



Examining Antifungal Resistance in Candida Isolates from Hospitalized Patients

Mr.Kuldeep Singh¹, Dr.Madhurendra Rajput², Mr.Shyam Sundar Bera³, Dr.Khyati Jain⁴, Mr Rajdeep Paul⁵

¹Research Scholar ,Malwanchal University, Indore

²Research Supervisor, Malwanchal University, Indore.

³Research Scholar, Malwanchal University, Indore.

⁴ Professor, People's Medical College, Bhopal.

⁵Research Scholar Malwanchal University, Indore

Introduction

Candida albicans is not the most prevalent kind of yeast in the Western world, despite popular belief. Non-*Albicans* species of *Candida* might develop if conditions favour more robust strains like *Candida glabrata* and *C. krusei*. Contrary to its more susceptible relative, *Candida glabrata*, *Candida krusei* is naturally resistant to the antifungal drug fluconazole. *C. tropicalis* is more challenging to treat with antifungal medications because it adheres to nonliving things like urine and vascular catheters. Resistance to azoles is increasing in both *C. tropicalis* and *C. albicans*. Because of this, determining fluconazole susceptibility and guiding early treatment decisions necessitates the use of fast speciation techniques. This is why we set out to identify the risk variables related to *Candida* species isolation from the urine of hospitalised patients and their sensitivity to antifungal therapy. The quick speciation technique was compared to conventional sugar fermentation processes. vaginal tract glycogen buildup, which may encourage the growth of *Candida*. *Candida* and yeast cytosol cells have been demonstrated to increase adhesion to vaginal epithelial cells through receptors or mechanisms for reproductive hormones. Many people enduring long-term catheterization have no symptoms, making it impossible for them to communicate their increasing frequency or dysuria. Individuals between the ages of 30 and 35, as well as those between the ages of 40 and 45, had the fewest symptoms overall. A total of 60 patients were found to have asymptomatic candidiasis; 25 were treated with antibiotics, and another 25 were catheterized as a precaution. In the 2018 research by Mauricio M et al., only 18% of participants had symptoms of a UTI. In this study, 27% of participants reported experiencing symptoms related to candiduria. In men, UTI symptoms were more common beyond the age of 45. Age-related increases in risk factors may be a contributing factor. Research shows that 29% of patients needed a more extensive catheterization due to unconsciousness. As far as symptoms went, they were all over the place. There were around 20 critically ill patients across three departments: eight in neurosurgery and 12 in medicine. Catheterization and prolonged antibiotic treatment were associated with this patient group. Fever was the major symptom in 18% of cases, and dysuria was the primary symptom in the same percentage of cases. Fevers were present in 19% of patients with catheterized UTIs, while dysuria affected 10%. Only patients with candiduria participated in the A. Paul et al. study. Urinary tract infections produced by contaminated urine may induce dysuria and urgency in non-catheterized people, although catheterization prevents these infections. If you have an irritable bladder or vesicoureteral reflux, a patent urinary catheter may help alleviate the pressure that builds up in your urinary tract. Pyelonephritis affected six persons, cystitis affected nine, and a total of twenty patients were found to have a urinary tract infection (UTI).

Our study population was much smaller than others of its kind in nephrology. The research on P. Stor was a major driving force for our project. There is a unique combination of dangers in each ward. Patients admitted to the medical floor were more likely to have preexisting conditions including diabetes, renal illness, or a UTI. The most common risk factors for patients in the ICU and post-operative patients were catheterization and the use of antibiotics. Over eighty-six percent of patients took antibiotics, and almost as many used catheters. Uma Chaudary et al. included critically ill people with candiduria instead of the 98.1% of our patients who had catheterization as a risk factor. Antibiotics alter the microbiota of the genito-urinary tract, making it more conducive to the growth of *Candida* species. Catheters can encourage the colonisation of their surfaces by *Candida* species other than *C. albicans*. Antibiotic use has been shown to increase the risk of nonalbican *Candida* spp. in a statistical study by 0.732%. The second most common condition was chronic kidney disease (38%), followed by type 2 diabetes (43%). The prevalence of diabetes identified in this study (22.1%) is higher than the prevalence found by Clark CB et al. (23.1%). The prevalence of diabetes is said to be highest in India. Indians are seeing an increase in the prevalence of metabolic syndrome, which includes insulin resistance, abdominal obesity, low levels of adiponectin, and high levels of C-reactive protein. The number of persons with diabetes is projected to increase from 42.0 million in 2017 to 72.3 million in 2025, as stated in the Diabetes Atlas 2021 published by the International Diabetes Federation. Some examples are oral and intravenous steroid treatment, tacrolimus, and mycophenolate mofetil. Similar medications were used to treat autoimmune diseases such as SLE and RPGN. Additionally, most patients took antibiotics and catheters for extended durations, making them more susceptible. Using the Chi-Square test, we found that having a transplant increased the likelihood that you would get candiduria due to *C. tropicalis* by 0.004.

According to some estimates, chronic kidney disease (CKD) is responsible for 36% of all UTIs. Renal insufficiency was found in just 18% of patients in Stephen P. S.'s study, which is on par with the proportion found by Mery S. et al. One possible explanation is that fifty out of every hundred patients also had diabetes. Patients with these characteristics are very susceptible to colonisation because of their glycosuria and deficiency of phagocytes. Patients on hemodialysis or with uremia are at a higher risk of contracting an infection than those without CKD. Metabolic acidosis, low albumin levels, iron overload, underlying disease, and immune system dysfunction all play a role. *C. albicans* and nonalbicans were not statistically distinct from one another. Patients with chronic renal illness and *Candida* species ($p = 0.242$) One-sixth of the people studied had candidiasis due to factors unrelated to the kidneys. These factors included calculi, BPH, neurogenic bladder, RPGN, hypospadias, phimosis, and prostate cancer. Catheterization was used to collect 78 percent of the urine samples, whereas midstream collection was used for just 29 percent. The investigations by Claudio CB, Artiaga K et al., and Arlene O.C. et al. all found the same thing: 81.1% of patients needed a catheter. Only 16% of the isolates were found to be *C. albicans*, whereas 85% belonged to other *Candida* species. *Candida* species other than *C. albicans* were detected in 71% of urine isolates analysed by Manisha Jain et al. *C. tropicalis* accounted for 63.33 percent of the population, *C. albicans* for 15.11 percent, *C. guilliermondii* and *C. krusei* for 9.3 percent each, *C. parapsilosis* for 6.8 percent, and *C. kefyr* for 7.23 percent. Similar findings to those of Masak J et al. It was shown by her research that 52.3% of the isolates were *C. tropicalis* and 24.8% were *C. albicans*. Febre N et al. and Elza H.D. Silva et al. found that *C. albicans* was the most common species, making up 45–49% of the total. In the study, more uncommon *Candida* species than *Candida albicans* were discovered to be on the rise. More than 97% (97/99) of the urine samples analysed contained just one organism. The findings matched those of Agarwal. Plan a certain time and day. *C. krusei* was the most common type of *Candida* found in mixed isolates, with *C. tropicalis* coming in second. Since *C. krusei* is hardier, it has probably displaced the native species. *Candida* spp. were present in both catheterized and uncatheterized urine. This was also true for individuals who were catheterized and who had a *Candida* species other than *Albicans* (0.005). Notably, fungi are responsible for 28.3% of all infections caused by catheters. Catheter biofilm production by *C. tropicalis* may aid in the colonisation of catheter users who use urological draining catheters. *C. tropicalis* biofilms were more resistant to antifungal medications due to their wide, hexamine-rich matrix, whereas *C. albicans* biofilms were more sensitive due to their narrow, glucose-rich matrix. The causes of the spread of *Candida* species other than *C. albicans* are not well understood. Patients who were catheterized had a lower prevalence (32.8%) than those who were sampled in the middle of the flow (7.9%). Catheterized patients had a higher prevalence of *C. albicans* in midstream urine samples compared to noncatheterized patients. Assimilation was the most effective technique for distinguishing the species of *Candida*, correctly identifying 97.6 percent of these organisms. The percentage of correct species identifications made using Hi-Chrom agar ranged from 82.4% to 97.1%. However, the sensitivity of Hi-Chrom agar varied by species, and while this was significant ($p = 0.0301$), this made it difficult to positively identify isolates. A sensitivity of 98 percent was found in all three species of *C. parapsilosis*, with the exception of *C. tropicalis* (82.4%) and *C. guilliermondii* (82.1%). This test was 83.2% as sensitive as Chromagar. According to research by Chaudary et al., the sensitivity of different species of *Candida* to Hi-Chrom agar ranges from 88% to 100%. Hi-Chrom agar was more sensitive in our study than in Baradhkar VP et al.'s, which may be due to the huge number of *C. tropicalis* isolates we examined. *C. albicans* and *C. tropicalis* both showed 96.1% and 96.2% specificity on Hi-Chrom Agar, respectively. *Clostridium albicans* had a sensitivity of 98.2 percent, *Clostridium tropicalis* had a sensitivity of 93.1 percent, and *Clostridium parapsilosis* had a sensitivity of 88.0 percent. Our findings vary from those of other research because we analysed a much larger collection of *C. parapsilosis* isolates. Green colonies may only be used to identify *C. albicans* 68% of the time, even when the fungus is present. Steel Blue successfully detected *C. tropicalis* 95.1% of the time. *C. parapsilosis*, *C. krusei*, and *C. guilliermondii* could all be distinguished from one another by their distinctive cream, dry pink, and light pink to purple colony colours. In this investigation, 23.1% of *C. tropicalis* isolates and 19.1% of *C. albicans* isolates were shown to be resistant to fluconazole. This is why Ariane Bruder-Nascimento and coworkers found that 18.3% of *C. tropicalis* and 25% of *C. albicans* strains were resistant. *C. albicans*, however, was shown to be very susceptible by Ariane BN et al. Due to the strains' minor variations, only 8.9% of SDD isolates demonstrated sensitivity or insensitivity to DD or MBD tests. Researchers Barry A. L. et al. found an error rate of 7.2%. There was a low level of itraconazole sensitivity among MBD and DD isolates (77.1 and 72.3%, respectively). These two species showed much greater resistance to itraconazole than the others. Despite the low sensitivity of their samples (74.1%), Riane B.N. et al. also discovered the same thing. There was a 13.5 percent risk of a moderate error and a 2.8 percent possibility of a catastrophic error because of the mismatch between the MBD and DD. The MBD strategy was more successful with itraconazole. While MBD revealed 100% sensitivity, DD only discovered 87.5% responsiveness across the isolates. Without MBD, susceptibility testing for amphotericin B is useless. Colleagues of Ariane BN also saw the incident.

REFERENCES

1. Achkar JM, Fries BC. Candidal infections of the genitourinary tract. *Clin Microbiol Rev.* 2010;23:253–273. doi: 10.1128/CMR.00076-09.
2. Sardi JCO, Scorzoni L, Bernardi T, Fusco-Almeida AM, Mendes Giannini MJS. *Candida* species: current epidemiology, pathogenicity, biofilm formation, natural antifungal products and new therapeutic options. *J Med Microbiol.* 2013;62:10–24. doi: 10.1099/jmm.0.045054-0.
3. Gullo A. Invasive fungal infections: the challenge continues. *Drugs.* 2009;69(Suppl 1):65–73. doi: 10.2165/11315530-000000000-00000.
4. Yang YL, Cheng HH, Ho YA, Hsiao CF, Lo HJ. Fluconazole resistance rate of *Candida* species from different regions and hospital types in Taiwan. *J Microbiol Immunol Infect.* 2003;36:187–191
5. Cannon RD, Lamping E, Holmes AR, Niimi K, Baret PV, Keniya MV, Tanabe K, Niimi M, Goffeau A, Monk BC. Efflux mediated antifungal drug resistance. *Clin Microbiol Rev.* 2009;22:291–321. doi: 10.1128/CMR.00051-08.
6. White TC, Marr KA, Bowden RA. Clinical, cellular, and molecular factors that contribute to antifungal drug resistance. *Clin Microbiol Rev.* 1998;11:382–402.

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7. Hospenthal DR, Beckius ML, Floyd KL, Horvath LL, Murray CK. Presumptive identification of *Candida* species other than *C. albicans*, *C. krusei*, and *C. tropicalis* with the chromogenic medium CHROMagar *Candida*. *Ann Clin MicrobiolAntimicrob*. 2006;5:1. doi: 10.1186/1476-0711-5-1.
 8. Isenberg HD. Mycology and Antifungal Susceptibility Testing. In: Gracia LS, Isenberg HD, editors. *Clinical microbiology procedure handbook*. 2. Washington, DC: ASM Press; 2004. pp. 8.0.1–8.10.7.
 9. Kauffman C, Fisher J. *Candida* urinary tract infections: diagnosis. *Clin Infect Dis*. 2011;52(suppl 6):S452–S456. doi: 10.1093/cid/cir111.
 10. Yucesoy M, Esen N, Yulung N. Use of chromogenic agar for the identification of *Candida albicans* strains. *Kobe J Med Sci*. 2001;47:161–167.