as size, morphology, composition, mechanical stability, sterility issues, contact duration, and degradation. To investigate the inflammatory response, several quantitative assays have been employed. After being evaluated by multiple scoring systems, cell adhesion, one of the examined criteria, has been used as a biocompatibility indicator.

#### Prospective features:

The advancement of additive manufacturing (AM) techniques over the past ten years has been beneficial to tissue engineering (TE), since it has led to the production of free-form porous scaffolds with customized topologies. The most recent ASTM standards define additive manufacturing (AM) as "a process of combining materials to produce items using three-dimensional (3-D) model data, usually layer upon layer, as opposed to subtractive manufacturing methodologies." Additive manufacturing builds the finished thing by adding material layers from a 3-D computer model, in contrast to traditional subtractive processes that remove material from a 3-D block. Two-dimensional (2-D) layers from the three-dimensional (3-D) model are "sliced" and sent to the AM device to create the final result. Precision extrusion deposition (PED), fused deposition modelling (FDM), stereolithography (SLA), and selective laser sintering (SLS). The number of strategies aimed at establishing the optimal scaffold design for a particular therapeutic application has increased dramatically since the widespread usage of computer-aided tissue engineering (CATE).

#### Conclusion:

In summary, the development of scaffold-based delivery systems represents a significant advancement in medical science, with the potential to revolutionize the way drugs and biologics are administered, ultimately leading to better patient outcomes and enhanced therapeutic efficacy.

#### REFERENCE:

1. SCAFFOLDS: A PANACEA FOR DRUGS AND BIOLOGICS DELIVERY", IJCSPUB - INTERNATIONAL JOURNAL OF CURRENT SCIENCE (www.IJCSPUB.org), ISSN: 2250-1770, Vol. 12, Issue 3, page no. 428-435, September 2022, Available: <https://rjpn.org/IJCSPUB/papers/IJCSP22C1271.pdf>
2. Dutta, R. C., & Dutta, A. K. (2009). Cell-interactive 3D-scaffold; advances and applications. *Biotechnology Advances*, 27(4), 334-339. <https://doi.org/10.1016/j.biotechadv.2009.02.002>
3. Lee, J. Y., Bashur, C. A., Goldstein, A. S., & Schmidt, C. E. (2009). Polypyrone-coated electro spun PLGA nanofibers for neural tissue applications. *Biomaterials*, 30(26), 4325-4335. <https://doi.org/10.1016/j.biomaterials.2009.04.042>
4. Nguyen, M. K., & Lee, D. S. (2010). Injectable biodegradable hydrogels. *Macromolecular Bioscience*, 10(6), 563-579. <https://doi.org/10.1002/mabi.200900358>
5. O'Brien, F. J. (2011). Biomaterials & scaffolds for tissue engineering. *Materials Today*, 14(3), 88-95. [https://doi.org/10.1016/S1369-7021\(11\)70058-X](https://doi.org/10.1016/S1369-7021(11)70058-X)
6. Wang, H., Boerman, O. C., Sariibrahimoglu, K., Li, Y., Jansen, J. A., & Leeuwenburgh, S. C. G. (2012). Preparation and characterization of injectable cements based on brushite nanoparticles. *Acta Biomaterial*, 8(8), 2732-2739. <https://doi.org/10.1016/j.actbio.2012.03.021>
7. Bose, S., Aghazadeh, S., & Bandyopadhyay, A. (2013). Bone tissue engineering using 3D printing. *Materials Today*, 16(12), 496-504. <https://doi.org/10.1016/j.mattod.2013.11.017>
8. Chen, Q., & Zhu, C. (2016). Fabrication of polymer scaffolds for biomedical applications. *Macromolecular Materials and Engineering*, 301(1), 17-25. <https://doi.org/10.1002/mame.201500176>
9. Kim, T. G., & Park, T. G. (2014). Surface functionalized electro spun biodegradable nanofibers for tissue engineering and drug delivery. *Advanced Drug Delivery Reviews*, 61(6), 301-316. <https://doi.org/10.1016/j.addr.2009.12.009>
10. Li, J., He, Y., Hao, Y., & Qian, Y. (2014). Injectable hydrogel systems based on functionalized natural polymers for tissue regeneration. *Journal of Biomedical Materials Research Part A*, 102(4), 1150-1162. <https://doi.org/10.1002/jbm.a.34811>
11. Ma, P. X. (2008). Biomimetic materials for tissue engineering. *Advanced Drug Delivery Reviews*, 60(2), 184-198. <https://doi.org/10.1016/j.addr.2007.08.041>
12. Place, E. S., George, J. H., Williams, C. K., & Stevens, M. M. (2009). Synthetic polymer scaffolds for tissue engineering. *Nature Materials*, 8(6), 457-470. <https://doi.org/10.1038/nmat2446>

13. Raghunath, J., Rollo, B., Sales, K. M., Butler, P. E., & Seinfeldian, A. M. (2007). Biomaterials and scaffold design: Key to tissue-engineering applications. *Biotechnology and Applied Biochemistry*, 46(2), 73-84. <https://doi.org/10.1042/BA20060128>
14. Saravanan, S., Leena, R. S., & Velmurugan, N. (2016). Chitosan based bio composite scaffolds for bone tissue engineering. *International Journal of Biological Macromolecules*, 93, 1354-1365. <https://doi.org/10.1016/j.ijbiomac.2016.06.017>
15. Stuckey, D. J., & Luol, M. P. (2009). Nanostructured hydrogels for 3D cell culture: R&D using microfluidic systems. *Lab on a Chip*, 9(1), 45-52. <https://doi.org/10.1039/B811124A>
16. Zhang, S., Geryak, R., Geld Meier, J., Kim, S., & Tsuru, V. V. (2017). Synthesis, assembly, and applications of hybrid nanostructures for biosensing. *Chemical Reviews*, 117(20), 12942-13038. <https://doi.org/10.1021/acs.chemrev.7b00204>