

International Journal of Research Publication and Reviews

Journal homepage: www.ijrpr.com ISSN 2582-7421

Phytochemical and Antibacterial Screening: A Review on Leaves and Latex of *Calotropis Procera*

^aTripti Joshi, ^bNeetu Pandey, ^cAlok Maithani

Student, Sardar Bhagwan Singh University Balawala, Dehradun, 248001, Uttarakhand, India Assistant Professor, Sardar Bhagwan Singh University Balawala, Dehradun, 248001, Uttarakhand, India Associate Professor, Sardar Bhagwan Singh University Balawala, Dehradun, 248001, Uttarakhand, India

ABSTRACT

Various investigations exposed antibacterial activity and phytochemical screening on leaves and latex extract of *Calotropisprocera*. Over all methodused for study was first to isolate the extracts of petroleum ether, methanol, ethanol, ethanol-170%, ethyl acetate, chloroform and water and all the above extract were than evaluated for the phytochemical test and the antibacterial activity. The results shows that the extract of leaves and latex of *Calotropisprocera* shows remarkable antibacterial activity specially with five bacteria's namely *Salmonella typhyii, Escheria coli, S. aureus, S. flexerni, E. facalis* along with this the results also revealed that the best extraction solvent for antibacterial activity of leaf and latex extract is alcohol followed by chloroform and water.

Keywords: Antibacterial, Phytochemical, Screening, Evaluated

INTRODUCTION

India has been known for its use on herbal drugs since the ancient past. Looking back in times we could find the people are very much dependent on plant sources as they are beneficial. The home remedies and DIY has been the most common and favourite way of dealing with diseases for us Indian as we've practiced this since ages and now it has been incorporated in our genes to search for these ingredients in our kitchen or in the nature. Research has explored the nature of the secondary metabolites that in various medicinal plants. Medicinal plants shows a precious, renewable source for new drugs. The world health organisation has claimed that 80% of the world's population count on the herbal method of treatment of various diseases Around 500,000 plant spices were estimated but only a small amount has been investigated phytochemically. More than 130 drugs in the world's markets comes from higher plants either directly or synthetically [1], [2], [3], [4], [5]. Although hundreds of plants were tested for antifungal and antibacterial properties, the majority of them have not been adequately evaluated and processed well. [6]. The whole Himalayan belt of gods world is the home to several medicinal plants, as in India not only the mythological facts says so but always the science has given approval to this very fact. Himalayan Region with background information on family, habit and nativity. In India, out of 17000 species of the higher plants, 7500 are known for their medicinal uses. Currently 25% of the drugs are obtained from plants, many others are synthetic analogous built on the protype substances obtained from the plant. [7]

Asia and Africa, which is commonly known as Milkweed, Apple of Seldom and Madar. *Calotropis procera* is used as a folk medicine and is not a new name in Indian household as it is used as ornamental plant due to beautiful white flower. It has been reported that the plant possess potential antimicrobial, anthelmintic, ant inflammatory, anticancer, purgative, anticoagulant, analgesic, and antipyretic characteristics and is also used in the treatment of leucoderma, leprosy, liver and abdomen diseases [8]. The latex of *Calotropisprocera* has been known for important indigenous medicinal uses due to its laxative, antisyphilitic and analgesic action [9]. *Calotropisprocera* flowers causes temporary paralysis of red stomach worm in sheep and notably reduces egg count percent of gastrointestinal nematodes in naturally infected sheep [10]. Dry latex of *Calotropis procera* has potential anti-cancer properties due to its differentiable targets and non-interference with regular pathway of apoptosis [11]. The pharmacological properties of *Calotropisprocera* is a versatile plant for the pharmaceutical industry to develop new drugs [12]. Medicinal plants have no doubt remained the major sources of traditional medicine worldwide [13]. The main objective of this review is to study the effect of various solvent extracts obtained from the leaf and latex of *Calotropisprocera*, also to compare their qualitative and phytochemical screening by using standard tests because successful extraction, determination and isolation of biologically active components from plant material are largely dependent on the type of solvent used [14].

* Corresponding author. Tel.:9410314963; fax: +01352686286.

E-mail address: neetu_bhtt@yahoo.co.in

PHYTOCHEMICAL DISCUSSION OF PLANT

The whole plant contain is incorporated with loads of phyto constituent which possess different remedic effects, it is said to be used in digestion, diarrhoea, jaundice, boils etc. The plant was also recommended by some to treat leprosy, splenic enlargement, dropsy, hepatic and worms [15]

LATEX

The latex and fresh leaves are recommended to be applied on painful sour joints. The latex of *Calotropisprocera* is acidic with the density of 1.02g/ml the composition of the latex depends on the environment season [15]. The latex possess a rigid composition of with compounds namely calactin (1), cholin (2), calotropin (3), calotoxin (4), uscharine (5), syriogenin (6) uzarigenine (7), voruscharin (8), proceragenine (9). L-lactucerol, rpoceroid, tetraxasterol, β -amyrin, calotropeol 3-epimoretenol lupeol trypsin [16]. 3, 7, 11-Trimethyl 2, 6, 10, 12-pentadecatrien-1-ol, 2, 6 dimethyl tetra-1, 5-decaene and were also isolated from the latex [17].

The latex contains two distinct cysteine peptidase, procerain and procerain B. However, a new cysteine peptidases were purified from Calotropis procera latex. The purified enzymes exhibited plasma-clotting activity mediated by a thrombin-like mechanism [18].

It also possesses aspartic acid, glutamic acid, serine, glycine, histidine, arginine, threonine, alanine, proline, tyrosine, valine, methionine, isoleucine, phenylalanine and lysine [19]. Several studies have been made on the latex of *Calotropisprocera* are said to reportedly possess anti helmintic, antileprotic antirheumatism, anti-cold activity [20]

The Albino rats were evaluated with latex of *Calotropisprocera* for the protection against isoproutenol 20mg/100g induced for myocardial infarct. The prior treatment with the ethanolic extract of *Calotropisprocera* latex at a dose of 300 MG /KG body weight when given orally three times a day for consecutively 30 days reduce significantly the elevated makers enzyme level in serum and heart harmonized in isoproterenol in our induce my cardial infarction [21]

The latex of *Calotropicprocera* has been studied to possess anti diarrheal activity when studied against the rats it has been observed that the administration of single dose of dry Latex reduces the frequency of the reaction by a significant percentage when compare to the animals treated with castor oil.[22]

Dry latex of *Calotropis procera* has the potential for anti-cancer effect due to its differentiable targets and non-interference with regular pathway of apoptosis. Dry latex treatment of mice showed a complete protection against hepato carcinogenesis. No adverse effect was observed in these animals. [23] It has been observe that daily administration of oral dose of latex in Diabetic rats with the dosage of 100 and 400 mg/Kg decreases the Glucose level and consequently increases the hepatic glycogen level of the treated animals, it has also been reported that the restorement of the daily water consumption and body weight has seen. [24]

Inflammatory capabilities has also been observed by air pouch and pedel oedema models of inflammation, when the rats were treated with aqueous subcutaneous injection of dry latex into plantar surface of the paw induced inflkammation, the maximum inflammation was obtained for an hour and was maintained for the next hour, the response of the inflammation was determined by the increase in vascular permeability as it reached its Maxima in 15 minutes [25].

The latex has been observed for the antibacterial activity by the Agar diffusion method when streptomycin was taken as a positive control with a concentration of 2 mg/ ml. The methanolic extract was supposed to possess the maximum antibiotic effect [26].

CH







(4) Calotoxin

(5) Uscharine

(6) Syriogenin

CHO



Figures- 1-9, Compounds Isolated from Latex of Calotropis procera

LEAVES

The leaves of *Calotropis procera* possess cardiac glycosides, and sterols and terpenoides. Two major cardiac glycosides that has been isolated from the leaves are calotropin and calotropagenin, whereas terpenoids were mainly alpha-acetate (10), beta-amyrin, 3- thiozoline (11). The only sterol that is present in the leaf is Beta sitosterol (12) Ratinoside (13) Ursolic acid (15), alpha amyline (16), were also isolated from the leaves of Calotropis procera. The leaves of plant possess antihelmathetic, antirheumatism, cytotoxic, antiasthmatic, antioxidant activity. [27]

Anti helmethetic activity of the leaf has been valuated against Indian earthworm *Pheritomapostuma*, 70% hydro ethanolic extract in 12.5 mg/ ml concentration showed death and paralysis in 29.05 m and 18.58 m respectively. This effect depends totally on the concentration taken. [28] Cytotoxity against hepatic to has been observed by the leaf extract. [28]

Antioxidant potential of Calotropis procera's methanolic and aqueous extract of leaves by using scavenging activity of the stable (2, 2-diphenyl-1-picrylhydrazyl-hydrate) DPPH free radical. Half- maximal inhibitory concentration (IC50) of the methanolic extract was 110.25 mg/ml that represents that aqueous extract has strong antioxidant effect. [29]





MATERIAL AND METHODS

Preparation of plant extract - Generally the standard process which is used in the preparation of plant extract is as follows. Leaves of *Calotropisprocera* dried in shade in order to avoid direct contact of sunlight for this pulverization method is used. The dried leaves are macerated in the liquid i.e hexane, isopropyl, ethyl acetate, ethanol, methanol, acetone, for 48 hours. The latex is collected in sterile plastic/glass bottle by squeezing the apex and tips of leaves and kept in refrigerator at 4 °Celsius [30]. The latex dried under shade at ambient temperature to remove the chlorophyll content then take measured amount of latex and extracted with petroleum ether in the separating funnel after the formation of two separate layers of petroleum ether and residue the same is repeated with other solvent. [30]

Test on organism- Studies reveals that the antibacterial effect on the extract of leaves and latex were mainly performed on these bacteria's namely, Shigella boydii [31], Streptococcus pyogen, Streptococcus pneumoniae, Staphylococcus albus [32], Staphylococcus epidermis, Staphylococcus saprophylics, Shigella dysenbria, Plesiamonas shigelloides, Vibrio cholera, Shigella sonneria, Shigella flexneria, Pseudomonas aeruginosa [33], Escheria coli, Salmonella typhyii, S.aureus, S.flexerni, E.facalis [34].

Biochemical standards and photo chemical screening test were reported to use the following methods. [35], [36], [37], [38], [39], [40], [41], [42], [43], [44]

- 1. Test for carbohydrates: Molisch's test: Benedict's solution test
- 2. Test for alkaloids: Dragendorff's reagent: Mayer's reagent: Hager's reagent
- 3. Test for proteins and amino acids: Biuret test: Ninhydrin solution test
- 4. Tests for steroids: Liebermann-Burchard reaction: Salkowaski reaction
- 5. Test for flavonoids: Shinoda's test: Lead acetate solution test
- 6. Test for glycosides: Bontrager's test
- 7. Test for Phenols: Ferric chloride test
- 8. Test for Amino acid and protein: 1% ninhydrin solution in acetone test
- 9. Test for Saponins: Foam test
- 10. Test for Sterols: Liebermann- Burchard test
- 11. Test for Tannins: Braymer's test
- 12. Test for Terpernoids: Salkowki's test
- 13. Test for Quinones: A small amount of extract was treated with concentrated HCL and observed for the formation of yellow precipitate (or colouration) confirms the presence.

ANTIBACTERIAL TEST [45]

Antibacterial activity on hexane, petroleum ether, ethyl acetate, isopropyl alcohol, ethanol, chloroform, methanol, diethyl ether and aqueous extract of *Calotropis procera* studied over by the researchers and they mainly used disc diffusion method for the antibacterial determination.

In Disc diffusionmethod a petri-disc of 90 mm diameter is taken and media of MHA is prepared and the test organism is spread uniformly in the petridisc, now sterile disc are placed in the media infused with the extract that needs to be evaluated for antibacterial activity, negative control (pure solvent) and positive control (chloroemphicol) were taken as a reference to compare the result, this is placed inside incubator for 24 hours at 37 degree celsius temperature the zone of inhibition is measured with respect to a control.

Phytochemistry [46]

When the leaf extracts of acetone, petroleum ether, chloroform, ethyl acetate, ethanol, methanol, isopropanol, hexane, petroleum ether and water were analysed or tested for the presence of alkaloids, flavonoids, saponins, terpenes, tannins, stair instance glycoside, phenolic compounds, amino acid, carbohydrate, steroid, quinone it was found that the different extract shows positive test for the certain fight of constituent as shown in table-1

RESULTS

Solvent	%yiel	Alkaloid	Flavnoid	Saponin	Terpene	Tannin	glycosid	Phenoli	Aminoaci	Carbohydrat	Quinon
	d	s	s	s	s	s	e	c	d	e	e
Acetone	9.68	+	_	+	_	_	+	_	I	_	_
Diethylethe	3,01	-	-	++	_	+	_	_	-	-	-
r											
Chlorofor	3.6	+	-	++	++	+	-	-	-	++	-
m											
Ethylacetat	5.6	_	_	_	_	_	+++	+	_	_	_
e											
Ethanol	6.8	+++	-	+++	-	+	++	+	_	+	-
Methanol	8.9	+	+	++	_	+	+	+	+	+	+
Isopropano l	2.5	+	+	+	+	+	+	-	-	_	_
N hexane	1.2	_	+	+	_	-	++	_	_	_	_
Pet.Ether	0.76	_	_	_	+	_	+	_	+	_	+
Water	6.0	+	-	-	-	+	+	+	-	+	-

Table-1 Phytochemical Screening of Calotropis procera leaf extract [46, 47]

+ = Present

- = Absent

Ingredient	Ethanol	Aqueous
Reducing sugar	+	_
Tannins	+	+
Steroid glycoside	+	+
Resin	_	+
Alkaliods	_	_
Saponins	+	+
Flavonoids	+	+

Table-2 Phytochemical screening of latex of *Calotropis procera* [48]

Table-3 Inhibition diameter [48]

Plant extract	S.aureus		S.flexerni		E.coli		E.facalis		S.typhii	
	Leaf	Latex	Leaf	Latex	Leaf	Latex	Leaf	Latex	Leaf	Latex
Methanol	12.5±0.5	11±0.2	-ve	10.01	-ve	-ve	10±0.1	-ve	8±0.6	7±0.6
Ethanol	11.2±0.7	11±0	24±0.5	-ve	11±0.5	7±0.1	-ve	-ve	16±0.4	-ve
Ethanol70%	11±0.1	13±0.1	-ve	12±0.1	9±0.1	12±0.4	-ve	9±0.4	13±0.0	15±0.1
Water	11±0.2	12.05±0.5	-ve	9±0.3	10±0.1	7±0.6	10±0.1	_ve	8±0.6	7±0.6
Chloroform	-ve	10±0.0	8±0.5	-ve	9±0.5	9±0.4	-ve	-ve	9.1±0.1	-ve
Chloroamphicinol	14±0.3		17.3±0.1		13±0.06		15±0.1		11±0.4	

ANTIBACTERIAL EFFICACY [49], [50]

The zone of inhibition diameter varies for different extract as antibacterial effect of the plant *Calotropis procera* varies for different microbes. Pure solvent are used as negative control. S.aureus was the bacteria that was very sensitive to every latex and leave extract to the plant as can be concluded by the data. *Calotropis procera* for ethanol extract of leaves and 70% ethanol extract and leave extract for E coli (17mm and 15 mm respectively) with MIC of 20 mg/ml.E.facalis is resistant to ethanolic extract, while S. flexerni, S.typhii, E.coli were sensitive to some of the extracts mention above. Ethanol extract of leaves and ethanol 70% extract of leaves on salmone in a typhoid 1615 and 13 mm with mic of 20 and 50 mg/mm respectively and E coli (11 12 and 9 mm) with MIC of 20 and 40 mg/ml respectively

Only ethanol and 70% ethanolic extract of Calotropisprocera recorded and zone of inhibition than chloroamphicinol in S. typhyii.

Discussion/Conclusion

Drug resistant antibiotic has increased alarmly all over the globe as they are becoming as they are being used indiscriminately. Nature has been a supplier of medicines and medicinal agents for years and an impressive number of modern drugs have been isolated from natural sources [45]. In vitro and bacterial essay has been reported as the first step toward the development of novel therapeutic agents studies have been recorded for the antibacterial activities of aqueous and other extract of Calotropis procera against human pathogen it has been seen in the earlier studies that the extract of ethanol shoes maximum zone of inhibition because ethanol itself possese some antibacterial activity, some work has also reported that methanolic extract shows greater Zone of inhibition then hexane and ethylacetate.

REFERENCES

- 1. Borris RP, Natural products research: Perspectives from a major pharmaceutical company, J Ethnopharmacol 51 1996:29–38.
- Kong JM, Goh NK, Chia LS and Chia TF, Recent advances in traditional plant drugs and orchids, Acta Pharmacol Sin 24, 7–21. N. Morsy et al. / Phytochemical analysis of Calotropis procera 2003 :273
- 3. Rates SMK.Plants as source of drugs, Toxicon 39 2001:603-613.
- 4. Gurib-Fakim A. Medicinal plants: Traditions of yesterday and drugs of tomorrow, Mol Aspects Med 27 2006:1–93.
- 5. Newman DJ, Cragg GM and Snader KM. The influence of natural products upon drug discovery, Nat Prod Res 17 2000:215-234.
- 6. Balandrin MF, Klocke JA, Wurtele ES and Bollinger WH. Natural plant chemicals: Sources of industrial and medicinal materials, Science 228 1985:1154–1160
- 7. Jain SC, Sharma R, Jain R and Sharma RA. Antimicrobial activity of Calotropisprocera, Fitoterapia 67 1996:275–277.
- 8. Derek V, Banthrope, John JW. Phytochemistry 1995; 38(1):107-111.
- Iqbal Z, Lateef M, Jabbar A, Muhammad G and Khan MN. Anthelmintic activity of Calotropis procera (Ait.) Ait. F. flowers in sheep, J Ethnopharmacol 2005; 102:256–277.
- 10. Larhsini M, Bousaid M, Lazrek HB, Jana M and Amarouch H, Evaluation of antifungal and molluscicidal properties of extracts of Calotropis procera, Fitoterapia 1997;68: 371–373.
- 11. Choedon T ,Mathan G,Arya S, Kumar VL and Kumar V. Anticancer and cytotoxic properties of the latex ofCalotropis procera in a transgenic mousemodel of hepatocellular carcinoma, World J Gastroenterol 2006;12: 2517–2522.
- Silva MCC, da Silva AC, Teixeira FM, de Sousa PCP, Rondon RMM and. Junior JERH. Therapeutic and 'biological activities of Calotropis procera (Ait.) R, Br Asian Pac J Trop Med 2010; 3: 332–336.
- 13. Goyal, M. and Mathur, R. Antimicrobial potential and Phytochemical analysis of plant extracts of Calotropis procera. Int. J. of Drug

Discovery and Herbal Research, 2001; 1(3):138-143.

- 14. Tiwari P, Kumar B, Kaur M, Kaur G, Kaur H. Int. Pharm. Sciencia, 2011;1:98-106.
- 15. Al-Snafi AE, Department of Pharmacology, College of Medicine, Thi qar University, Nasiriyah,
- Shukla OP, Krishnamurti CR. Properties and partial purification of bacteriolytic enzyme from the latex of Calotropis procera (Madar). Journel
 of Science industrial Research, 1961; 20:109-112.
- Misra MK, Mohanty MK, Kdas P. Studies on the method-ethnobotany of Calotropis gigantean and C. procera. Ancient Science of Life, XIII (1&2)1993: 40-56.
- 18. Doshi1 HV, Parabia FM, Sheth FK, Kothari IL, Parabia MH, Ray A. Phytochemical analysis revealing the presence of two new compounds from the latex of *CalotropisProcera* (Ait.) R.Br. International Journal of Plant Research, 2012; 2:28-30.
- Ramos MV, Bandeira GP, de Freitas CDT, Nogueira NA, Alencar NMN, de Sousa PAS, Carvalho AFU. Latex constituents from Calotropis procera (R. Br.) Display toxicity upon egg hatching and larvae of Aedes aegypti (Linn.). Mem Inst Oswaldo Cruz, Rio de Janeiro, 2006; 101 (5): 503-510.
- 20. Maikhuri A, Parcha V, Fatma N, Kumar D, Karn SK. Phytopharmacoglogical reciew of *Calotropisprocera* Department of applied science, Sardar Bhagwab Singh University Balawal Dehradun 2020
- Kumar VL, Padhy BM, Sehgal R, Roy S. Antioxidant and protective effect of latex of *Calotropis procera* against alloxan-induced diabetes in rats. Journal of Ethnopharmacology, 2005; 102 (3): 470-473.
- 22. Kumar VL,Samgraula H, Dewan S,Kumar S.Antidiarreaheal activity of the latex of Calotropis procera Journal of Ethnopharmacology,2001;76(1):115-118,
- Magalh HIF, Ferreira PMP, Moura ES, Torres M, Alves ANN, Pessoa ODL, Lotufo LC. In vitro and in vivo antiproliferative activity of *Calotropis procera* stem extracts. Anais da Academia Brasileira de Ciências, 2010; 82(2): 407-416.
- Kumar VL, Padhy BM, Sehgal R, Roy S. Antioxidant and protective effect of latex of Calotropis procera against alloxan-induced diabetes in rats. Journal of Ethnopharmacology, 2005; 102(3):470-473.
- Al-YahyaMA Al-Meshal IA, Mossa, JS, and Tariq M.. Phytochemical and pharmacological studies on *Calotropis procera*. Proceeding of the 3rd International Conference of Traditional and Folk Medicine. Lecatecas, Mexico, 1985.
- Uddin, G., Rauf, A., Muhammad, N. and Mulik, N. 2012. Phytochemical and pharmacological studied of the whole plant of *Calotropis* procera. Middle-East Journal of medicinal Plants Research.2012; 1(4): 71-74.
- Yasmin, N., Uddin, S.N., Mubassara, S. and Akond, M.A. 2008. Antioxidant and antibacterial activity of *Calotropis procera* linn. Ammerican -EurasianJournal of agricultural and environmental science.2008; 4(5): 550-553.
- 28. Brain KR, Tuner TD, The practical evaluation of phytopharmaceuticals, Wright Scientectica Publishers: Bristol, 1975; 57–58.
- Freitas CD, Lopes JL, Beltramini LM, de Oliveira RS, Oliveira JT,Ramos MV. Osmotin from Calotropis procera latex: new insights into structure and antifungal properties. Biochim Biophys Acta, (2011; 8(10): 2501-2507.
- Yasmin, N., Uddin, S.N., Mubassara, S. and Akond, M.A. 2008. Antioxidant and antibacterial activity of calotropis procera linn. Ammerican -EurasianJournal of agricultural and environmental science. 2008; 4(5): 550-553.
- Verastegui MA, Sanchez CA, Heredia NL, Santos GAJ (1996) Antimicrobial activity of ectracts of three major plants from the Chihuahuan desert J. Ethnopharmacol. 52:175-177.
- 32. Mst Nazma Yesmin, Sardar Nasir Uddin, Sanzida Mubassara, Muhammed Ali Akand. Department of Botany and Genetic Engineering Displine American – Eurasian Journal of Agriculture and Enivronmental Science. 2008;4(5):550-553.
- 33. A Kareem SO, AkpanI,Ojo OP, Department of Microbiology. African Journal of Biomedical Research. 2008;(11):105-110.
- 34. Mali Rohit P, Rao Priya S, Jadhav RS. Department of Pharmacogonosy. Journal of Drug Delivery and Therapeutics. 2019; 9(3-5):947-951.
- 35. Kawo AH, Mustapha A, abdullahi BA, Rogo LD, Gaiya ZA, Kumurya AS. Bayero Journal of Pure and Applied Science. 2009; 2(1):34-40.
- Doshi H, Satodiya H, Thakur MC and Parabia F, Khan A. Phytochemical Screening and Biological Activity of Calotropis procera (Ait). R. Br. (Asclepiadaceae) Against Selected Bacteria and Anopheles stephansi Larvae, Int J Plant Res 1 (2011) 29–33.
- Mossa JS, Tariq M, Moshin A, Angeel AM, Al-Yahya MA, Al-Said MS and Rafatullah S. Pharmacological studies on aerial parts of Calotropis procera, Am J Chin Mesd 3 (1991), 223–231.
- Ramaprabha M, and VasanthaK. Phytochemical and antibacterial activity of Calotropis procera (Ait.) R.Br. flowers. Int J of Pharma and Biosci 3 (2012), 1–6.
- Mainsara MM, Aliero BL, Aliero AA and YakubuM. Phytochemical and Antibacterial properties of root and leaf extracts of Calotropis procera, NJBAS 2012; 20: 1–6.
- 40. A. Mohsin, A.H. Shah, M.A. Alaha, M.O. Tariqi and A.M. Ageel, Analytic anti-pyretic activity and phytochemical
- 41. N.R. Farnsworth, Biological and phytochemical screening of plants, J pharm Sci 55 (1996), 225-276.
- 42. J.B. Harborne, Phytochemical Methods, A guide to modern techniques of plant analysis, Chapman and Hall: New York, 1998; 1-150.
- 43. A. Sofowora, Medicinal plants and traditional medicine in Africa, Spectrum books limited: Ibadan, Nigeria, 1993; 150-289.
- 44. K.R. Brain, T.D. Tuner, The practical evaluation of phytopharmaceuticals, Wright Scientectica Publishers: Bristol, 1975; 57-58.
- 45. Trease GE, and Evans WC. A textbook of pharmacognosy, Academic press: London, (1989). 22-40.
- 46. Jorgensen JH, Turnidge JD and Washington JA.Antibacterial susceptibility tests: Dilution and disk diffusion methods. In: Manual of clinical microbiology, Washington, DC: ASM Press, 1999;7: 1526–1543
- 47. Abegunde S. M.1, Ayodele-Oduola R.O.2 1, 2 Department of Science Technology, The Federal Polytechnic, P.M.B. 5351, Ado Ekiti, Nigeria.
- 48. Raginee Verma", G. P. Satsangi, and J. N. Shrivastava Microbiology Laboratory, Department of Botany, Faculty of Science, Dayalbagh Educational Institute, Dayalbagh, Agra.
- 49. SalemWM, Sayed W F HaridyM. and HassanNH.Department of Botany, Faculty of Science, South Valley University, Qena, Egypt Department of Pathology and Clinical Pathology, Faculty of Veterinary Medicine, South Valley University, Qena, Egypt
- Sofwara A. Medicinal Plants and Traditional Medicines in Africa. Spectrum Books Ltd., Ibadan, Nigeria, 1993: 289.