



## **A Review on Cubosome: The Novel Drug Delivery System**

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### **Abstract-**

Because of the extraordinary advancements in biomedical nanotechnology over the last few decades, traditional drug delivery systems have been transformed into smart drug delivery systems that respond to stimuli. These well-defined nanoplateforms can boost therapeutic targeting efficiency while reducing the side effects/toxicities of payloads, which are vital variables for enhancing patient compliance by responding to specific internal or external triggers. Cubosomes are lipid-based nano systems that are analogous to familiar vesicular systems, such as lipo- and niosomes. They could be used as part of a exceptional drug delivery system that includes hydro-, lipophilic, and amphiphilic drug molecules. In this review, we critically analyze the relevant literature on cubosomes regarding theories of cubosome self-assembly, composition, and manufacturing methods, with an emphasis on tumor-targeted drug delivery applications. Due to the bioadhesive and compatible nature of cubosome dispersion, this review also focuses on a variety of drug delivery applications, including oral, ophthalmic and transdermal.

**Keywords-** Cubosome, Monoglycerides, Monoolein, Controlled release

### **Introduction [1-6]**

The Father of word “Cubosome” was Larsson since the structure resembles cubic molecular crystals, its internal structure was confirmed in 1967. Cubosomes having particle size ranging between 10-500 nm in diameter and square-shaped structure. Cubosomes are distinct, sub-micron, nanostructured specks of the bicontinuous cubic liquid crystalline phase. Cubosomes are nanoparticles which are self-assembled liquid crystalline particles of certain surfactants with proper ratio of water with microstructure. Cubosomes refer as “bicontinuous” because it is having two distinct large interfacial area (continuous, but non-intersecting) consisting hydrophilic regions separated by the bilayer. Cubosomes are composed of polymers, lipids and surfactants with polar and nonpolar constituents hence said as amphiphilic.

### **Structure [7-10]**

It consists of honeycombed (cavernous) structures splitting two internal aqueous channels and a large interfacial area. They contain indistinguishable microstructure as that of its parent with high surface area and their dispersions are less viscid than the parent cubic phase. Cubosomes were first recognized by using X-Ray scattering technique. These appear like square shaped or spherical dots consistent to the presence of pore with aqueous cubic phases in lipid-water system. Cubosomes are merely the liquid crystalline nanoparticles or nanostructure particles.

### **Advantage [3, 5, 7, 8]**

1. It is economic, biocompatible and non-toxic.
3. The preparation process is easy.
4. It has outstanding bio adhesive qualities.
5. They are thermodynamically stable for a longer period of time.
6. The ability to encapsulate compounds that are amphiphilic, hydrophobic, and hydrophilic.
7. Controlled and targeted release of bioactive substances.
8. High drug loading is caused by increased internal surface area and cubic crystalline formations.
9. These are excellent solubilizers when compared to lipid or non-lipid carriers.
10. The bioavailability range of water- soluble peptides would increase 20-100 times with these cuboidal systems.
11. Cubosomes serve as an excellent vehicle for protecting the sensitive drug from enzymatic degradation.

12. For sparingly soluble drugs cubosomes show high drug carrier capacity.
13. Increased convenience and compliance (orally, topically and intravenously).
14. Improved bioavailability due to size
15. Improved efficacy

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**Disadvantage** [5, 7, 10]

1. Entrapment of water-soluble drugs is less due to of the presence of a large amount of water exclusive cubosomes.
2. Due to high viscosity, large-scale production of cubosome is difficult.
3. Without a specific polymer, controlled drug delivery is not possible. They may lead to leakage during storage.

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**Material used in cubosomes** [8, 13]

Bicontinuous cubic phases are found in -

- 1) Natural lipids
- 2) Cationic and nonionic surfactants
- 3) Polymer system

- 1) Natural lipids

Though the lipid most commonly used to construct bicontinuous cubic phases are Monoglyceride and Monoolein.

- 2) Monoglycerides

Monoglycerides are freely form bicontinuous cubic phases upon the addition of water, are relatively insoluble are impervious to changes in temperature.

- 3) Monoolein

The main precursor of cubosome conformation is monoolein. Monoolein or glyceryl monooleate is a combination of the glycerides of oleic acid and other fatty acids, containing mainly of the monooleate. Commercially available monoolein may be obtained in two forms, a mixed glyceride form or as distilled monoolein the distilled monoolein is preferred for medical operations latterly of its high chastity. It swells in water, giving rise to several lyotropic liquid crystalline structures Monoolein is a nontoxic, ecological and biocompatible material classified as GRAS (generally recognized as safe) and it is included in the FDA inactive constituents companion and is Non - parenteral drugs licensed in the United Kingdom. Monoolein show the mesomorphic phase, important in making further accessible the implicit medicinal operation of the lipid.

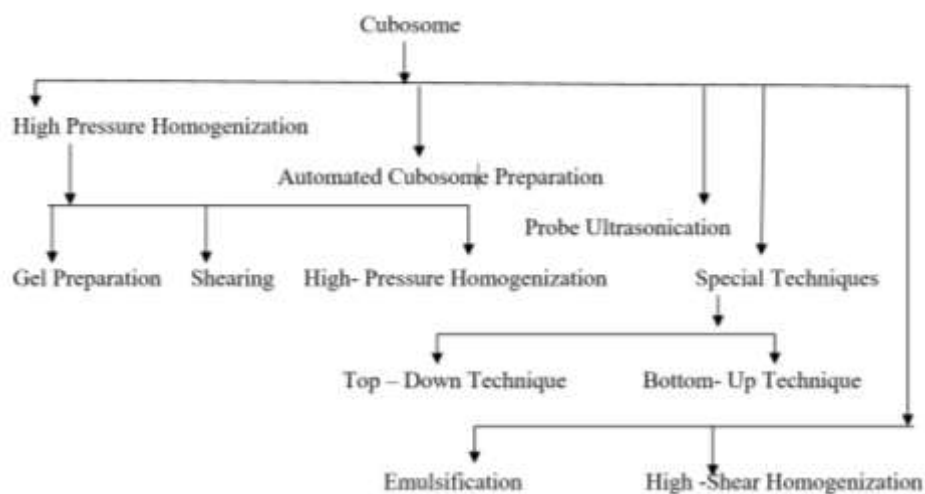
- 4) Surfactant

Surfactants which are used in the timber of cubosomes are poloxamer 407 in attention range between 0% and 20% w/w with respect to the disperse phase. The attention of the surfactant admixture typically takes between 2.5% and 10% w/w w.r.t. the total weight of the dissipation.

- 5) Polymer system

Polyvinyl alcohol (PVA) used in addition to poloxamer as a stabilizing agent of the dissipation.

## Method of Preparation [5]



## Evaluation [7, 8, 9, 10]

### 1. Visual examination:-

The cubosomes are visually assessed for optical appearance colour, turbidity, unity, presence of macroscopic patches.

### 2. Shape of the cubosome:-

Transmission electron microscopy can be used to view the shape of the cubosomes.

### 3. Flyspeck size distribution:-

Flyspeck size distributions of cubosomes are mainly determined by dynamic light scattering using Zeta seizer (Photon correlation spectroscopy). The sample adulterated with a suitable detergent is acclimated to light scattering intensity of about 300 Hz and measured at 25°C in triplet. The data can be collected and generally shown by using average volume weight size. The zeta eventuality and polydispersity indicator can also be recorded.

### 4. Zeta implicit:-

The magnitude of zeta eventuality indicates the degree of electronic aversion between acclimate, also charge flyspeck. Zeta eventuality is crucial index of the stability of formulation.

### 5. Entrapment efficiency:-

The ruse effectiveness of cubosomes can be determined using ultra filtration ways. In the later fashion, untrapped medicine attention is determined, which is abated from the total drug added. The quantum of medicine is anatomized by using spectrophotometer.

### 6. Dimension of drug release:-

Medicine release from cubosomes can be done by pressure ultrafiltration method. It's grounded on that proposed by Magenheim et al. using an Amicon pressure ultrafiltration cell fitted with a Millipore membrane at ambient temperature (22±2) °C.

### 7. Stability studies:-

The physical stability can be studied by disquisition of organoleptic and morphological aspects as a function of time. Flyspeck size distribution and drug content can be assessed at different time intervals can also be used to estimate the possible variations by time.

## Applications

1. Cubosomes as tumor targeted drug delivery
2. Skin cancer therapy
3. Glioblastoma Multiforme therapy
4. Lung cancer treatment

5. Colorectal cancer therapy
6. Liver cancer treatment
7. Ovarian cancer treatment
8. Cervical carcinoma
9. Hepatocellular carcinoma therapy
10. Brain tumor therapy

### Drug embedded in cubosomes

Drugs	Objectives of study	Outcome of study	Ref.no.
<b>Docetaxel</b>	Synthesis and evaluation of controlled release of cubosomes incorporated with docetaxel as thermo-sensitive depot.	The depot offered gradual drug release , preparation was free flowing at room temperature and changed to the depot at body temperature	16
<b>Antimicrobial Peptide</b>	The antimicrobial potential of cubosomal LL-37 was evaluated using in vitro adenovirus	The formulation provides superior protection to LL-37 against enzymatic degradation and significant bacterial effects and ensures a controlled release.	15
<b>Ketorolac</b>	Monoolein and poloxmer cubic nanoparticles for ocular delivery of ketorolac	Optimized cubosomes loaded with ketorolac provided transcorneal permeation and retention	17
<b>Indomethacin</b>	Evaluation of Indomethacin fabricated cubosome for anti-inflammatory activity	Homogenized monoolein and poloxamer containing cubosomes prolonged the delivery of lipophilic drug through the skin.	14
<b>Flurbiprofen</b>	NSAID used for treatment of ocular inflammation.	The formulation expressed less ocular irritation and enhanced trans-corneal permeation of FB	17
<b>Erythromycin</b>	Treatment and prevention of several types of acne as a result of its bacteriostatic activity against propionibacterium acnes.	The formulation prevents the acne due to the topical application of erythromycin impregnated with cubosomes	18
<b>Insulin</b>	Tested against the C-Type -1-diabetic induced rat	Cubosomes provide shield to insulin against proteolysis. It is found to be stable at normal temperature and controlled the hyperglycemia in a reproducible manner.	19

### References:-

#### 1. A NOVEL CORTICOSTEROID CUBOSOMES – FOR OCULAR DRUG DELIVERY

Snehal Shashaikant Chakorkar<sup>1</sup>, Jameel Ahmed S. Mulla

#### 2. A REVIEW ON: CUBOSOMES DRUG DELIVERY SYSTEM V. Ramya Sri, A. Madhusudhan reddy, R.Karthikeyan, P.Srinivasababu

3.CUBOSOMES: A NOVEL DRUG DELIVERY SYSTEM OVERVIEW Lakshmi Prasanna Yalavarthi \*, Pavan Kumar Jonnadula, Mohan Varma Manthina, Anand Addagalla Department of Pharmaceutics, Shri Vishnu College of Pharmacy, Bhimavaram, West Godavari District, Andhra Pradesh, India

4. review on cubosome: The novel drug delivery system Sadhu Venkateswara Rao <sup>1</sup> , Beram Naga Sravya <sup>1</sup> and Kantamneni Padmalatha Department of Pharmaceutics, Vijaya Institute of Pharmaceutical Sciences for Women, Enikepadu, Vijayawada–521108, India. Department of Pharmacology, Vijaya Institute of Pharmaceutical Sciences for Women, Enikepadu, Vijayawada–521108, India. Publication history: Received on 21 August 2018; revised on 28 September 2018; accepted on 05 October 2018

#### 5AN OVERVIEW ON THE STUDY OF CUBOSOMES –NANOPARTICLES

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#### 6 Cubosomes: As a Drug Delivery Carrier

Gupta Mukesh Department of Pharmaceutics, Alwar Pharmacy College, IET, North extension MIA, Alwar, (Raj.) India

#### 7 CUBOSOMES-A DRUG DELIVERY SYSTEM

Komal Tilekar\*, Prashant Khade, Sujit Kakade, Sachin Kotwal and RavindraPatil

#### 8. CUBOSOMES-A NOVEL DRUG DELIVERY SYSTEM

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9 .REVIEW ON CUBOSOMES AND THEIRNOVEL APPROACHES 1P. Shailaja, 2Y.Samukhya, 3J. Ramya, 4K. Satya Ashok, 1Associate professor, 2,3Student, 4Research scholar1Department of pharmaceutical technology1A.U. College of Pharm aceutical Sciences Andhra University, Visakhapatnam, India

#### 10. A REVIEW ON: CUBOSOMES DRUG DELIVERY SYSTEM

V. Ramya Sri, A. Madhusudhan reddy, R.Karthikeyan, P.Srinivasababu

#### 11. AN OVERVIEW ON THE STUDY OF CUBOSOMES –NANOPARTICLES

Arun T S\*1, Subash Chandran M.P1, Prasobh G.R1, Aparna P1, Remya S. B Department of Pharmaceutics, SreeKrishna College of Pharmacy and Research Centre, Parassala, Thiruvananthapuram, Kerala, India. 695502

12 FORMULATION AND INVITRO EVALUATION OF GASTRO RETENTIVE INSITU FLOATING GELS OF LOSARTAN POTASSIUM CUBOSOMES Dr.M.Sunitha Reddy\* and N.Nagadurga Centre for Pharmaceutical Sciences, IST, JNTUH Kukatpally, Hyderabad.

13.Cubosomes: a vehicle for delivery of various therapeutic agentsVolume 4 Issue 1 – 2018 Nilesh R Rarokar, Pramod B KhedekarDepartment of Pharmaceutical Sciences, Rashtrasant Tukadoji

14. Esposito E., Cortesi R., Drechsler M., Paccamiccio L., Mariani P., Contado C., Stellin E., Menegatti E., Bonina F., Puglia C. Cubosome dispersions as delivery systems for percutaneous administration of indomethacin. *Pharm. Res.* 2005;22:2163–2173. doi: 10.1007/s11095-005-8176-x. [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]

15. Boge L., Hallstenson K., Ringstad L., Johansson J., Andersson T., Davoudi M., Larsson P.T., Mahlapuu M., Håkansson J., Andersson M. Cubosomes for topical delivery of the antimicrobial peptide LL-37. *Eur. J. Pharm. Biopharm.* 2019;134:60–67. doi: 10.1016/j.ejpb.2018.11.009. [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]

16. Rarokar N.R., Saoji S.D., Raut N.A., Taksande J.B., Khedekar P.B., Dave V.S. Nanostructured cubosomes in a thermoresponsive depot system: An alternative approach for the controlled delivery of docetaxel. *AAPS PharmSciTech.* 2016;17:436–445. doi: 10.1208/s12249-015-0369-y. [[PMC free article](#)] [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]

17. Ali Z., Sharma P.K., Warsi M.H. Fabrication and evaluation of ketorolac loaded cubosome for ocular drug delivery. *J. Appl. Pharm. Sci.* 2016;6:204–208. doi: 10.7324/JAPS.2016.60930. [[CrossRef](#)] [[Google Scholar](#)]

18. Morsi N.M., Abdelbary G.A., Ahmed M.A. Silver sulfadiazine based cubosome hydrogels for topical treatment of burns: Development and in vitro/in vivo characterization. *Eur. J. Pharm. Biopharm.* 2014;86:178–189. doi: 10.1016/j.ejpb.2013.04.018. [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]

19. Boyd B.J., Khoo S.-M., Whittaker D.V., Davey G., Porter C.J. A lipid-based liquid crystalline matrix that provides sustained release and enhanced oral bioavailability for a model poorly water soluble drug in rats. *Int. J. Pharm.* 2007;340:52–60. doi: 10.1016/j.ijpharm.2007.03.020. [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]