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Screening of Anticonvulsant Profile of *Habenaria Intermedia* Tubers in Rats Using Well Established Model

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ABSTRACT

A review of the literature found that *Habenaria intermedia* has not had any systematic pharmacological research done to support the traditional claims made for the majority of its neuropharmacological activities, particularly for its anticonvulsant profile. Therefore, it was thought to be beneficial to test *H. intermedia* tubers anticonvulsant profile. The hydroalcoholic extract and its ethyl acetate fraction of plant tubers were prepared using reference procedures and showing presence of alkaloids, flavonoids, coumarins and tannins as major classes of compounds. The two parametres were accessed such as mean onset of action (sec) and mortality (%) in anticonvulsant activity screening and results are described in table 15 and figure 2 following the administration of hydroalcoholic extract (100, 200, or 400 mg/kg, p.o.), ethyl acetate fraction (25 or 50 mg/kg, p.o.), diazepam (2 mg/kg, p.o.), and the control (vehicle, p.o.). The experimental protocol was divided into three groups such as control, standard and test. Each group of animals received pentylenetetrazole (80 mg/kg, i.p.) before 45-60 min prior takes control, standard and test drugs. The hydroalcoholic extract (400 mg/kg) and its ethyl acetate fraction (50 mg/kg) of *Habenaria intermedia* tubers were exhibit anticonvulsant activities, it can be suggested that anticolvulsant activity of *Habenaria intermedia* tubers are attributed to these natural classes of compounds.

Key words: Habenaria intermedia, anticonvulsant, pentylenetetrazole, flavonoids, phenols.

Introduction

Mainly found in India (Himalayas, Himachal Pradesh, Kashmir, Uttarakhand, and Sikkim), Pakistan, Nepal, and Bhutan is *Habenaria intermedia* D. Don (Riddhi; family: Orchidaceae) (Balkrishna et al., 2012). Traditional medicine has made extensive use of *Habenaria intermedia* D. Don to treat a variety of illnesses. It serves as a tonic for health. It is a component of the Astavarga formulation, which consists of eight rejuvenating medications. Additionally, it goes into making "Chyawanprash," a popular Ayurvedic tonic. According to Khare (2007), Kirtikar and Basu (1994), Warrier et al. (1994), its tubers are used as an emollient (having a soothing/cooling effect), an aphrodisiac, a depurative (removing impurities from the body), an anthelmintic, a nervine, and a cardiac tonic. They are also used to treat leprosy, burning sensations, asthma, fever, cold, insanity, and other skin diseases (Prajapati et al., 2003). According to Jagetia et al. (2004), it has antioxidant action. Additionally, it is utilized in the production of "Salep," a tuber-based flour (Ahuja, 2003). Following a review of the literature, it was discovered that H. intermedia includes flavonoids, tannins, phenol (gallic acid) (Goudar et al., 2015; Habbu et al., 2012), coumarin (scopoletin) (Habbu et al., 2012; Reynolds and Wilson, 1991), and alkaloids (Rajashekhar et al., 2012), and hepatoprotective (Goudar et al., 2015) properties *of H. intermedia* have all been thoroughly studied. A study of the literature revealed that the majority of the conventional claims made for *H. intermedia* neuropharmacological activities—particularly for its anticonvulsant profile—have not been supported by systematic pharmacological research. Testing the anticonvulsant profile of *H. intermedia* tubers was therefore deemed advantageous.

MATERIALS AND METHODS

Plant material

Habenaria intermedia tubers were procured from Himalaya Herbs Store, Madhav Nagar, Saharanpur, (Uttar Pradesh), India in January, 2024. The identity of plant was confirmed from online literature related to their microscopic characters.

Preparation of test samples and their chemical testing

Using Soxhlet apparatus technology, the hydroalcoholic extract from dried plant material was made after defating with n-hexane solvent. Additionally, reflux technology was used to generate the ethyl acetate fraction from the hydroalcoholic extract (Kumar and Kumar, 2015; Prakash et al., 2015). To

determine if phytomolecules were present or absent, a chemical screening process was applied to both the hydroalcoholic extract and its ethyl acetate fraction (Farnsworth, 1966).

Anticonvulsant activity

Animals

Male SD rats of body weight 250-300 g, procured from Akal College of Pharmacy and Technical Education, Mastuana Sahib, Sangrur were used for anticonvulsant activity using using pentylenetetrazole seizure model (Vogel, 2002). The animals were fed with normal laboratory pellet diet and water *ad libitum*. The approval was taken from Institutional Animal Ethics Committee of Akal College of Pharmacy and Technical Education, Mastuana Sahib, Sangrur before carrying out animal studies (ATRC/30/23, dated 26/12/2023).

Vehicle and standard drugs

Distilled water + Tween 80 (2%) was used as vehicle for preparing various test doses of crude extract and fraction in such a concentration as to administer a volume ranging 1-2 ml to the rats. Phenytoin sodium injection was used as a standard anticonvulsant drug at the dose of 20 mg/kg, *i.p.* Pentylenetetrazole was used to induce convulsions at the dose of 80 mg/kg, *i.p.*

Experimental protocol

Experimental protocol comprising groups 1-7 was designed to assess anticonvulsant activity of various crude extract / fractions of plant tubers.

Group 1 - Control group received pentylenetetrazole (80 mg/kg, i.p.).

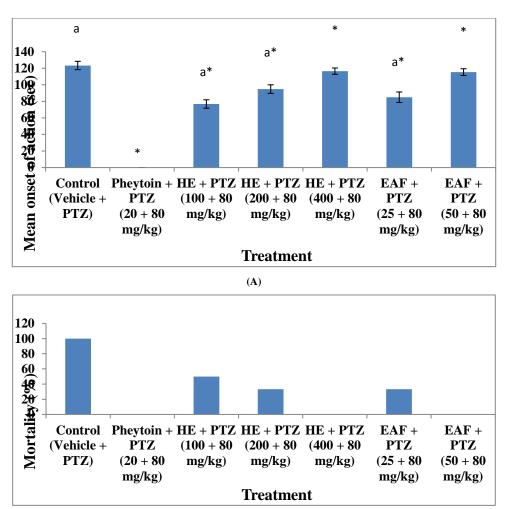
- Group 2 Standard group received phenytoin (20 mg/kg, i.p.) + pentylenetetrazole (80 mg/kg, i.p.).
- Groups 3 Test groups received 100 mg/kg doses of hydroalcoholic extract + pentylenetetrazole (80 mg/kg, *i.p.*).
- Groups 4 Test groups received 200 mg/kg doses of hydroalcoholic extract + pentylenetetrazole (80 mg/kg, i.p.).
- Groups 5 Test groups received 400 mg/kg doses of hydroalcoholic extract + pentylenetetrazole (80 mg/kg, i.p.).
- Groups 6 Test groups received 25 mg/kg doses of ethyl acetate fraction + pentylenetetrazole (80 mg/kg, i.p.).
- Groups 7 Test groups received 50 mg/kg doses of ethyl acetate fraction + pentylenetetrazole (80 mg/kg, i.p.).

Statistics

The results were expressed as mean \pm standard deviation (S.D.). The anticonvulsant activity of test drugs was compared with that of standard drug and control by one way analysis of variance (ANOVA) followed by Student-Newman-Keul's test (Scheffer, 1980).

RESULTS AND DISCUSSION

The ethyl acetate fraction (in relation to hydroalcoholic extract) and the hydroalcoholic extract's percentage yield (w/w) were found to be 5.90 and 20.80 % w/w respectively. The main purpose of extraction of plant material with n-hexnae is to remove various greasy materials from plant to avoid any interference for further extraction and pharmacological profiles. The hydroalcoholic extract of plant tubers showing presence of alkaloids, flavonoids, coumarins, proteins, tannins and carbohydrates. Further, the hydroalcoholic extract was purified to generate ethyl acetate fraction to remove (phytoconstituent rich fraction) to remove highly polar substances which are not responsible for pharmacological activities. The phytochemical testing of ethyl acetate fraction showing presence of alkaloids, flavonoids, coumarins and tannins. The hydroalcoholic extract and its ethyl acetate fraction of *Habenaria intermedia* tubers were investigated for anticonvulsant activity screening using well reported model in scientific databases named using pentylenetetrazole induced seizure model. The two parametres were accessed such as mean onset of action (sec) and mortality (%) in anticonvulsant activity screening and results are described in figure 1 following the administration of hydroalcoholic extract (100, 200, or 400 mg/kg, p.o.), ethyl acetate fraction (25 or 50 mg/kg, p.o.), diazepam (2 mg/kg, p.o.), and the control (vehicle, p.o.). The experimental protocol was divided into three groups such as control, standard and test. Each group of animals received pentylenetetrazole (80 mg/kg) of *Habenaria intermedia* tubers were exhibit anticonvulsant action of *Habenaria intermedia* tubers were exhibit anticonvulsant action in a dose dependent manner. The hydroalcoholic extract (400 mg/kg) and its ethyl acetate fraction (50 mg/kg) of *Habenaria intermedia* tubers were exhibit anticonvulsant action statistically equivalent to standard drug. Other doses hydroalcoholic extract (100 or 200 mg/kg) and its ethyl acetate fraction (25 mg/kg)



(B)

Figure 1: Anticonvulsant activity of *Habenaria intermedia* tubers using pentylenetetrazole induced seizure model. HE, Hydroalcoholic extract; EAF, Ethyl acetate fraction.

n=6; The data is expressed as Mean ± S.D.; *P<0.05 vs. Control; *P<0.05 vs. Standard; one way ANOVA followed by Student Newman Keul's test.

Conclusion

According to Kumar et al. (2014), bioactive extract and fractions of *Habenaria intermedia* tubers may have anticonvulsant properties by altering excitatory and inhibitory neurotransmission through effects on voltage-gated ion channels, GABA(A) receptors, and glutamate-mediated excitatory neurotransmission. Additionally, potentiating GABA neurotransmission may be achieved by inhibiting calcineurin (protein phosphatase 2B), reducing activation of NMDA extracsynaptic receptors, and preventing the activation of extracellular regulated kinase and $p38\alpha$ mitogen activated protein kinase pathways.

According to the literature that is currently accessible, coumarins, alkaloids, and flavonoids are essential for the treatment of neurological illnesses (Mittal et al., 2016). There are several plant reports wherein flavonoids have demonstrated central nervous system (CNS) activities. These include the flavonoids hesperidin (Marder et al., 2003), gossypin (Duraisami et al., 2009), diosmin (Fernandez et al., 2006), naringin (Fernandez et al., 2009), and quercetrin (Kang et al., 2010); alkaloids hypaconitine (Nesterova et al., 2010), montanine (Silva et al., 2006), neferine (Sugimoto et al., 2008), pseudo-akuammigine (Duwiejua et al., 2002), and yohimbine (Chermat et al., 1997); coumarins esculetin (Wang et al., 2012), imperatorin (Luszcki et al., 2009), osthole (Luszcki et al., 2009), and marmesinin (Kang and Kim 2007). Our findings imply that these naturally occurring groups of chemicals are responsible for the anticolvulsant activity of *Habenaria intermedia* tubers, which is consistent with recent observations.

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References

Ahuja PS. 2003. Medicinal plants in India: Report and Directory. New Delhi, India: Institute of Economic and Market Research, 65-89

Balkrishna A, Srivastava A, Mishra RK, Patel SP, Vashistha RK, Singh A, Saxena P. 2012. Astavarga plants-threatened medicinal herbs of the North-West Himalaya. *International Journal Medicinal and Aromatic Plants*. 2: 661-676.

Chermat R, Lachapelle F, Baumann N and Simon P, 1979. Anticonvulsant effect of yohimbine in quaking mice: antagonism by clondine and prazosine. Life Science, 25(17), 1471–1475.

Duraisamy S, Rasilingam D and Subramanian R, 2009. Anticonvulsant activity of bioflavonoid gossypin. Bangladesh Journal of Pharmacology, 4, 51-54.

Duwiejua M, Woode E and Obiri DD, 2002. Pseudo-akuammigine, an alkaloid from *Picralima nitida* seeds, has anti-inflammatory and analgesic actions in rats. Journal of Ethnopharmacology, 81(1), 73-79.

Farnsworth NR. (1966). Biological and phytochemical screening of plants. J Pharm Sci, 55, 225-76.

Fernandez SP, Nguyen M, Yow TT, Chu C, Johnston GAR, Hanrahan JR and Chebib M, 2009. The Flavonoid glycosides, myricitrin, gossypin and naringin exert anxiolytic action in mice. Neurochemical Research, 34, 1867-1875.

Fernandez SP, Wasowski C, Loscalzo LM, Granger RE, Johnston GAR, Paladini AC and Marder M, 2006. Central nervous system depressant action of flavonoid glycosides. European Journal of Pharmacology, 539, 168–176.

Goudar MA, Jayadevappa H, Mahadevan KM, Shastry RA, Habbu PV, Sayeswara HA. 2015. Isolation and characterization of secondary metabolites from *Habenaria intermedia* D. Don for evaluation of hepatoprotective activity against carbon tetrachloride induced liver damage in albino rats. *Asian Journal of Pharmaceutical and Clinical Research*. 8: 194-198.

Habbu PV, Smita DM, Mahadevan KM, Shastry RA, Biradar SM. 2012. Protective effect of *Habenaria intermedia* tubers against acute and chronic physical and psychological stress paradigms in rats. *Revista Brasileira de Farmacognosia*. 22: 568-579.

Jagetia GC, Rao SK, Baliga MS, Babu SK. 2004. The evaluation of nitric oxide scavenging activity of certain herbal formulations *in vitro*. A preliminary study. *Phytotherapy Research*. 18: 561-565.

Kang SY and Kim YC, 2007. Neuroprotective coumarins from the root of *Angelica gigas*: structure-activity relationships. <u>Archives of Pharmacal</u> Research, 30(11), 1368-1373.

Kang TH, Jeong SJ, Kim NY, Higuchi and Kim YC, 2010. Sedative activity of two flavonol glycosides isolated from the flowers of *Albizzia julibrissin* Durazz, Journal of Ethnopharmacology, 127(2), 551-554.

Khare CP. 2007. Indian Medicinal Plants: An Illustrated Dictionary. New York, USA: Spinger Science and Business Media, 233 Spring Street, 301.

Kirtikar KR, Basu BD. 1994. Indian Medicinal plants. Volume 4. New Delhi, India: Bishen Singh Mahendra Pal Singh, 2413.

Kumar D, Kumar S. Screening of antianxiety activity of *Abies pindrow* Royle aerial parts. Indian Journal of Pharmaceutical Education and Research, 2015, 49(1), 66-70.

Kumar, A., Lalitha, S., Mishra, J. 2014. Hesperidin potentiates the neuroprotective effects of diazepam and gabapentin against pentylenetetrazole-induced convulsions in mice - Possible behavioral, biochemical and mitochondrial alterations. *Indian Journal of Pharmacology*, 46, 309-315.

Luszczki JJ, Andres-Mach M, Cisowski W, Mazol I, Glowniak K and Czuczwar SJ, 2009. Osthole suppresses seizures in the mouse maximal electroshock seizure mode. European Journal of Pharmacology, 607(1–3),107–109.

Luszczki JJ, Wojda E, Andres-Mach M, Cisowski W, Glensk M, Glowniak K and Czuczwar SJ, 2009. Anticonvulsant and acute neurotoxic effects of imperatorin, osthole and valproate in the maximal electroshock seizure and chimney tests in mice: a comparative study. Epilepsy Research, 85(2-3), 293-299.

Marder M, Viola H, Wasowski C, Fernandez S, Medina JH and Paladini AC, 2003. 6-Methylapigenin and hesperidin: new valeriana flavonoids with activity on the CNS. Pharmacology, Biochemistry and Behavior, 75, 537–545.

Mittal P, Kumar D, Kumar S. Screening of anticonvulsant activity of *Viscum album* L. and estimation of hesperitin in plant using TLC densitometry. Indian Drugs, 2016, 53(7), 25-29.

Nesterova YV, Povetieva TN, Suslov NI, Semenov AA and Pushkarskey SV, 2010. Antidepressant activity of diterpene alkaloids of *Aconitum baicalense* Turcz. Bulletin of Experimental Biology and Medicine, 151(4), 425-428.

Prajapati NS, Purohit SS, Sharma AK, Kumar T. 2003. A Handbook of Medicinal Plants: A Complete Source Book. Jodhpur, India: Agrobios, 191-245.

Prakash O, **Kumar D**, Kumar S. Screening of methanol extract and ethyl acetate fraction of *Abies webbiana* Lindl. for neuropharmacological activities. Indian Journal of Pharmaceutical Sciences, 2015, 77(5), 536-541.

Rajashekhar I, Hiren R, Hardik D. 2015. A short review on Astavarga plants- losing their existence. *International Journal of Ayurveda and Pharma Research*, 3: 32-38.

Reynolds LD, Wilson NG. 1991. Scribes and Scholar. London: Oxford, 193-194.

Sahu MS, Sahu RA, Verma A. 2013. Immunomodulatory activity of alcoholic extract of Habenaria intermedia in mice. International Journal of Pharmacy and Pharmaceutical Sciences. 5: 406-409.

Scheffer WC. (1980). Statistics for the Biological Sciences. Philippines: Addison-Wesley Publishing Company, 121-41.

Silva AF, de Andrade JP, Bevilaqua LR, de Souza MM, Izquierdo I, Henriques AT and Zuanazzi JA, 2006. Anxiolytic-, antidepressant- and anticonvulsant-like effects of the alkaloid montanine isolated from *Hippeastrum vittatum*. Pharmacology, Biochemistry and Behavior, 85(1), 48-54.

Sugimoto Y, Furutani S, Itoh A, Tanahashi T, Nakajima H, Oshiro H, Sun S and Yamada J, 2008. Effects of extracts and neferine from the embryo of *Nelumbo nucifera* seeds on the central nervous system. Phytomedicine, 15(12), 1117-1124.

Vogel HG (2002): In: Drug Discovery and Evaluation: Pharmacological Assays. New York, Springer, pp. 487-488.

Wang C, Pei A and Chen J, 2012. A natural coumarin derivative esculetin offers neuroprotection on cerebral ischemia/ reperfusion injury in mice. Journal of Neurochemistry, 121(6), 1007–1013.

Warrier PK, Nambiar VPK, Thakur RS, Ramankuntty C. 1994. Indian Medicinal Plants: A Compendium of 500 Species. New Delhi, India: Orient Longan, 191.