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Brucellosis and Its Treatment: An Overview

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ABSTRACT

The facultative intracellular transmitted Bacteria is the Brucella bacteria which is mainly non motile, non spore forming and gram negative bacteria, named after the microbiologist David Bruce in (1855-1931). This bacterium primarily spreads between various type of infected animals to human and the disease is called Brucellosis. This pathogenic zoonoses bacteria has various specises of which most infectious for human's is Brucella melitensis primarily affects camel, goat ,sheep. Again Brucella abortus, Brucella canis, Brucella suis infects hogs,cows,dogs and also others animal's. When this zoonoses bacteria is transmitted in Human's body then it infects our multi organ system specially respiratory, genital system and digestive system. This intracellular pathogen accumulated in the cells and organs of human by the microphase mechanism typically in liver and spleen. The general mechanism of human cell phagocytosis can kill 90-95% of Brucella bacteria after getting internalized by microphages but still few bacteria remain alive which is enough for replication into the host cell.

Keywords: Intracellular Bacteria, Brucella, Zoonoses, Pathogeenic Disease , Non Spore Forming

Introduction

Brucellosis is the generic name used for infections in animals and humans caused by several species of the genus Brucella, mainly Brucella abortus, B. melitensis and B. suis. Brucella infection in cattle is usually caused by B. abortus, less commonly by B. melitensis, and sometimes B. suis. Brucella melitensis is the primary causative agent of Brucella infections in sheep and goats [1]. Infection in Brucella pigs is caused by B. suis 1-3 biovars, but the disease caused by biovar 2 differs in host range, limited geographic coverage and pathogenicity. In some areas, infection with B. suis was found in feral pigs. Clinically, Brucella infection in animals is characterized by one or more of the following : abortion, placental retention, orchitis, epididymitis and, less commonly, arthritis, with microbial excretion in uterine and milk secretions. An unequivocal diagnosis depends on the isolation of Brucella from the abortion material, udder secretions, or tissue removed during dissection. Since B.abortus, B. melitensis and B. suis are highly pathogenic to humans, so they potentially contaminatetissues, cultures and materials must be handled under appropriate conditions [2]. This Bacteria is a harmful pathogenic zoonotic [3] intracellular proteobacteria (Gram-negative bacteria) which infect the cells of human-being, [4]. The term 'zoonosis' can be described as an infectious disease caused by microorganisms that transferred or jumped from animal to a human being.Brucellosis mainly occurs from the domestic animals like sheeps, pigs, rams, cows . This zoonotic bacteria, Brucella consists of total seven species in which Brucella melitensis, Brucella abortus, Brucella canis, Brucella suis are known to induce human disease whereas on the other hand, Brucella neotomae and Brucella ovis are not harmfulfor the human being [5]. Among various species of this zoonotic bacteria Brucella melitensis, Brucella abortus show prominent attack, which mainly infects the multi organ system of the body includingrespiratory and genital system in both human and animal [6]. Thesymptoms include joint infection, fever, cough, cold and its virulence depends on the duration of infection [7]. Brucellosis is also known as 'malta fever' or 'undulant fever' [8].WHO declared this disease to be the most common in every parts of the world [9].

Symptoms and Signs

The patient had multiple acute symptoms and 4,444 signs of human brucellosis, such as wave fever, myalgia (Table 1), and other clinical symptoms such as spleen enlargement, hepatomegaly, and spondylitis. Infective endocarditis, although rare, is the most serious complication of systemic brucellosis and may require surgery. This is a harmful zoonotic disease [10] for human and animal both [11]. The Primary symtoms of brucella is respiratory infection, digestive infection, genital infection but it may be change as per duration of time of the brucellosis infection [12]. As per WHO there are several infection present in Brucella like as join pain, fever, back pain, weight loss, anorexia, fatigue, arthralgia [13]. As per WHO Table 1 : reports symptoms and sings in 500 patient with brucellosis due to brucella melitensis [14].

Symptoms and Signs	Number of Patient	Percentage (%)
Lack of energy	473	95
Fever	464	93
Aches	457	91
Sweats	437	87
Arthritis	202	40
Chills	410	82
Abdominal pain	225	45
Cough	122	24
Headache	403	81
Joint and back pain	431	86
Testicular pain/epididymo-orchitis	62	21
Loss of appetite	388	78
Diarrhoea	34	7
Sleep disturbance	185	37
Pallor	110	22
Cardiac murmur	17	3
Spinal tenderness	241	48
Weight loss	326	65
Central nervous system abnormalities	20	4
Hepatomegaly	97	19
Constipation	234	47
III appearance	127	25
Splenomegaly	125	25
Jaundice	6	1
Lymphadenopathy	160	32
Rash	72	14

Table 1 : Symptoms and signs in 500 patients with brucellosis due to B. melitensis.

Diagnosis

Diagnostic tools for brucella inculdes isolation and identification of Brucella from clinical sample, detection of genome, antigen and antibodies [15]. **1.Culture:**Blood cultures mainly provide the solid evidence of brucellosis, but do not provide positive results for the all patients. Dissolved centrifugation and thrombus culture techniques have calculated promising results in terms of sensitivity and rapidity [16]. The latest automated blood culture system has slightly improved detection speed. Poor quarantine of thisBacterial techniques in blood culture are impractical for routine diagnosis. **2.Antigen detection:** The antigen detection method with the help of enzyme-linked immunosorbent assay (ELISA) [17] is potentially useful and several recent antigen detection systems are under development. **3. Genome detection:** The polymerase chain reaction (PCR) [18]was sought for rapid detection and confirmation of Brucella. The technology of molecular properties is Brucella spp. Is a very useful tool for distinguishing follow-up tests, especially for abnormal phenotypic results. The supply of equipment and reagents is a limitation of this technology [19].

4. Antibody detection: Due to the limitations of the above tools, the serum indicated for antibody detection is the most useful tool. Antibodies generally begin to appear in the blood at the end of the first week of infection [20].

(A) Genathering Test

Testing Bengal Rose Plate (RBPT) is one of the experiments called Brucella buffer antigen tests called the principle based on the principle that the ability of IgM-related antibodies [21], low pH Value is marked. It takes place on glass slides with antigen color bacteria, and this test is as a screening test, especially in rural areas at high risk, which is not able to show SAT [22]. Whenever possible, a serum brings positive results, so that is confirmed by a more specific test. RBPT [23]also plays a big role in the fast confirmation of neurobrucellose, arthritis, curingitis and hydrocele from Brucella. The test is an excellent screening test, but can be overdown to diagnose in each animal, in particular vaccine rates [24].

Serum agglutination test (SAT) remains the most popular and globally used diagnostic tool. SAT measures the total amount of agglutinating antibodies (IgM and IgG) [25] and is determined by the amount of specific IgG 2mercaptoethanol (2ME). IgG antibody is considered a better indicator of active infections, and the type of antibody is important because a sharp decrease in IgG antibody levels is a prognosis for successful treatment [26]. This test is easy, cheap to run, but lacking in sensation.

The complement binding reaction (CFT) is a complex method that, although sensitive and specific, requires excellent laboratory facilities and skilled staff. If these are available and tests are run regularly with sufficient attention to quality assurance, it can be very satisfactory reaction test [27].

(B) ELISA Test

The ELISA test provides excellent sensitivity and specificity while being robust and very simple to run the with minimal equipment and is readily available from many commercial sources in kit format. Compared to SAT, ELISA [28]offers high sensitivity and specificity. It is reported that the ELISA is the most sensitive test for the diagnosis of neurological brucellosis [29]. The omp28 protein is currently used in an indirect plate ELISA system and has been evaluated for its excellent sensitivity and specificity in a number of clinical samples.

Fluorescence polarization analysis (FPA) requires special reagents and readers and is known to have advantages over other methods in sensitivity and specificity [30]. However, evaluation is limited to and the procedure is not widely available. More information is needed before evaluating the overall value. This technique is readily applicable to the detection of either the antibody or antigen through simultaneous changes in rotational speed [31]. Diagnosis of Brucella disease by test using S phase, Rose Bengal plate test (RBT) serum aggregation test (SAT), comb antiglobulin, complement binding test (CFT), ELISA. [32]. The results of combining tests such as the SAT or comb antiglobulin can be used to evaluate the steps in disease progression at diagnosis. ELISA with the appropriate IgM or IgG specificity of and a conjugate of SLPS (lipopolysaccharide) can replace the established test, but new standardization and validation are required. Other methods are useful, but have low specificity and are not sufficiently evaluated [33].

Protein Diagnostics Test

Antigens of proteins that are important to gin can be classified into four groups: LPS, external membrane proteins, peripheral cytoplasmic proteins, and cellular proteins. [34].

- i) LPS: Smoothness Attention LPS consists of a core area containing lipidA and mannose, Quinova Social Democrats, 3DeoxyDmannose2octulosonate (KDO) and the offside chain [35]
- ii) Outer membrane protein: Brucella spp. Major outer membrane proteins (OMP). was first identified in the early 1980s and was characterized by potential immunogenicity and protective antigens. OMPs are classified into Group I (88-94 kDa), Group II (35-39 kDa) and Group III (25-31 kDa) proteins based on the mass of the molecule. These proteins feature additional monoclonal antibodies, a) lipoproteins (Omp10, 16, 19), b) two homologous groups (Omp25, Op31), c) folliproteins (Omp2a / 2b), d). Contains Group 1 protein (Omp1) [18].iii)Periprasm Protein: Many immunogenic proteins have been identified in Peripurazumu space, such as BP26 and Cu/Zn SOD. iv)Cytoplasmic Proteins: Proteins in this category include heat shock proteins DnaK, HrtA, GroEL, GroES and ribosomal L7/L12 proteins [36].

Prevention

To prevent human brucellosis alawys avoid consuming milk derivatives and unpasteurized milk. **[37]**.Barrier precautions for endangered hunters and professionals (butchers, farmers, slaughterers, veterinarians) **[38]**. Carefully handle postpartum and disposal, especially in 4,444 abortion cases. Serological or other tests of animals; vaccination of unreasonable / herd may be expected. Remove infected herds / herds. Occupational and food hygiene; Vaccine is generally not recommended **[39]**.

Treatment

Brucella is a pathogenic bacteria mainly four species which are more pathogenic for human body like as *Brucella melitensis*, *Brucella suis*, *Brucella abortus*, *Brucella Canis* and this species are highly infectious by aerosoles [40]. Human to human transmission does not usually occur. The incubation time/period varies from 5 days to 6 months [41]. According to durstion of infection symtoms includes fever, joint pain, cough etc. There are variouse combination treatments with Doxycline along with Streptomycin and Refampicin or any amino glycosides act as fisrst line therapy for the treatment of Brucellosis for first six weeks [42]. Anothers Combination therapy is Tetracycline and Streptomycin also use for the treatment of Brucellosis [43].

	-	Post Exposure Prophylaxis in case of
	Confirmed Clinical Case	suspected or confirmed exposured to
	Duration Treatment 6 Weeks	the pathogen
		Duration Treatment 3 to 6 Weeks
Posology	For Adults	For Adults
	100mg iv twice daily	100mg orally twice daily
	100mg orally twice daily	
	For Children	<u>For Children</u>
	>8years and >45kg Adult dose	>8years and >45kg Adult dose orally
	>8 years and<45kg 2.2mg/kg iv twice	>8 years and<45kg 2.2 mg/kg orally
	daily and same for oral dose	twice daily
Contraindication	Should be considered in view of the prescribing information given in the different	
Pregnancy and lactation	Given only the seriousness condition	
	Contraindication	Posology For Adults 100mg iv twice daily 100mg orally twice daily 100mg orally twice daily For Children >8years and >45kg Adult dose >8 years and<45kg 2.2mg/kg iv twice daily and same for oral dose

Table 2 : Combination Medication Treatment and Their Dose . [44-49]

Name of Active Substance	Section	Treatment of Suspected or	Post Exposure Prophylaxis in case of
Role in Therapy and		Confirmed Clinical Case	suspected or confirmed exposured to
Prophylaxis		Duration Treatment 6 Weeks	the pathogen
			Duration Treatment 3 to 6 Weeks
Refampicin	Posology	For Adults	For Adults
use to first line treatment		10-15mg/kg iv daily	600-900mg orally daily
combination with doxycyline		600-900mg orally daily	
adults and children, children			
age above 8 years old.		For Children	For Children
		10-15mg/kg iv daily	10-15mg/kgorally daily
		10-15mg/kgorally daily	
First line prophylaxis in			
combination with	Contraindication	Should be considered in view of the prescribing information given in the different	
doxycycline in adults and	Pregnancy and lactation	Given only the seriousness condition	
child more than 8 years of			
age			

There are another alternative theray also present like combination therapy of sulfamethoxazole -trimethoprim and combination with ofloxacine or ciprofloxacine also used as a first line therapy for the treatment of brucellosis **[50]**.

Conclusion

Brucellosis has been eradicated in many developed countries, but remains an important public health problem in most developing countries due to the serious economic losses associated with abortion and unreasonable infertility. Human brucellosis is commonly reported among laboratory workers, slaughterhouse employees, farmers and veterinarians who can be exposed to infected animals.For its non-uniformity and lack of specificity, the diagnosis of Brucellosis has always required laboratory conformation, either by isolation of pathogen or demonstration of specific antibodies. The serological test available for the diagnosis of Brucellosis continues to be the most useful test for preliminary identification of the disease, as well as the limitation of for its low sensitivity.Therefore, it is necessary to develop a fast, reliable and user-friendly system for the diagnosis ofdiseases and alternative vaccine access. Because of the inherent challenges of in isolation, inefficiency, cost, risk, and other factors, most laboratories prefer to use other, more cost-effective methods. As a diagnostic tool, molecular biology is advancing and is soon to replace actual bacterial isolation. It's fast, safe and cost-effective, and the only real problem is some uncertainty about specificity.

References

1. Ko J, Splitter GA (2003) Molecular host-pathogen interaction in brucellosis: current understanding and future approaches to vaccine development for mice and humans. Clin Microbiol Rev 16:65–78

2. Corbel MJ, Elberg SS, Cosivi O, Brucellosis in humans and animals. World Health Organization, Geneva , 2020. 118-77

3.Ross H, Foster G, Reid R, Jahans K, MacMillan A. Brucella species infection in sea-mammals. Vet Rec. 1994; 134(14): 359.

4.Sankarasubramanian J, Vishnu US, Gunasekaran P, Rajendhran J. Identification of genetic variants of Brucella spp. through genomewide association studies. In: Infection, Genetics and Evolution. Elsevier; 2017. 92–8

5.Gorvel, J. P. and Moreno, E. (2002) Brucella intracellular life: from invasion to intracellular replication. Vet. Microbiol. 90, 281-297.

6. Bargn Kv, Gorvel JP, Salcedo SP. Internal affairs : Investigating the Brucella intracellular lifestyle. FEMS Microbio. Rev. 2012;36:533-562

7. Kim S. The interaction between Brucella and the Host cell in Phagocytosis. Updates of Brucellosis. Intech Open 2015

8. Greenfield RA. Drevets DA. Machado LJ. et al. et al. Bacterial pathogens as biological weapons and agents of bioterrorism. Am J Med Sci. 2002; 323: 299-315

9.Memish Z.Mah MW .Al Mahmoud S .Al Shaalan M .Khan MY.Brucella bacteraemia: clinical and laboratory observations in 160 patients.J Infect. 2000; 40: 59-63

10. Porte F. Naroeni A. Ouahrani-Bettache S. Liautard JP.Role of the *Brucella suis* lipopolysaccharide O antigen in phagosomal genesis and in inhibition of phagosome-lysosome fusion in murinmacrophages. *Infect Immun.* 2003; 71: 1481-1490

11. HEMMEN F, WEYNANTS V, SCARCEZ T, LETESSON JJ, SAMAN E: Cloning and sequence analysis of a newly identified Brucella abortos gene and serological evaluation of the I7-kilodaiton antigen that it encodes. Clin Diagn Lab Immunol 1995; 2 (3): 263-7

12. LEIVA-LEON J, DE-LA-ROSA M, PLATA J, MARTINEZ MJ, JIMENEZ-ALONSO J: An immunoblotting study of serologic response in patients with acute bruceliosis. Diagn Microbiol Infect Dis 1991; 14(6): 515-8.

13. NAVAS E, GUERRERO A, COBO J, LOZA E: Faster isolation of Brucelia spp. from blood by Isolator compared with BACTEC NR. Diagn Microbiol Infect Dis 1993; 16(I): 79-81.

14. ZAITSEVA MB, GOLDING H, BETFS M, YAMAUCHJ A, BLOOM ET, BUTLER LE, STEVAN L, GOLDING B: Human peripherai biood CD4+ and CD8+ T ecOs express Thi-hke cytokine rnRNA and proteins foilowing in vitro stimulation with heat-inactivat cd Brocelia abortos. infecl Immun 1995; 63 (7): 2720-8.

15. OLIVEIRA SC. HARMS IS. BANAI M, SPL1TTER GA: Recornbinant Brocelia abortos proteins that induce prohferation and gamma-mnterferon secretion by CD4+ T ecOs from Brucella-vaccinated mice and deiayed-type hypersensitivity in sensitized guinea pigs. Ccli Immunoi 1996; 172 (2): 262-8.

16. LIAUTARD JP, GROSS A, DORNAND J, KOHLER 5: Interactions between professional phagocytes and Bruceila spp. Microbiologia 1996; 12 (2): 197-206

17. OCON P, REGUERA JM, MORATA P, JUAREZ C, ALONSO A, COLMENERO JD: Phagocytic ccli function in active brucellosis. Infect immun 1994; 62 (3): 9 10-14.

18. . LINDLER LE, HADFIELD TL, TALL BD, SNELLINGS NJ, RUBIN FA, VAN DE VERG LL, HOOVER D, WARREN RL: Cioning of a Bruceila ,nelitensis group 3 antigen gene encoding 0mp28, a pro tem recognized by the humoral immune response during human bru celiosis. Infect Immun 1996; 64(7): 2490-9.

19. JIANG X, BALDWIN CL: Effects of cytokines on intraceliuiar growth of Broceila abortos. infect immun 1993; 61(1): 124-34.

20. OLIVEIRA SC, SPLITT'ER GA.: CD8+ type 1 CD44hi CD45 RB lo T iymphocytes control intraceHuiar Broceila abortos infection as demonstrated in major histocompatibility complex class 1 and class II deficient mice. EurJ Immunol 1995; 25(9): 255 1-7.

21. SPLITTER G, OLIVEIRA 5, CAREY M, MILLER C, KO J, COVERT J: T Iymphocyle mediated protection against facuitative intracellular bacteria. Vet Immunopathol 1996; 54 (1-4): 309-19.

22. BALDWIN CL, WINTER Ai: Macrophages and Bruceila. irnmunoi Ser 1994; 60: 363-80.

23. RODRIGUEZ-ZAPATA M, ALVAREZ-MON M, SALMERON 1, PRIETO A, MANZANO L, SALMERON 01, CARBALL1DO J: Diminished T lymphocyte proiiferative response to polycionai milo gens in acute brucellosis patients. Infection 1996; 24(2): 115-20.

24. Al-Aska AK, Chagla AH. Laboratory-acquired brucellosis. Journal of Hospital Infection. 1989;14(1):69-71. pmid:2570105

25. Connolly JP, Comerci D, Alefantis TG, Walz A, Quan M, Chafin R, Grewal P, Mujer CV, Ugalde RA, DelVecchio VG (2006) Proteomic analysis of *Brucella abortus* cell envelope and identification of immunogenic candidate proteins for vaccine development. Proteomics 6:3767–3780

26. Papasergi S, Garibaldi M, Tuscano G, Signorino G, Ricci S, Peppoloni S, Pernice I, Lo Passo C, Teti G, Felici F, Beninati C (2010) Plasminogenand fibronectin-binding protein B is involved in theadherence of *Streptococcus pneumoniae* to human epithelial cells. J Biol Chem 285:7517–7524

27. Xolalpa W, Vallecillo AJ, Lara M, Mendoza-Hernandez G, Comini M, Spallek R, Singh M, Espitia C (2007) Identification of novel bacterial plasminogen-binding proteins in the human pathogen *Mycobacterium tuberculosis*. Proteomics 7:3332–3341

28. Schwarz-Linek U, Höök M, Potts JR (2006) Fibronectin-binding proteins of gram-positive cocci. Microbes Infect 8:2291-2298

30. Rosas, G., Fragoso, G., Ainciart, N., Esquivel-Guadarrama, F., Santana, A., Bobes, R.J., 2006. Brucella spp. lumazine synthase: a novel adjuvant and antigen delivery system to effectively induce oral immunity. Infect Immun 8, 1277–86.

31. Sciutto, A., Toledo, C., Cruz, G., Rosas, G., Meneses, D., Laplagne, N., 2005. Brucella spp. lumazine synthase: A novel antigen delivery system. Vaccine 23, 2784-2790.

32. Yousefi, S., Tahmoorespur, M., Sekhavati, M.H., 2016. Cloning, expression and molecular analysis of Iranian *Brucella melitensis* Omp25 gene for designing a subunit vaccine. Res Pharm Sci 11, 405-411.

33. Delpino, M.V., Estein, S.M., Fossati, C.A., Baldi, P.C., Cassataro, J., 2007. Vaccination with Brucella recombinant DnaK and SurA proteins induces protection against *Brucella abortus* infection in BALB/c mice. Vaccine 25, 6721-6729.

34. Goldbaum, F.A., Leoni, J., Wallach, J.C., Fossati, C.A., 1993. Characterization of an 18-kilodalton Brucella cytoplasmic protein which appears to be a serological marker of active infection of both human and bovine brucellosis. J Clin Microbiol Infect Dis 31, 2141–2145.

35. Cassataro, J., Estein, S.M., Pasquevich, K.A., Velikovsky, C.A., de la Barrera, S., Bowden, R., Fossati, C.A., Giambartolomei, G.H., 2005. Vaccination with the recombinant Brucella outer membrane protein 31 or a derived 27-amino-acid synthetic peptide elicits a CD4+ Thelper 1 response that protects against *Brucella melitensis* infection. Infect Immun 73, 8079-8088.

38. Cassataro, J., Estein, S.M., Pasquevich, K.A., Velikovsky, C.A., de la Barrera, S., Bowden, R., Fossati, C.A., Giambartolomei, G.H., 2005. Vaccination with the recombinant Brucella outer membrane protein 31 or a derived 27-amino-acid synthetic peptide elicits a CD4+ Thelper 1 response that protects against *Brucella melitensis* infection. Infect Immun 73, 8079-8088.

39. Yaramis A, Kervancioglu M, Yildirim I, Soker M, Derman O, Tas MA. Severe microangiopathic hemolytic anemia and thrombocytopenia in a child with *Brucella* infection. Ann Hematol 80: 546-548, 2001.

40. Colmenero JD, Reguera JM, Martos F, et al. Complications associated with *Brucella melitensis* infection: a study of 530 cases. Medicine (Baltimore) 75: 195-211, 1996.

41. Colmenero JD, Reguera JM, Martos F, et al. Complications associated with *Brucella melitensis* infection: a study of 530 cases. Medicine (Baltimore) 75: 195-211, 1996.

42. King-Brown R, Recht W. Brucella spondylitis; review of the literature and report on two cases. Postgrad Med J. 1952;28:250-4.

44. Bingol A, Yucemen N, Meco O. Medically treated intraspinal "Brucella" granuloma. Surg Neurol. 1999;52:570-6.

45. Hausler WJ Jr, Moyer NP, Holcomb LA: *Brucella*; in Lennette EH, Balows A, Hausler WJ Jr, Shamody HJ (eds): Manual of Clinical Microbiology, ed 4. Washington, American Society for Microbiolgy, 1985, pp 382–386.

46.Gonzalez Sanchez FJ, Encinas Gaspar MB, Napal Lecumberri S, Rajab R: Brucellar orchiepididymitis with abscess. Arch Esp Urol 1997;50:289–292.

47. Castillo Soria JL, Bravo de Rueda Accinelli C: Genital brucellosis: a rare cause of testicular abscess. Arch Esp Urol 1994;47:533-536.

48. Bandara AB, Contreras A, Contreras-Rodríguez A, Martins AM, Dobrean V, Poff-Reichow S, et al. *Brucella suis* urease encoded by *ure1* but not *ure2* is necessary for intestinal infection of BALB/c mice. *BMC Microbiol*. (2007) 7:57. doi: 10.1186/1471-2180-7-57

49. Ibañez AE, Coria LM, Carabajal MV, Delpino MV, Risso GS, González-Cobiello P, et al. A bacterial protease inhibitor protects antigens delivered in oral vaccines from digestion while triggering specific mucosal immune responses. *J Control Release*. (2015) 220:18–28. doi: 10.1016/j.jconrel.2015.10.011

50. Ferrero MC, Fossati CA, Rumbo M, Baldi PC. *Brucella* invasion of human intestinal epithelial cells elicits a weak proinflammatory response but a significant CCL20 secretion. *FEMS Immunol Med Microbiol*. (2012) 66:45–57. doi: 10.1111/j.1574-695X.2012.00985.x