



A Review on Chemical Nature and Pharmacological actions of Multifunctional Osthole

Barate S.A.¹, Deshmukh S.², Wasmate D.³, Bawage S.⁴

¹Department of pharmaceutical chemistry, Latur college of pharmacy, Hasegaon, Tq. Ausa, Dist. Latur. 413512.

²Department of pharmaceutical chemistry, Latur college of pharmacy, Hasegaon, Tq. Ausa, Dist. Latur. 413512.

³Department of pharmaceutical analysis, Latur college of pharmacy, Hasegaon, Tq. Ausa, Dist. Latur. 413512.

⁴Department of pharmacognosy, Latur college of pharmacy, Hasegaon, Tq. Ausa, Dist. Latur. 413512.

ABSTRACT-

Osthole, better known as osthol, comes from the coumarin found in *Cnidium monnieri* and *Angelica pubescens*, among other medicinal plants. It has been reported that osthole from *Cnidium monnieri* (L.) Cusson can inhibit plant pathogen. It can be extracted and separated from plants or fully integrated. Several studies have found that osthole has anti-cancer, anti-inflammatory, neuroprotective, osteogenic, cardiovascular protective, antibacterial, and antiparasitic properties. In addition, some research has been done on the efficiency and effectiveness of osthole. This article covers a comprehensive list of Osthole sources and the current state of change. It also covers the most recent biological work of osthole, which could be very helpful in future research. Osthole has been shown to have anti-inflammatory and anti-proliferative properties. Osthole inhibited cell proliferation and caused cell cycle arrest in lung and ovarian cancer. Osthole regulated phosphorylation of signaling proteins in human breast cancer cells. Furthermore, osthole-induced activation of JNK protein-mediated apoptosis in both cell lines.

Keyword: -*Cnidium monnieri*, Breast Cancer, Osthole, Ion Channel Control, Hepatitis.

Introduction-

Osthole (also known as osthol) is a natural coumarin found in the *Cnidium* plant. 7-methoxy-8-(3-methyl-2-butenyl)-2H-1-benzopyran-2-one. The ripe fruit *Cnidium monnieri* (Fructus *Cnidii*), commonly used in Traditional Chinese Medicine (TCM) clinics, contains high levels of osthole, as do other medicinal plants such as *Angelica*, *Archangelica*, *Citrus*, and *Clausena*. Most of the therapeutic effects of *Fructus Cnidii* are considered to be one of its most important bioactive compounds, osthole [1, 2], which stimulates the immune system and improves male function while reducing rheumatic pain and removing moisture. Antioxidant, anticancer, anti-inflammatory, and immunomodulatory functions of osthole have been found in recent studies [1, 3, 4]. The development of osthole and the emergence of alternatives as a targeted treatment should be encouraged, given the various activities of osthole described. As a result, it is important to examine the medical and biological studies of this coumarin, as well as the basic mechanisms of its action and the full picture of its various roles. Osthole has a wide variety of biological and pharmacological effects. Although the effects of osthole are divided into different biological processes in this study, there is a strong correlation between them (Figure 1).

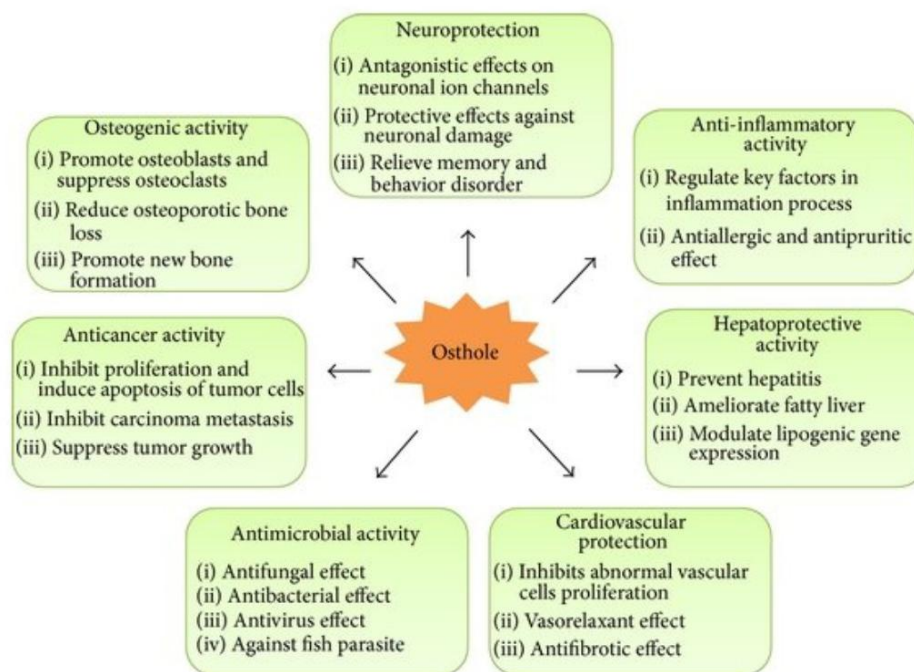


Fig.1- Several systemic pharmacological and therapeutic effects, as well as related experimental findings

7-methoxy-8-(3-methyl-2-butenyl)-2H-1-benzopyran-2-one, commonly known as osthole, is a natural coumarin isolated from the *Cnidium* plant. The small fruit *Cnidium monnieri* (*Fructus Cnidii*), widely used in traditional Chinese medicine, contains high levels of osthole [5, 6]. *Angelica*, *Archangelica*, citrus, and *Clausena* are a few medicinal plants that contain osthole. Osthole shows anti-inflammatory, anticancer, antiapoptosis, antithrombosis, and anti-aggregation properties [7]. Several studies have shown that osthole has a hepatoprotective effect. Osthole has been shown to reduce the release of hepatitis B virus (HBV) from cell culture and to protect mice against hepatitis caused by concanavalin A or the anti-Fas antibody [8,9,10]. Osthole promotes hepatic fibrosis, inhibits hepatic stellate cell function [10], enhances antitumor immune responses [12,13,14], and reduces the development of hepatocellular carcinoma [15]. It also slows the growth of hepatocellular carcinoma. In the mouse model of bleeding caused by trauma, osthole also protects against liver damage [16]. Osthole, as shown in a recent study, is an effective treatment for acetaminophen liver injury (APAP) [17], a condition characterized by oxidative stress [18]. On the other hand, the effect of osthole on hepatotoxicity induced by TMX was not studied. In this work, we examined the effect of osthole on TMX caused by severe liver damage and the underlying mechanism.

Pharmacokinetics and Metabolism of Osthole -

The pharmacokinetics of osthole in mouse plasma after oral or intravenous treatment were investigated using the HPLC method, which produced a concentration / time curve with a rapid distribution phase followed by a long term phase [19 - 22]. Ozone absorption in the rat single pass intestine perfusion (SPIP) model was investigated using HPLC, and the results revealed that osthole absorption was a diffusion process performed in all parts of the intestine [23]. In male SD mice, osthole metabolism was assessed following oral administration, and 10 phase I and 3 phase II metabolites were excreted and detected in urine. Hydroxylation, demethylation, and hydrogenation were the most important phases of metabolic I, while glucuronidation contributed to phase II metabolism [24]. In the human colorectal Caco2 cell model, osthole absorption and metabolism were also examined. Desmethyl-osthol and its various isomers have been an important phase I metabolite for Caco-2 cells, which have a strong penetration and accumulation [25,26]. cAMP and cGMP levels. In a study of osthole biological functions, it was found that osthole induces indirect proliferation of intracellular and tissue cAMP and cGMP, which may be involved in the basic mechanism of some of the osthole organisms. The second messengers, cAMP and cGMP, produced at ATP and GTP, respectively, are important in many biological processes. cAMP significantly affects protein kinase A (PKA), as well as ion channels and growth hormone, by activating it. cGMP regulates ion channels involved in cell cycle binding, apoptosis, and smooth muscle relaxation. Osthole increases cAMP and cGMP levels by inhibiting phosphodiesterases (PDEs), which in turn hydrolyze cAMP and cGMP by destroying the phosphodiester link [27,28]. Increased cAMP and cGMP

-osthole was found to be associated with an action to prevent vascular smoothness. muscle cells [29] .when cGMP / PKG-dependent pathway stimulation was linked to the release of osthole-induced glutamate into the hippocampus synaptosomes [30]. Increases in cGMP have been found to be responsible for the inhibition of osthole-mediated inflammatory substances in the carrageenan model [31]. In addition, the researchers speculated that the osorelaxantosthole properties were linked to higher levels of cAMP and cGMP as a result of osthole treatment [32, 28, 33]. (Figure 1).

Ion Channel Regulator-

In many cells and tissues, osthole has been reported to alter membrane and other ion channels, including sodium channels, acid-sensing ion channels, CFTR chloride channels, and especially calcium channels. The action of osthole in these ion channels is linked to osthole bioactivities, which include vasorelaxant, immunomodulatory, and antifibrotic effect, as well as its neuronal and neuroendocrine activities, which include neuroprotective, anticonvulsant, and pain relievers. Calcium depletion, on the other hand, affects the activity of osteoprogenitors, which play an important role in both bone homeostasis and regeneration [34,35], making the calcium channel a promising way to investigate the mechanism of osteogenic osthole effect. cAMP and cGMP have long been known to play a role in regulating L-type Ca^{2+} channels and other ion channels [36, 37, 38]. As a result, mutations in cAMP and cGMP levels are at least a major cause of the osthole effect on ion channels (Figure 2).

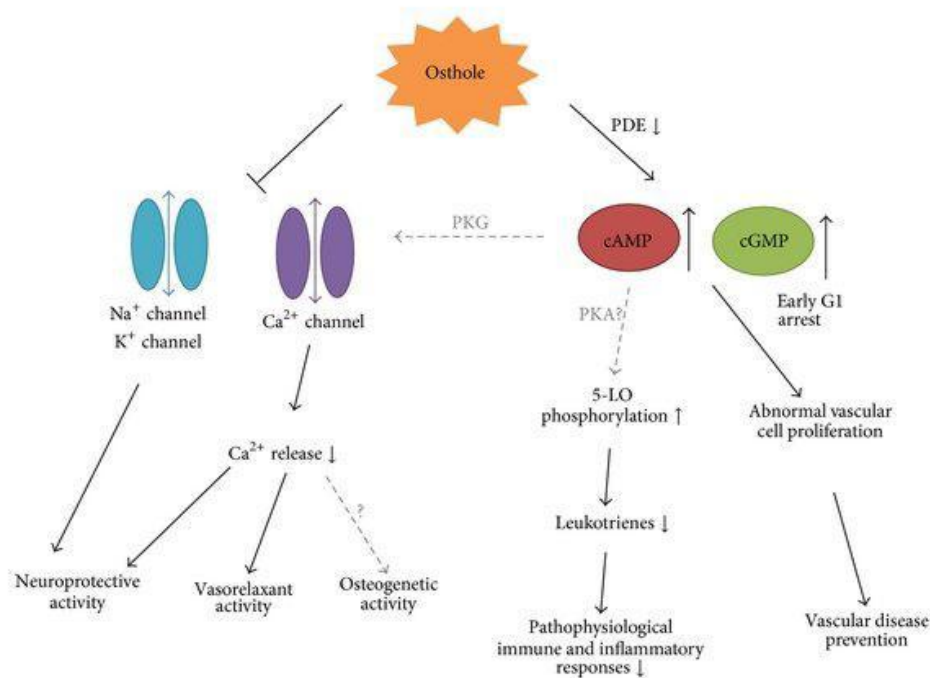


Fig.2.Possible connections between osthole's influence on intracellular ion channels, cyclic adenosine monophosphate (cAMP) levels, and cyclic guanosine monophosphate (cGMP) levels, as well as some of their pharmacological action. (A dashed line indicates unconfirmed possibilities in osthole research.)

Osthole (7-methoxy-8-isopentenoxycoumarin) is a monomer isolated from *Cnidium monnieri* (L.) Cusson that exhibits anti-inflammatory, anti-inflammatory, and leishmanial properties (39,40). Traditional Chinese medicine has been used to treat a variety of ailments, including allergies, inflammation, HIV infection, and diabetes, in clinics for many years. In several types of human cancer, osthole has been shown to have anti-inflammatory and anti-proliferative properties. Osthole, in particular, promotes tumor cell death, inhibits cell cycle development, and inhibits tumor cell migration (41-48). In addition, osthole has been shown to enhance the anticancer activity of cisplatin in rhabdomyosarcoma cells (49) and to prevent hepatocellular carcinoma in other studies (46). These findings suggest that osthole may play a role in the treatment of human cancer, particularly cervical cancer. In this study, the anticancer activity of osthole was tested in vitro as a single agent or in combination with irradiation. In cervical cancer cells, the basic cellular events of osthole treatment were also considered. This should have been the first step in determining whether osthole should be used to treat cervical cancer. Recent research suggests that osthole is a potent antidepressant for many diseases of the nervous system and effectively crosses the blood-brain barrier. Oral osthole treatment reduces the inflammatory response to local

ischemic stroke [50], whereas intraperitoneal injection has a neuroprotective effect on traumatic brain injury due to its antioxidative and antiapoptotic properties [51]. In transient cerebral ischemia, osthole treatment enhances neurobehavioral function and reduces infarct volume [52]. In addition, osthole has been shown to protect against acute ischemic stroke caused by middle cerebral ischemia when administered intraperitoneally. Potato Fusarium wilt is a common soil-borne disease that causes severe symptoms of rot and ultimately plant death, which has a direct impact on potato harvesting and quality [54,55,56]. The disease is widespread worldwide, and often results in a 30% loss of productivity. According to Rakhimov et al. (2000), potato Fusarium wilt is caused by one of the five species of *Fusarium*: *Fusariumoxysporum*Schlecht, *Fusariumsolani* (Mart.) Sacc., *Fusariummoniliforme*Sheld, *Fusariumsambucinum*Fuckel, and *Fusariumnivale* (Fr.) Ces. *F. oxysporum*Schlecht, *F. sol* (Fr.)

In Inner Mongolia, China, Ces are agents of potato Fusarium wilt. Because the disease is a soil-borne pathogen-resistant strain, chemical treatment of soil in the field is a challenge [57]. In the fight against potato Fusarium wilt, contamination and persistence of fungicide residues in potato stages are major challenges. On the other hand, fungicides derived from plants, are attractive options because they are not phytotoxic, easily biodegradable, and environmentally safe [58]. Proteins, alkaloids, flavonoids, phenols, essential oils, and polysaccharides are examples of plant components that have antifungal effects [58 - 63]. *Cnidiummonnieri* (L.) is a traditional Chinese plant found throughout China [64 - 62]. *C. monnieri* contains a total of 429 chemicals, 56 of which were chemically identified [67]. Osthole is an O methylated coumarin that was obtained and purified from *C*-seeds. *monnieri*. It has been shown to have pharmacological properties in humans, including anti-allergic, anti-pruritic, anti-bacterial, anti-dermatophytic, anti-osteoporotic, and anti-fungal effects [68 - 72]. In addition, osthole has been shown to promote osteogenesis in osteoblasts by regulating transcription factor osterix by cAMP / CREB signaling [73].

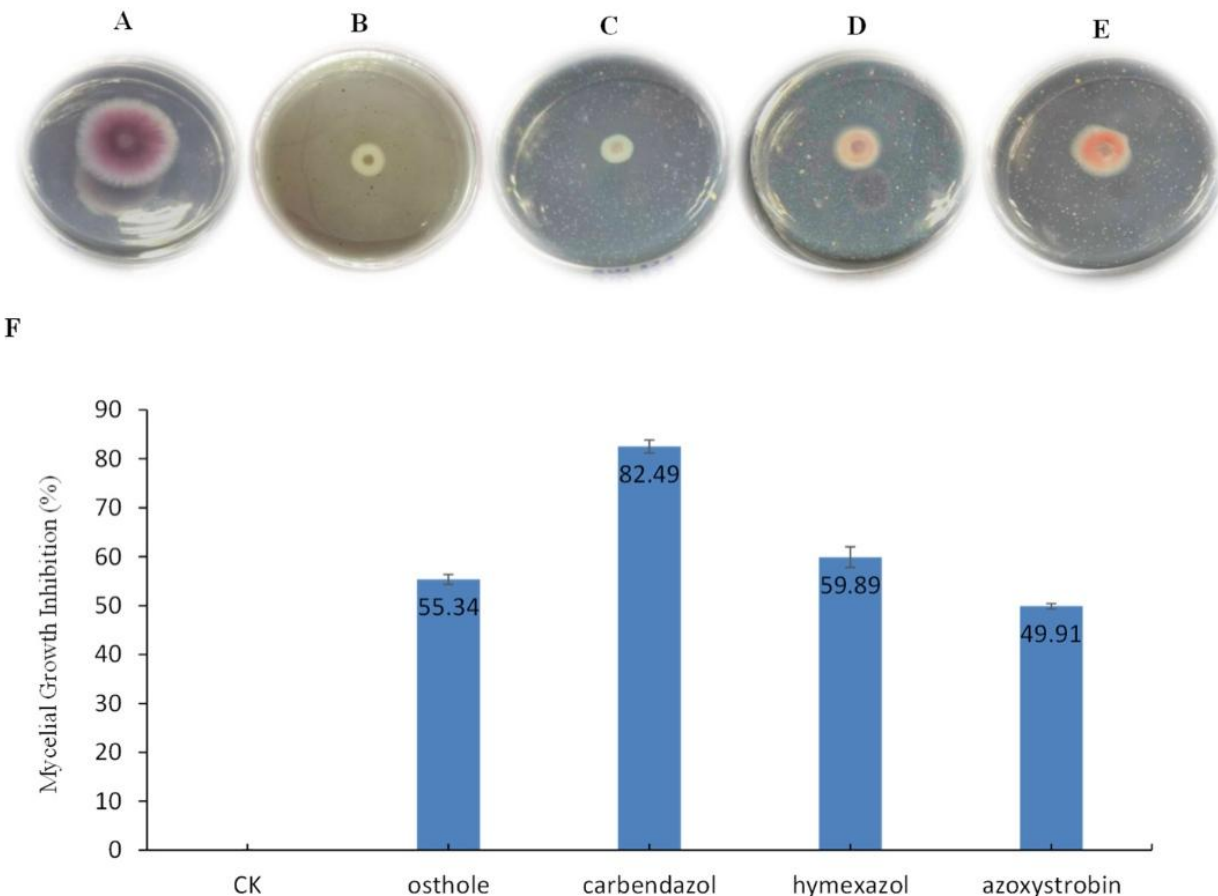


Fig. 3: 3 days after injection, osthole has a preventive effect on *Fusariumoxysporum*. (A) CK, potato dextrose agar (PDA) medium; (B) PDA medium plus 5 mg / mL osthole; (C) PDA medium plus 5 mg / mL carbendazol powder wettable; (D) PDA medium plus 5 mg / mL hymexazol aqueous solution; (E) PDA medium plus 5 mg / mL azoxystrobin; (F) In PDA medium, CK The normal deviation is indicated by the bars in the columns.

In the United States, breast cancer is the leading cause of death by cancer [74]. The incidence of breast cancer has been steadily increasing and it is predicted that it will continue [75]. The pathogenesis of breast cancer is accelerated by altering reproductive patterns, such as pregnancy in later years and shorter breastfeeding [76]. Because fewer symptoms appear in the early stages of breast cancer, it is more commonly found in the advanced stages. Although diagnostic and therapeutic approaches have improved, many barriers to treatment persist, including metastasis, adverse pharmacological effects, and drug resistance [77]. About 50% of patients with advanced breast cancer are resistant to radiation due to the hypoxic tumor nature [78]. In addition, the heterogeneity of breast cancer makes it difficult to treat with certain chemotherapeutic drugs [79]. Based on gene expression profiles, breast cancer cells can be divided into five subtypes: luminal A, luminal B, HER2-overexpression, basal, and normal similarities. Clinical outcomes vary depending on the subtype [80].

Coumarin and its derivatives have been extensively investigated in terms of their biological effects. The molecular structure of coumarin is thought to play a role in its pharmacological activities [81]. Coumarin extracts have been used to treat CNS infections as well as anticoagulants and anti-HIV drugs [82,83,84].

Osthole (7-methoxy-8-(3-methyl-2-butenyl) coumarin) is a phytochemical found in *Cnidium monnieri* (L.) Cusson, a plant of traditional medicine. Osthole is an anti-inflammatory, antimicrobial, and anti-allergic [85,86]. It has received many reports for its anti-cancer properties. Osthole has been shown to be effective in treating cancers such as lung, liver, cervical and ovary cancers.

Osthole also induced apoptosis in immature hepatocellular carcinoma cells and prevented the formation of hepatic tissue in mice [87]. Osthole inhibited cell proliferation and caused cell cycle arrest in lung and ovarian cancer [88,89]. It inhibits cell growth and metastasis, which has anti-cancer properties in breast cancer [90]. According to a recent study [91], Osthole blocked triple-negative breast cancer cell lines by blocking the STAT3 signaling pathway. Reduces cell proliferation and stops cell cycle. Osthole weakens the mitochondrial membrane and increases the levels of cytosolic calcium, which disrupts cellular homeostasis. In addition, osthole induced apoptosis in breast cancer cells by activating ER stress proteins and regulating the activity of PI3K / Akt and MAPK signaling pathways (Figure 4).

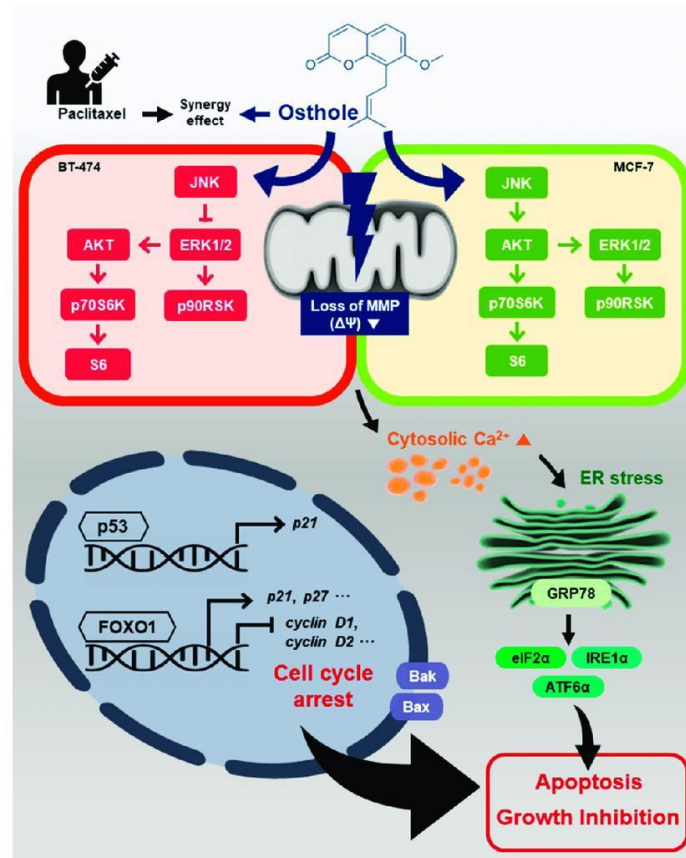


Figure 4 shows a possible osthole mechanism in human breast cancer cell lines. Osthole resulted in the loss of MMP and an increase in cytoplasmic calcium levels. In BT-474 and MCF-7 cells, protein-response proteins and pro-apoptotic proteins are increased. Osthole inhibited JNK in both cell lines, reduced the hallmark modes of Akt and ERK1 / 2 in BT-474 cells, and inhibited these pathways in MCF-7 cells. In breast cancer cells, Osthole caused growth inhibition and apoptosis.

We also looked at osthole (7-methoxy-8-(3-methyl-2-butenyl)-2H-1-benzopyran-2-one), C prenylatedcoumarin derivative and neuroprotective, osteogenic, anticancer, hepatoprotective, and possible antioxidant. results (fig.5). The leaves of citrus [92] and *Cnidium monnieri*, a flowering plant used in traditional Chinese medicine, contain osthole. For centuries, *C. monnieri* is used to treat itching, irritation, and other skin conditions [93].

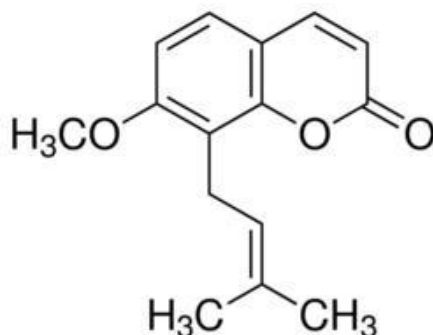


Fig.5 Chemical structure of osthole

Table 1. X-ray Crystal Structural Characterization of Osthole

Name	Osthole
Empirical Formula	C ₁₅ H ₁₆ O ₃
Formula Weight	244.28
Crystal System	Triclinic
Space Group	P-1
Unit Cell Dimensions	7.4543(11) Å 65.145(3)°
	9.4107(14) Å 74.836(3)°
	10.7594(16) Å 70.553(3)°
Volume	639.23(17) Å ³
Z	2
Absorption Coefficient	0.088 mm ⁻¹
Crystal size	0.280 × 0.190 × 0.050 mm ³
Θ range data collection	2.107 to 32.054°
Reflections Collected	16,847
Independent Reflections	4224 [R(int) = 0.0273]
Completeness to Θ = 25.242°	100.0%
Absorption correction	Empirical
Data/restraints/parameters	4224/0/228
Goodness of fit on F ²	1.034
Final R indices [I > 2 σ(I)]	R1 = 0.0425, wR2 = 0.1169
R indices (all data)	R1 = 0.0560, wR2 = 0.1276
Largest diff peak and hole	0.417 and -0.248 e.Å ⁻³

X-ray osthole data. All data collected at temperature = 125 (2) K and (Mo) λ = 0.71073 Å. The structure of the osthole has already been defined at room temperature [94]. The boundaries of our cell and the structure of 1989 (CCDC code JAKFIK) are related to our reduced cell. (Table 1 summarizes our osthole crystal structure data). Figure 6, shows the formation of osthole cells, which do not produce unique features. As shown in Figure 5, the crystal structure shows intermolecular and osthole interactions with offset coagulation pattern. Figure shows how the molecule is folded into C10, at an angle of 74° between the aromatic coumarin system and the alkyl component. Offset stacking is similar to bergamottin, but with CH...O (carbonyl) hydrogen bond 3.576 (3) and CHO angle 161 (1) instead of 3.366 and CH...O weak (carbonyl) hydrogen bond of 3.576 (3).

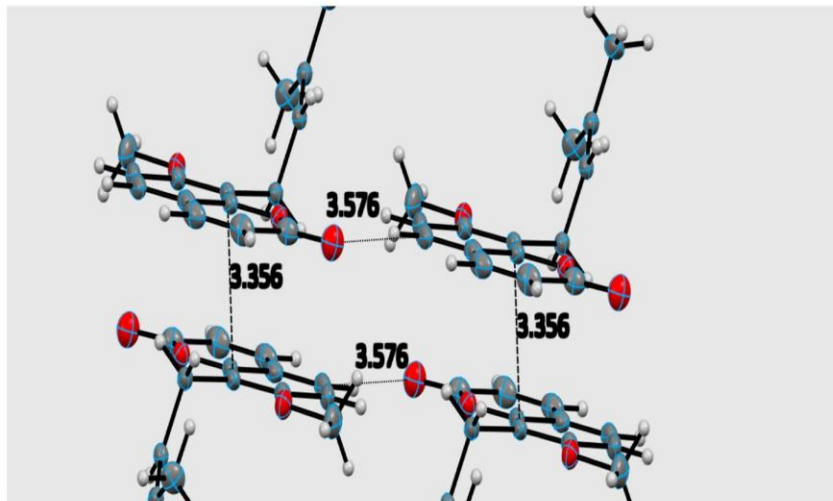


fig.6 Hydrogen interaction and accumulation of osthole crystal structure

conclusion-

Numerous studies have shown that osthole has therapeutic benefits such as neuroprotection, osteogenesis, immunomodulation, and anti-cancer properties, making it an additional multitarget drug and effective diet. Although the basic mechanisms of these characteristics are unknown, the action of osthole control at cAMP and cGMP levels, as well as specific ion channels, can be considered as a contribution to their value. Further research is needed to improve our understanding of the effects of osthole medication and to address safety concerns in order to use this natural substance and its extracts as a preventive and curative agent in humans. In human breast cancer cells, we have found that osthole has anti-cancer properties. Osthole reduced cell growth by lowering the levels of PCNA and caused cell cycle arrest. Collectively, these findings suggest that osthole may have significant therapeutic promise in glioma patients. Further research is needed to determine the exact effect that osthole has on cervical cancer. We also found that osthole has an anti-tumor effect on TNBC cells, as well as mechanisms that may support it.

Reference:

1. L. You, S. Feng, R. An, and X. Wang, "Osthole: a promising lead compound for drug discovery from a traditional Chinese medicine (TCM)," *Natural Product Communications*, vol. 4, no. 2, pp. 297–302, 2009.
2. Z.-W. Zhou and P.-X. Liu, "Progress in study of chemical constituents and anti-tumor activities of *Cnidiummonnieri*," *ZhongguoZhongyaoZazhi*, vol. 30, no. 17, pp. 1309–1313, 2005.
3. J. R. S. Hoult and M. Paya, "Pharmacological and biochemical actions of simple coumarins: natural products with therapeutic potential," *General Pharmacology*, vol. 27, no. 4, pp. 713–722, 1996.
4. R. Wang, J. Kong, D. Wang, L. L.-M. Lien, and E. J.-C. Lien, "A survey of Chinese herbal ingredients with liver protection activities," *Chinese Medicine*, vol. 2, article 5, 2007.
5. Wang R, Kong J, Wang D, Lien LL, Lien EJ. A survey of Chinese herbal ingredients with liver protection activities. *Chin Med*. 2007;2:5.
6. You L, Feng S, An R, Wang X. Osthole: a promising lead compound for drug discovery from a traditional Chinese medicine (TCM). *Nat Product Commun*. 2009;4:297–302.
7. Hoult JR, Paya M. Pharmacological and biochemical actions of simple coumarins: natural products with therapeutic potential. *Gen Pharmacol*. 1996;27:713–22.
8. Huang RL, Chen CC, Huang YL, Hsieh DJ, Hu CP, Chen CF, et al. Osthole increases glycosylation of hepatitis B surface antigen and suppresses the secretion of hepatitis B virus in vitro. *Hepatology*. 1996;24:508–15.
9. Okamoto T, Yoshida S, Kobayashi T, Okabe S. Inhibition of concanavalin A-induced mice hepatitis by coumarin derivatives. *Jpn J Pharmacol*. 2001;85:95–7.
10. Okamoto T, Kawasaki T, Hino O. Osthole prevents anti-Fas antibody-induced hepatitis in mice by affecting the caspase-3-mediated apoptotic pathway. *Biochem Pharmacol*. 2003;65:677–81.
11. Qi Z, Xue J, Zhang Y, Wang H, Xie M. Osthole ameliorates insulin resistance by increment of adiponectin release in high-fat and high-sucrose-induced fatty liver rats. *Planta Med*. 2011;77:231–5.

12. Zhang J, Xue J, Wang H, Zhang Y, Xie M. Osthole improves alcohol-induced fatty liver in mice by reduction of hepatic oxidative stress. *Phytother Res: PTR*. 2011;25:638–43.
13. Liu YW, Chiu YT, Fu SL, Huang YT. Osthole ameliorates hepatic fibrosis and inhibits hepatic stellate cell activation. *J Biomed Sci*. 2015;22:63.
14. Zhang L, Jiang G, Yao F, Liang G, Wang F, Xu H, et al. Osthole promotes antitumor immune responses in tumor-bearing mice with hepatocellular carcinoma. *ImmunopharmacolImmunotoxicol*. 2015;37:301–7.
15. Zhang L, Jiang G, Yao F, He Y, Liang G, Zhang Y, et al. Growth inhibition and apoptosis induced by osthole, a natural coumarin, in hepatocellular carcinoma. *PLoS ONE*. 2012;7:e37865.
16. Yu HP, Liu FC, Tsai YF, Hwang TL. Osthole attenuates hepatic injury in a rodent model of trauma-hemorrhage. *PLoS ONE*. 2013;8:e65916.
17. Cai Y, Sun W, Zhang XX, Lin YD, Chen H, Li H. Osthole prevents acetaminophen-induced liver injury in mice. *ActaPharmacol Sin*. 2018;39:74–84.
18. James LP, McCullough SS, Knight TR, Jaeschke H, Hinson JA. Acetaminophen toxicity in mice lacking NADPH oxidase activity: role of peroxynitrite formation and mitochondrial oxidant stress. *Free Radic Res*. 2003;37:1289–97.
19. T. H. Tsai, T. R. Tsai, C. C. Chen, and C. F. Chen, “Pharmacokinetics of osthole in rat plasma using high-performance liquid chromatography,” *Journal of Pharmaceutical and Biomedical Analysis*, vol. 14, no. 6, pp. 749–753, 1996.
20. Y. Li, F. Meng, Z. Xiong, H. Liu, and F. Li, “HPLC determination and pharmacokinetics of osthole in rat plasma after oral administration of fructuscnidii extract,” *Journal of Chromatographic Science*, vol. 43, no. 8, pp. 426–429, 2005.
21. J. Zhou, S. W. Wang, and X. L. Sun, “Determination of osthole in rat plasma by high-performance liquid chromatograph using cloud-point extraction,” *AnalyticaChimicaActa*, vol. 608, no. 2, pp. 158–164, 2008.
22. J. Zhou, P. Zeng, Z. H. Cheng, J. Liu, F. Q. Wang, and R. J. Qian, “Application of hollow fiber liquid phase microextraction coupled with high-performance liquid chromatography for the study of the osthole pharmacokinetics in cerebral ischemia hypoperfusion rat plasma,” *Journal of Chromatography B: Analytical Technologies in the Biomedical and Life Sciences*, vol. 879, no. 23, pp. 2304–2310, 2011.
23. Y.-N. Wu and L.-B. Luan, “In situ rats single pass perfusion intestinal absorption of the effective components in *Radix AngelicaePubescentis*,” *Yao XueXueBao*, vol. 43, no. 1, pp. 102–107, 2008.
24. X. Lv, C.-Y. Wang, J. Hou et al., “Isolation and identification of metabolites of osthole in rats,” *Xenobiotica*, vol. 42, no. 11, pp. 1120–1127, 2012.
25. X.-W. Yang, Q.-M. Guo, and Y. Wang, “Absorption and transport of 6 coumarins isolated from the roots of *Angelica pubescens f. biserrata* in human Caco-2 cell monolayer model,” *Zhong Xi Yi Jie He XueBao*, vol. 6, no. 4, pp. 392–398, 2008.
26. Z. Yuan, H. Xu, K. Wang, Z. Zhao, and M. Hu, “Determination of osthol and its metabolites in a phase I reaction system and the Caco-2 cell model by HPLC-UV and LC-MS/MS,” *Journal of Pharmaceutical and Biomedical Analysis*, vol. 49, no. 5, pp. 1226–1232, 2009.
27. C.-M. Teng, F.-N. Ko, J.-P. Wang et al., “Antithrombotic effect of some antiplatelet agents isolated from Chinese herbs,” *The Journal of Pharmacy and Pharmacology*, vol. 43, no. 9, pp. 667–669, 1991.
28. C.-M. Teng, C.-H. Lin, F.-N. Ko, T.-S. Wu, and T.-F. Huang, “The relaxant action of osthole isolated from *Angelica pubescens* in guinea-pig trachea,” *Naunyn-Schmiedeberg’s Archives of Pharmacology*, vol. 349, no. 2, pp. 202–208, 1994.
29. J.-H. Guh, S.-M. Yu, F.-N. Ko, T.-S. Wu, and C.-M. Teng, “Antiproliferative effect in rat vascular smooth muscle cells by osthole, isolated from *Angelica pubescens*,” *European Journal of Pharmacology*, vol. 298, no. 2, pp. 191–197, 1996.
30. T. Y. Lin, C. W. Lu, W.-J. Huang, and S.-J. Wang, “Involvement of the cGMP pathway in the osthole-facilitated glutamate release in rat hippocampal nerve endings,” *Synapse*, vol. 66, no. 3, pp. 232–239, 2012.
31. J. Liu, W. Zhang, L. Zhou, X. Wang, and Q. Lian, “Anti-inflammatory effect and mechanism of osthole in rats,” *Journal of Chinese Medicinal Materials*, vol. 28, no. 11, pp. 1002–1006, 2005.
32. F.-N. Ko, T.-S. Wu, M.-J. Liou, T.-F. Huang, and C.-M. Teng, “Vasorelaxation of rat thoracic aorta caused by osthole isolated from *Angelica pubescens*,” *European Journal of Pharmacology*, vol. 219, no. 1, pp. 29–34, 1992.
33. W.-F. Chiou, Y.-L. Huang, C.-F. Chen, and C.-C. Chen, “Vasorelaxing effect of coumarins from *Cnidiummonnieri* on rabbit corpus cavernosum,” *Planta Medica*, vol. 67, no. 3, pp. 282–284, 2001.
34. H. Yang, L. N. Xu, Y. J. Sui et al., “Stimulation of airway and intestinal mucosal secretion by natural coumarin CFTR activators,” *Frontiers in Pharmacology*, vol. 2, article 52, 2011.
35. A. M. C. Barradas, H. A. M. Fernandes, N. Groen et al., “A calcium-induced signaling cascade leading to osteogenic differentiation of human bone marrow-derived mesenchymal stromal cells,” *Biomaterials*, vol. 33, no. 11, pp. 3205–3215, 2012.
36. C. L. Chik, Q.-Y. Liu, B. Li, E. Karpinski, and A. K. Ho, “cGMP Inhibits L-type Ca²⁺ channel currents through protein phosphorylation in rat pinealocytes,” *Journal of Neuroscience*, vol. 15, no. 4, pp. 3104–3109, 1995.
37. T. J. Kamp and J. W. Hell, “Regulation of cardiac L-type calcium channels by protein kinase A and protein kinase C,” *Circulation Research*, vol. 87, no. 12, pp. 1095–1102, 2000.
38. B. Ay, A. Iyanoye, G. C. Sieck, Y. S. Prakash, and C. M. Pabelick, “Cyclic nucleotide regulation of store-operated Ca²⁺ influx in airway smooth muscle,” *The American Journal of Physiology: Lung Cellular and Molecular Physiology*, vol. 290, no. 2, pp. L278–L283, 2006.
39. Zimecki M, Artym J, Cisowski W, Mazol I, Włodarczyk M and Gleńsk M: Immunomodulatory and anti-inflammatory activity of selected osthole derivatives. *Z Naturforsch C* 64: 361-368, 2009.

40. Hao Y and Liu Y: Osthole alleviates bleomycin-induced pulmonary fibrosis via modulating angiotensin-converting enzyme 2/angiotensin-(1-7) axis and decreasing inflammation responses in rats. *Biol Pharm Bull* 39: 457-465, 2016.
41. Jiang G, Liu J, Ren B, Tang Y, Owusu L, Li M, Zhang J, Liu L and Li W: Anti-tumor effects of osthole on ovarian cancer cells in vitro. *J Ethnopharmacol* 193: 368-376, 2016.
42. Ding D, Wei S, Song Y, Li L, Du G, Zhan H and Cao Y: Osthole exhibits anti-cancer property in rat glioma cells through inhibiting PI3K/Akt and MAPK signaling pathways. *Cell PhysiolBiochem* 32: 1751-1760, 2013.
43. Yang D, Gu T, Wang T, Tang Q and Ma C: Effects of osthole on migration and invasion in breast cancer cells. *BiosciBiotechnolBiochem* 74: 1430-1434, 2010.
44. Wu C, Sun Z, Guo B, Ye Y, Han X, Qin Y and Liu S: Osthole inhibits bone metastasis of breast cancer. *Oncotarget* 8: 58480-58493, 2017.
45. Chou SY, Hsu CS, Wang KT, Wang MC and Wang CC: Antitumor effects of Osthol from *Cnidiummonnieri*: An in vitro and in vivo study. *Phytother Res* 21: 226-230, 2007.
46. Okamoto T, Kobayashi T and Yoshida S: Chemical aspects of coumarin compounds for the prevention of hepatocellular carcinomas. *Curr Med Chem Anticancer Agents* 5: 47-51, 2005.
47. Yang LL, Wang MC, Chen LG and Wang CC: Cytotoxic activity of coumarins from the fruits of *Cnidiummonnieri* on leukemia lines. *Planta Med* 69: 1091-1095, 2003.
48. Kao SJ, Su JL, Chen CK, Yu MC, Bai KJ, Chang JH, Bien MY, Yang SF and Chien MH: Osthole inhibits the invasive ability of human lung adenocarcinoma cells via suppression of NF- κ B-mediated matrix metalloproteinase-9 expression. *ToxicolApplPharmacol* 261: 105-115, 2012.
49. Jarzab A, Luszczki J, Guz M, Skalicka-Wozniak K, Halasa M, Smok-Kalwat J, Polberg K and Stepulak A: Combination of osthole and cisplatin against rhabdomyosarcoma TE671 cells yielded additive pharmacologic interaction by means of isobolographic analysis. *Anticancer Res* 38: 205-210, 2018.
50. Li, F.; Gong, Q.; Wang, L.; Shi, J. Osthole attenuates focal inflammatory reaction following permanent middle cerebral artery occlusion in rats. *Biol. Pharm. Bull.* 2012, 35, 1686–1690.
51. He, Y.; Qu, S.; Wang, J.; He, X.; Lin, W.; Zhen, H.; Zhang, X. Neuroprotective effects of osthole pretreatment against traumatic brain injury in rats. *Brain Res.* 2012, 1433, 127–136.
52. Mao, X.; Yin, W.; Liu, M.; Ye, M.; Liu, P.; Liu, J.; Lian, Q.; Xu, S.; Pi, R. Osthole, a natural coumarin, improves neurobehavioral functions and reduces infarct volume and matrix metalloproteinase-9 activity after transient focal cerebral ischemia in rats. *Brain Res.* 2011, 1385, 275–280.
53. Chao, X.; Zhou, J.; Chen, T.; Liu, W.; Dong, W.; Qu, Y.; Jiang, X.; Ji, X.; Zhen, H.; Fei, Z. Neuroprotective effect of osthole against acute ischemic stroke on middle cerebral ischemia occlusion in rats. *Brain Res.* 2010, 1363, 206–211.
54. Rakhimov, U.K.; Khakimov, A.K. Wilt of potatoes in Uzbekistan. *Zashchita i Karantin Rastenii* 2000, 3, 46.
55. Peng, X.W.; Zhu, J.H. Species and Distribution of potato Fungal Diseases in Hebei Province, China. *Chin. Potato J.* 2008, 22, 31–33.
56. An, X.M.; Hu, J.; Wu, J.H.; Liu, Z.H.; Meng, M.L. Overview of pathogen causing potato Fusarium Wilt. *Chin. Potato J.* 2017, 31, 302–306.
57. Khedher, S.B.; Mejdoub-Trabelsi, B.; Tounsi, S. Biological potential of *Bacillus subtilis* V26 for the control of Fusarium wilt and tuber dry rot on potato caused by Fusarium species and the promotion of plant growth. *Biol. Control* 2020, 152, 104444. [CrossRef]
58. Chen, Y.H.; Lu, M.H.; Guo, D.S.; Zhai, Y.Y.; Miao, D.; Yue, J.Y.; Yuan, C.H.; Zhao, M.M.; An, D.R. Antifungal Effect of Magnolol and Honokiol from *Magnolia officinalis* on *Alternaria alternata* Causing Tobacco Brown Spot. *Molecules* 2019, 24, 2140. [CrossRef]
59. Mi, Y.Y.; Byeong, J.C.; Jin, C.K. Recent Trends in Studies on Botanical Fungicides in Agriculture. *Plant Pathol.* 2013, 29, 1–9.
60. Kijjoa, A.; Pinto, M.M.M.; Tantisewie, B.; Herz, W. A biphenyl type neolignan and biphenyl ether from *Magnolia henryi*. *Phytochemistry* 1989, 28, 1284–1286. [CrossRef]
61. Du, C.M.; Wu, Y.H.; Zhao, X.X.; Zhu, C.Y.; Jiang, G.; Yan, X.M. Recent development in research of natural antiphytoviral substances. *Acta Tabacaria Sin.* 2004, 10, 34–40.
62. Chen, Y.H.; Ru, B.L.; Zhai, Y.Y.; Li, J.; Cheng, J.L. Screening and inhibitory effects of plant extracts against Tobacco mosaic virus (TMV). *J. Plant Prot.* 2018, 45, 463–469.
63. Zhao, L.; Feng, C.; Wu, K.; Chen, W.B.; Chen, Y.J.; Hao, X.A.; Wu, Y.F. Advances and prospects in biogenic substances against plant virus: A review. *Pestic. Biochem. Phys.* 2016, 135, 15–26. [CrossRef]
64. Kitajima, J.; Aoki, Y.; Ishikawa, T.; Tanaka, Y. Monoterpenoid glucosides of *Cnidiummonnieri* fruit. *Chem. Pharm. Bull.* 1999, 47, 639–642. [CrossRef]
65. Oh, H.; Kim, J.S.; Song, E.K.; Cho, H.; Kim, D.H.; Park, S.E.; Lee, H.S.; Kim, Y.C. Sesquiterpenes with hepatoprotective activity from *Cnidiummonnieri* on tacrine-induced cytotoxicity in Hep G2 cells. *Planta Med.* 2002, 68, 748–749. [CrossRef] [PubMed]
66. Zhao, J.Y.; Zhou, M.; Liu, Y.; Zhang, G.L.; Luo, Y.G. Chromones and coumarins from the dried fructus of *Cnidiummonnieri*. *Fitoterapia* 2011, 82, 767–771. [CrossRef] [PubMed]
67. Sun, Y.; Yang, A.; Lenon, G.B. Phytochemistry, Ethnopharmacology, Pharmacokinetics and Toxicology of *Cnidiummonnieri* (L.) Cusson. *Int. J. Mol. Sci.* 2020, 21, 1006. [CrossRef] [PubMed]
68. Basnet, P.; Yasuda, I.; Kumagai, N.; Tohda, C.; Nojima, H.; Kuraishi, Y.; Komatsu, K. Inhibition of itch-scratch response by fruits of *Cnidiummonnieri* in mice. *Biol. Pharm. Bull.* 2001, 24, 1012–1015. [CrossRef] [PubMed]
69. Bao, J.J.; Xie, M.L.; Zhu, L.J. Treatment of osthol on osteoporosis in ovariectomized rats. *Chin. Pharm. Bull.* 2011, 27, 591–592.

70. Matsuda, H.; Ido, Y.; Hirata, A.; Ino, Y.; Naruto, S.; Amamiya, T.; Kubo, M. Antipruritic effect of *Cnidiummonnieri* Fructus (fruits of *Cnidiummonnieri* Cusson). *Biol. Pharm. Bull.* 2002, 25, 260–263. [CrossRef] [PubMed]
71. Matsuda, H.; Tomohiro, N.; Ido, Y.; Kubo, M. Anti-allergic effects of *Cnidiummonnieri* fructus (dried fruits of *Cnidiummonnieri*) and its major component, osthole. *Biol. Pharm. Bull.* 2002, 25, 809–812. [CrossRef]
72. Li, Y.M.; Jia, M.; Li, H.Q.; Zhang, N.D.; Wen, X.; Rahman, K.; Zhang, Q.Y.; Qin, L.P. *Cnidiummonnieri*: A Review of Traditional Uses, Phytochemical and Ethnopharmacological Properties. *Am. J. Chin. Med.* 2015, 43, 835–877. [CrossRef]
73. Zhang, Z.R.; Leung, W.N.; Li, G.; Kong, S.K.; Lu, X.; Wong, Y.M.; Chan, C.W. Osthole Enhances Osteogenesis in Osteoblasts by Elevating Transcription Factor Osterix via cAMP/CREB Signaling In Vitro and In Vivo. *Nutrients* 2017, 9, 588. [CrossRef]
74. Siegel, R.L.; Miller, K.D.; Jemal, A. Cancer statistics, 2018. *CA A Cancer J. Clin.* 2018, 68, 7–30. [CrossRef]
75. Winters, S.; Martin, C.; Murphy, D.; Shokar, N.K. Breast Cancer Epidemiology, Prevention, and Screening. *Prog. Mol. Biol. Transl. Sci.* 2017, 151, 1–32. [CrossRef] [PubMed]
76. Winters, S.; Martin, C.; Murphy, D.; Shokar, N.K. Chapter One—Breast Cancer Epidemiology, Prevention, and Screening. In *Progress in Molecular Biology and Translational Science*; Lakshmanaswamy, R., Ed.; Academic Press: Cambridge, MA, USA, 2017; Volume 151, pp. 1–32.
77. Redig, A.J.; McAllister, S.S. Breast cancer as a systemic disease: A view of metastasis. *J. Intern. Med.* 2013, 274, 113–126. [CrossRef] [PubMed]
78. Sinha, D.; Sarkar, N.; Biswas, J.; Bishayee, A. Resveratrol for breast cancer prevention and therapy: Preclinical evidence and molecular mechanisms. In *Seminars in Cancer Biology*; Academic Press: Cambridge, MA, USA, 2016; Volume 40, pp. 209–232. [CrossRef]
79. Rattani, N.S.; Swift-Scanlan, T. Deconstructing breast cancer heterogeneity: Clinical implications for women with Basal-like tumors. *Oncol. Nurs. Forum* 2014, 41, 639–646. [CrossRef] [PubMed]
80. Sorlie, T.; Perou, C.M.; Tibshirani, R.; Aas, T.; Geisler, S.; Johnsen, H.; Hastie, T.; Eisen, M.B.; van de Rijn, M.; Jeffrey, S.S.; et al. Gene expression patterns of breast carcinomas distinguish tumor subclasses with clinical implications. *Proc. Natl. Acad. Sci. USA* 2001, 98, 10869–10874. [CrossRef] [PubMed]
81. Hoult, J.R.; Paya, M. Pharmacological and biochemical actions of simple coumarins: Natural products with therapeutic potential. *Gen. Pharmacol.* 1996, 27, 713–722. [CrossRef]
82. Skalicka-Woźniak, K.; Orhan, I.E.; Cordell, G.A.; Nabavi, S.M.; Budzyńska, B. Implication of coumarins towards central nervous system disorders. *Pharmacol. Res.* 2016, 103, 188–203. [CrossRef] [PubMed]
83. Holbrook, A.M.; Pereira, J.A.; Labiris, R.; McDonald, H.; Douketis, J.D.; Crowther, M.; Wells, P.S. Systematic overview of warfarin and its drug and food interactions. *Arch. Intern. Med.* 2005, 165, 1095–1106. [CrossRef]
84. Yu, D.; Suzuki, M.; Xie, L.; Morris-Natschke, S.L.; Lee, K.H. Recent progress in the development of coumarin derivatives as potent anti-HIV agents. *Med. Res. Rev.* 2003, 23, 322–345. [CrossRef]
85. Okamoto, T.; Yoshida, S.; Kobayashi, T.; Okabe, S. Inhibition of concanavalin A-induced mice hepatitis by coumarin derivatives. *Jpn. J. Pharmacol.* 2001, 85, 95–97. [CrossRef]
86. Matsuda, H.; Tomohiro, N.; Ido, Y.; Kubo, M. Anti-allergic effects of *Cnidiummonnieri* fructus (dried fruits of *Cnidiummonnieri*) and its major component, osthole. *Biol. Pharm. Bull.* 2002, 25, 809–812. [CrossRef]
87. Zhang, L.; Jiang, G.; Yao, F.; He, Y.; Liang, G.; Zhang, Y.; Hu, B.; Wu, Y.; Li, Y.; Liu, H. Growth inhibition and apoptosis induced by osthole, a natural coumarin, in hepatocellular carcinoma. *PLoS ONE* 2012, 7, e37865. [CrossRef]
88. Xu, X.; Zhang, Y.; Qu, D.; Jiang, T.; Li, S. Osthole induces G2/M arrest and apoptosis in lung cancer A549 cells by modulating PI3K/Akt pathway. *J. Exp. Clin. Cancer Res.* 2011, 30, 33. [CrossRef]
89. Jiang, G.; Liu, J.; Ren, B.; Tang, Y.; Owusu, L.; Li, M.; Zhang, J.; Liu, L.; Li, W. Anti-tumor effects of osthole on ovarian cancer cells in vitro. *J. Ethnopharmacol.* 2016, 193, 368–376. [CrossRef] [PubMed]
90. Yang, D.; Gu, T.; Wang, T.; Tang, Q.; Ma, C. Effects of osthole on migration and invasion in breast cancer cells. *Biosci. Biotechnol. Biochem.* 2010, 74, 1430–1434. [CrossRef] [PubMed]
91. Dai, X.; Yin, C.; Zhang, Y.; Guo, G.; Zhao, C.; Wang, O.; Xiang, Y.; Zhang, X.; Liang, G. Osthole inhibits triple negative breast cancer cells by suppressing STAT3. *J. Exp. Clin. Cancer Res.* 2018, 37, 322. [CrossRef] [PubMed]
92. Dugrand-Judek, A.; Olry, A.; Hehn, A.; Costantino, G.; Ollitrault, P.; Froelicher, Y.; Bourgaud, F. The distribution of coumarins and furanocoumarins in citrus species closely matches citrus phylogeny and reflects the organization of biosynthetic pathways. *PLoS ONE* 2015, 10, e0142757. [CrossRef] [PubMed]
93. Matsuda, H.; Tomohiro, N.; Ido, Y.; Kubo, M. Anti-allergic effects of *Cnidiummonnieri* fructus (dried fruits of *Cnidiummonnieri*) and its major component, osthole. *Biol. Pharm. Bull.* 2002, 25, 809–812. [CrossRef]
94. Borowiak, T.; Wolska, I. Structure of 7-methoxy-8-(3-methyl-2-butenyl) coumarin. *Acta Cryst. Sect. C* 1989, 45, 620–622. [CrossRef]